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Table A-1. Schedule for Baseline Monitoring Program activities.

Activity	Deadline (all days are calendar days)
Submission of draft QAPP to EPA	September 17, 2003
Submission of revised QAPP to EPA	21 days after receipt of USEPA comments on draft BMP QAPP (as per AOC)
Commencement of Velocity Profile Study	Spring 2004
Commencement of Year 1 field activities – routine water sampling.	30 days after USEPA approval of the BMP QAPP
Commencement of special studies including velocity profile study, Time of Travel Study, Dissolved/Particulate Study and Lock 1.	May 2004 contingent on river flow conditions
Commencement of Year 1 fish sampling program	May/June 2004 contingent on river flow conditions
Commencement of Year 2 field activities – routine water sampling.	January 2005
Submission of Data Summary Report for Year 1 to EPA	April 1, 2005
Commencement of Year 2 fish sampling program	May/June 2005
Commencement of Year 3 field activities – routine water sampling.	January 2006
Submission of Data Summary Report for Year 2 to EPA	April 1, 2006
Commencement of Year 3 fish sampling program	May 2006
Submission of Data Summary Report for Year 3 to EPA	April 1, 2007

Table A-2. Summary of data quality objectives and associated measurement performance criteria.

Data Quality Objective	Measurement Performance Criteria
<p>Establish baseline PCB load at Thompson Island, Schuylerville, Stillwater and Waterford to be used in determining dredging compliance with resuspension performance standards</p>	<ul style="list-style-type: none"> - Determine the width and depth-integrated TSS and PCB concentrations in water flowing past each of the main stem Upper Hudson River monitoring stations - Determine river flow at the time of sampling
<p>Provide a means to translate between the historical record of PCB concentrations at Thompson Island Dam (TID) and Schuylerville and the proposed baseline data</p>	<ul style="list-style-type: none"> - Paired measurements at the historical and baseline monitoring stations using the historical and baseline monitoring sampling protocols
<p>Establish the Baseline Annual PCB Load at Waterford and on the Mohawk River at Cohoes to provide a basis to assess the effectiveness of the remedy in reducing PCB Load to the Lower Hudson River</p>	<ul style="list-style-type: none"> - Weekly sampling will be conducted to provide sufficient data to capture seasonal trends - Monthly sampling at the Mohawk River is sufficient because of absence of significant sediment PCB source - Year-round sampling will be conducted to allow for an annual PCB loading estimate
<p>Establish baseline PCB concentrations upstream of the GE facilities to determine background PCB levels</p>	<ul style="list-style-type: none"> - A single sampling location will be used to provide an accurate representation of the PCB concentration and loading at the Bakers Falls Bridge sampling station - Low-level PCB collection, extraction and analysis methods will be used to quantify PCBs at the Bakers Falls Bridge sampling station
<p>Establish baseline PCB concentrations at Rogers Island to determine the PCB contribution downstream of the background station and upstream of the sediment remedial action</p>	<ul style="list-style-type: none"> - Centroid (approximate center channel) sampling will be performed in the East and West channels of river at Rogers Island - Sampling will occur year-round - Low-level PCB collection, extraction and analysis methods will be used to quantify PCBs at Rogers Island

Table A-2. Summary of data quality objectives and associated measurement performance criteria.

Data Quality Objective	Measurement Performance Criteria
<p>Establish reference concentrations of nutrients, metals and dioxins/furans prior to dredging</p>	<ul style="list-style-type: none"> - TAL metals will be monitored bi-weekly from May through November of each year at each of the Upper Hudson River Stations - Nutrients (TKN, nitrate, nitrite, total phosphorous) will be monitored weekly from May through November of 2004 at each of the Upper Hudson River Stations - Dioxins and furans will be monitored monthly from May through November of 2004 at Rogers Island, Thompson Island, Schuylerville, Stillwater, and Waterford - Sampling protocols are the same as for PCBs
<p>Establish a relationship between turbidity and meteorological events</p>	<ul style="list-style-type: none"> - Turbidity will be measured weekly at each of the main stem Upper Hudson River stations - Precipitation is measured at numerous stations within the tributary basins
<p>Establish baseline conditions of parameters potentially useful for comparison to conditions during the dredging operation</p>	<ul style="list-style-type: none"> - TSS, DOC and POC will be measured on water samples in conjunction with the PCB samples - Water quality (WQ) measurements including pH, dissolved oxygen (DO), conductivity, turbidity and temperature will be taken at each sampling station using a probe
<p>Confirm particulate and dissolved phase PCB partitioning behavior under baseline conditions to provide a means to evaluate the cause of elevated PCB levels that may potentially be observed during remedial action</p>	<ul style="list-style-type: none"> - High volume samples will be collected at Thompson Island and Schuylerville and filtered in the field once per month (May-November) during the 2004 field season - The aqueous and particulate phases will be analyzed separately

Table A-2. Summary of data quality objectives and associated measurement performance criteria.

Data Quality Objective	Measurement Performance Criteria
<p>Establish baseline PCB concentrations in the Lower Hudson River to assess remedy effectiveness and provide a baseline in the vicinity of the principal Lower Hudson River water intake</p>	<ul style="list-style-type: none"> - Depth integrated samples taken at the centroid (approximate center channel) of the river will be collected at Albany/Troy and Poughkeepsie on a monthly basis from May through November
<p>Monitor PCB levels in Upper Hudson River sport fish and forage fish to allow evaluation of long term recovery trends.</p>	<ul style="list-style-type: none"> - Fish species (including sport and forage fish) covering a range of association with sediment including black bass, yellow/brown bullhead, yearling pumpkinseed, yellow perch and spottail shiner or substitute forage fish will be collected from multiple locations within each river section - A maximum of 20-30 fish samples of each species (depending on location) will be collected - The Feeder Dam Pool will serve as a reference location - Fish will be collected at Albany/Troy just below the Federal Dam to determine when PCB concentrations in Lower Hudson River fish have reached levels that would permit relaxation of consumption advisories

Table A-3. New York State's ELAP and the National Environmental Accreditation Program (NELAP) Certifications Maintained by the Analytical Laboratories

Matrix	Category	Laboratory	Analyte Name	CAS number	Analytical Method	Certification
Water	PCBs	NEA	Congeners and Total PCBs (sum of congeners)	1336-36-3	Modified Green Bay Mass Balance Method (NEA 207_03)	No Certification Available
	Select Nutrients	Bender- St. Peter's	Nitrate	NT001	EPA 353.3	NYSDOH Non-Potable Water - EPA 353.3
			Nitrite	NT002	EPA 354.1	NYSDOH Non-Potable Water - EPA 354.1
			TKN	NT003	EPA 351.3	NYSDOH Non-Potable Water - EPA 351.3
			Total Phosphorus	NT004	EPA 365.2	NYSDOH Non-Potable Water - EPA 365.2
	TAL Metals	STL Pittsburgh	Ag (Silver)	7440-22-4	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Al (Aluminum)	7429-90-5	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			As (Arsenic)	7440-38-2	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Ba (Barium)	7440-39-3	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Be (Beryllium)	7440-41-7	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Ca (Calcium)	7440-70-2	EPA 200.8	No Certification Available
			Cd (Cadmium)	7440-43-9	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Co (Cobalt)	7440-48-4	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Cr (Chromium)	7440-47-3	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Cu (Copper)	7440-50-8	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Fe (Iron)	7439-89-6	EPA 200.8	No Certification Available
			Hg (Mercury)	7439-97-6	EPA 245.1	NYSDOH Non-Potable Water - EPA 245.1
			K (Potassium)	7440-09-7	EPA 200.8	No Certification Available
			Mg (Magnesium)	7439-95-4	EPA 200.8	No Certification Available
			Mn (Manganese)	7439-96-5	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Na (Sodium)	7440-23-5	EPA 200.8	No Certification Available
			Ni (Nickel)	7440-02-0	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Pb (Lead)	7439-92-1	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Sb (Antimony)	7440-36-0	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
			Se (Selenium)	7782-49-2	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8
	Tl (Thallium)	7440-28-0	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8		
	V (Vanadium)	7440-62-2	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8		
Zn (Zinc)	7440-66-6	EPA 200.8	NYSDOH Non-Potable Water - EPA 200.8			
Other	NEA	TSS	WQ001	EPA 160.2	NYSDOH Non-Potable Water - EPA 160.2	
		DOC and POC	OC001/ OC002	NE128_03	NYSDOH Non-Potable Water - TOC by EPA 415.1 (No certification available for POC/DOC or "Tekmar Dohrmann")	

Table A-3. New York State's ELAP and the National Environmental Accreditation Program (NELAP) Certifications Maintained by the Analytical Laboratories

Matrix	Category	Laboratory	Analyte Name	CAS number	Analytical Method	Certification
Water (Cont'd)	Dioxins and Furans	Paradigm	Total-TCDD	41903-57-5	USEPA 1613	No Certification Available
			2378-TCDD	1746-01-6	USEPA 1613	NELAC NY Lab Non-Potable Water by EPA 1613
			Total-TCDF	55722-27-5	USEPA 1613	No Certification Available
			2378-TCDF	51207-31-9	USEPA 1613	No Certification Available
			Total PeCDD	36088-22-9	USEPA 1613	No Certification Available
			12378-PeCDD	40321-76-4	USEPA 1613	No Certification Available
			Total PeCDF	30402-15-4	USEPA 1613	No Certification Available
			12378-PeCDF	57117-41-6	USEPA 1613	No Certification Available
			23478-PeCDF	57117-31-4	USEPA 1613	No Certification Available
			Total-HxCDD	34465-46-8	USEPA 1613	No Certification Available
			123478-HxCDD	39227-28-6	USEPA 1613	No Certification Available
			123678-HxCDD	57653-85-7	USEPA 1613	No Certification Available
			123789-HxCDD	19408-74-3	USEPA 1613	No Certification Available
			Total HxCDF	55684-94-1	USEPA 1613	No Certification Available
			123478-HxCDF	70648-26-9	USEPA 1613	No Certification Available
			123678-HxCDF	57117-44-9	USEPA 1613	No Certification Available
			123789-HxCDF	72918-21-9	USEPA 1613	No Certification Available
			234678-HxCDF	60851-34-5	USEPA 1613	No Certification Available
			Total HpCDD	37871-00-4	USEPA 1613	No Certification Available
			1234678-HpCDD	35822-46-9	USEPA 1613	No Certification Available
			Total HpCDF	38998-75-3	USEPA 1613	No Certification Available
1234678-HpCDF	67562-39-4	USEPA 1613	No Certification Available			
1234789-HpCDF	55673-89-7	USEPA 1613	No Certification Available			
OCDD	3268-87-9	USEPA 1613	No Certification Available			
OCDF	39001-02-0	USEPA 1613	No Certification Available			
Fish	PCBs	NEA	Congeners and Total PCBs (sum of congeners)	1336-36-3	Green Bay Mass Balance Method (NEA 013_07)	No Certification Available
			Aroclor 1016	12674-11-2	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1221	11104-28-2	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1232	11141-16-5	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1242	53469-21-9	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1248	12672-29-6	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1254	11097-69-1	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Aroclor 1260	11096-82-5	SW846 8082 (NE148_04)	NYSDOH Solid and Hazardous Waste - SW846 8082
			Total PCBs (sum of Aroclors)	1336-36-3	SW846 8082 (NE148_04)	No Certification Available
			Other	NEA	Percent Lipid	LP001
Mercury	7439-97-6	SW-846 7471A	NYSDOH Solid and Hazardous Waste - SW846 7471A			

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Matrix	Category	Laboratory	Analyte Name	CAS number	Analytical Method	Certification
Fish (Cont'd)	Dioxins and Furans	Paradigm	Total-TCDD	41903-57-5	USEPA 1613	No Certification Available
			2378-TCDD	1746-01-6	USEPA 1613	No Certification Available
			Total-TCDF	55722-27-5	USEPA 1613	No Certification Available
			2378-TCDF	51207-31-9	USEPA 1613	No Certification Available
			Total PeCDD	36088-22-9	USEPA 1613	No Certification Available
			12378-PeCDD	40321-76-4	USEPA 1613	No Certification Available
			Total PeCDF	30402-15-4	USEPA 1613	No Certification Available
			12378-PeCDF	57117-41-6	USEPA 1613	No Certification Available
			23478-PeCDF	57117-31-4	USEPA 1613	No Certification Available
			Total-HxCDD	34465-46-8	USEPA 1613	No Certification Available
			123478-HxCDD	39227-28-6	USEPA 1613	No Certification Available
			123678-HxCDD	57653-85-7	USEPA 1613	No Certification Available
			123789-HxCDD	19408-74-3	USEPA 1613	No Certification Available
			Total HxCDF	55684-94-1	USEPA 1613	No Certification Available
			123478-HxCDF	70648-26-9	USEPA 1613	No Certification Available
			123678-HxCDF	57117-44-9	USEPA 1613	No Certification Available
			123789-HxCDF	72918-21-9	USEPA 1613	No Certification Available
			234678-HxCDF	60851-34-5	USEPA 1613	No Certification Available
			Total HpCDD	37871-00-4	USEPA 1613	No Certification Available
			1234678-HpCDD	35822-46-9	USEPA 1613	No Certification Available
			Total HpCDF	38998-75-3	USEPA 1613	No Certification Available
			1234678-HpCDF	67562-39-4	USEPA 1613	No Certification Available
			1234789-HpCDF	55673-89-7	USEPA 1613	No Certification Available
OCDD	3268-87-9	USEPA 1613	No Certification Available			
OCDF	39001-02-0	USEPA 1613	No Certification Available			

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Matrix	Category	Laboratory	Analyte Name	CAS number	Analytical Method	Certification
Fish (Cont'd)	Organochlorine Pesticides	NEA	Aldrin	309-00-2	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			α-BHC	319-84-6	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			β-BHC	319-85-7	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			γ-BHC (Lindane)	58-89-9	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			δ-BHC	319-86-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			α-Chlordane	5103-71-9	SW846 8081A (NE131_03)	No Certification Available
			γ-Chlordane	5103-74-2	SW846 8081A (NE131_03)	No Certification Available
			Chlordane	57-74-9	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			4,4'-DDD	72-54-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			4,4'-DDE	72-55-9	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			4,4'-DDT	50-29-3	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Dieldrin	60-57-1	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endosulfan I	959-98-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endosulfan II	33213-65-9	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endosulfan sulfate	1031-07-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endrin	72-20-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endrin aldehyde	7421-93-4	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Endrin ketone	53494-70-5	SW846 8081A (NE131_03)	No Certification Available
			Heptachlor	76-44-8	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
			Heptachlor epoxide	1024-57-3	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A
Methoxychlor	72-43-5	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A			
Toxaphene	8001-35-2	SW846 8081A (NE131_03)	NYSDOH Solid and Hazardous Waste - SW846 8081A			

Table B-1. Description of routine water column sampling locations, procedures, and significance.

Sampling Location ¹	Approx. HRM ²	Description	Approx. Water Depth ³	Sampling Method	Significance ⁴
Bakers Falls Bridge	196.9	Sample collected from the centroid (defined as approximate center of the channel) from the downstream side of the County Route 27 Bridge in Hudson Falls. Approximate distance from top of guardrail to river bed ~ 38 ft.	8 ft.	Single depth-integrated sample collected with multiple aliquot depth integrating sampler.	Historical Monitoring Station (PCRDMP). Measures background PCB concentrations in the Hudson River upstream of GE facilities, remnant deposits, and PCB-containing sediment.
Rogers Island - Route 197 Bridge	194.2	A centroid sample will be collected from the east and west channels of the Hudson River. The flow distribution will be used to weight the volumes of each sample in the composite. The east and west channels are sampled from the upstream side of the Route 197 Bridge in Fort Edward. Distance from concrete deck to river bed ~ 29 ft on the west side and ~ 34 ft on the east side.	8 ft. (West) 8 ft. (East)	Depth-integrated water samples collected at the centroid (defined as approximate center channel to be consistent with historical sampling stations) by lowering a multiple aliquot depth integrating sampler through the water column to approximately 75% of the location's depth. The entire volume collected during each deployment will be poured into composite sample containers.	Historical Monitoring Station (PCRDMP). Measures PCB concentrations in the Hudson River downstream of GE facilities - approximately 0.7 miles below the former Fort Edward Dam, the upstream end of River Section 1. Studies performed by O'Brien & Gere Engineers in 1995 ⁵ indicate that sampling from this location should provide representative data for estimating the PCB load past this station. Under mean flow conditions, approximately 65% of the river flow is in the west channel and 35% is in the east channel; however, the proportion of water flowing through each channel varies with flow rate.
Thompson Island	187.5	Transect sampling to occur at 6 equal-flow locations over the cross section at this station. Samples will be collected at the southern end of Thompson Island by boat. The historical single-point sampling locations at TID PRW2 will be sampled simultaneously with the transect sampling monthly from March to November during 2004.	11 ft.	Depth-integrated water samples collected at each EDI location by lowering a multiple aliquot depth integrating sampler through the water column to approximately 75% of the location's depth. The entire volume collected during each deployment will be poured into composite	Historical Monitoring Location. Located just downstream of Thompson Island Dam, the beginning of River Section 2. HRMP Station TID-PRW2, which was often inaccessible by boat during winter or high flow events, and TID-West, a shore-based station that has been shown to be biased ^{6,7} . Samples from the center of the channel below the dam have been shown to be representative of the PCB load entering River Section 2 ⁷ .

Table B-1. Description of routine water column sampling locations, procedures, and significance.

Sampling Location ¹	Approx. HRM ²	Description	Approx. Water Depth ³	Sampling Method	Significance ⁴
Schuylerville – Route 29 Bridge	181.4	<p>Transect sampling to occur at 6 equal-flow locations over the cross section at this station. Samples are collected from the upstream-side of the Route 29 Bridge in Schuylerville. Distance from the top of the guardrail to the Riverbed ~ 53 ft.</p> <p>The historical single-point sampling locations at Schuylerville will be sampled simultaneously with the transect sampling monthly (year-round) during 2004.</p>	21 ft.	<p>sample containers.</p> <p>Depth-integrated water samples collected at each EDI location by lowering a USGS “fish sampler” through the water column to approximately 75% of the location’s depth. The entire volume collected during each deployment will be poured into composite sample containers.</p>	<p>Historical Monitoring Station (HRMP). Samples collected from this location are assumed to be representative of PCB loading past this station, which is approximately 2 miles downstream of the Northumberland Dam, which divides River Sections 2 and 3.</p>
Stillwater – Route 125 Bridge	168.4	<p>Transect sampling to occur at 5 equal-flow locations over the cross section at this station. Samples are collected from the downstream side of the Route 125 Bridge in Stillwater. Distance from the top of the guardrail to the Riverbed (to be determined; TBD).</p>	5 ft.	<p>Depth-integrated water samples collected at each EDI location by lowering a multiple aliquot depth integrating sampler through the water column to approximately 75% of the location’s depth. The entire volume collected during each deployment will be poured into composite sample containers.</p>	<p>Samples collected at this station are assumed to be representative of the PCB load transported downstream from the upper reach (Northumberland Dam to Stillwater Dam) of River Section 3.</p>

Table B-1. Description of routine water column sampling locations, procedures, and significance.

Sampling Location ¹	Approx. HRM ²	Description	Approx. Water Depth ³	Sampling Method	Significance ⁴
Lock 1 – Route 4 Bridge	159.5	Transect sampling will occur at 5 equal-flow locations over the cross section at this station by boat.	(TBD)	Depth-integrated water samples collected at each EDI location by lowering a multiple aliquot depth integrating sampler through the water column to approximately 75% of the location’s depth. The entire volume collected during each deployment will be poured into composite sample containers.	New station, not historically sampled. Sampling may be discontinued after 2004 if good correlation with data collected from Waterford station is observed.
Waterford – Route 4 Bridge	156	Transect sampling occurs at 5 equal-flow locations over the cross section at this station. Samples are collected from the downstream side of the Rt. 4 Bridge in Waterford. Samples will be taken at the centroid (defined as approximate center of the channel to be consistent with historical sampling location) during high flow.	12 ft.	Depth-integrated water samples collected at each EDI location by lowering a multiple aliquot depth integrating sampler through the water column to approximately 75% of the location’s depth. The entire volume collected during each deployment will be poured into composite sample containers.	Historical USGS monitoring station for TSS. Samples collected at this station are assumed to be representative of the PCB load transported downstream from the lower reach (Waterford Dam at Lock 1 to Troy Dam) of River Section 3 to the Lower Hudson River below Troy. .Samples collected from this station will generate data to compute annual baseline loads to assess the effectiveness of the remedy in reducing the PCB load from the Upper Hudson River to the Lower Hudson River.
Mohawk River at Cohoes	NA	Transect sampling occurs at 5 equal-flow locations over the cross section at this station. Samples are collected from the downstream side of the Rt. 32 Bridge in Cohoes.	14 ft.	Single, depth-integrated sample collected multiple aliquot depth integrating sampler.	Samples collected from this station will generate data to compute annual baseline loads to assess the effectiveness of the remedy in reducing the PCB load from the Upper Hudson River to the Lower Hudson River.

Table B-1. Description of routine water column sampling locations, procedures, and significance.

Sampling Location ¹	Approx. HRM ²	Description	Approx. Water Depth ³	Sampling Method	Significance ⁴
Albany/Troy	145	Centroid (defined as approximate center of the channel).	25 ft.	Single, depth-integrated sample collected with multiple aliquot depth integrating sampler.	Samples collected from this station will provide a baseline to assess the remedy effectiveness.
Poughkeepsie	75	Centroid (defined as approximate center of the channel).	75 ft.	Single, depth-integrated sample collected with multiple aliquot depth integrating sampler.	Samples collected from this station will provide a baseline in the vicinity of the principal Lower Hudson water intake.

¹ Designations presented correspond to those used in the GE Hudson River Database.

² HRM refers to Hudson River Mile. HRM 0.0 is located at the Battery in New York City.

³ Approximate water depth at center of channel during typical mean flow of 5,000 cfs.

⁴ PCRDMP = Post-Construction Remnant Deposit Monitoring Program; HRMP = GE Hudson River Monitoring Program.

⁵ O'Brien & Gere, 1996. Hudson River Project, River Monitoring Test. O'Brien & Gere Engineers, Inc., Syracuse, New York January, 1996.

⁶ QEA, 1998. Thompson Island Pool Sediment PCB Sources. Quantitative Environmental Analysis, LLC. March 1998.

⁷ O'Brien & Gere, 1998. Hudson River Project, 1996-1997 Thompson Island Pool Studies. O'Brien & Gere Engineers, Inc., Syracuse, New York. February, 1998.

Table B-2. Hudson River water monitoring summary.

Station	Hudson RM	Sample Type ¹	Analyte and Sampling Frequency				
			PCBs, TSS, Suspended OC, Dissolved OC	Additional TSS	TAL Metals	Nutrients ⁶	Dioxins/Furans ⁶
Bakers Falls ²	197.0	Centroid (~center channel)	Year-round/weekly		May-Nov./bi-weekly	May-Nov./weekly	
Rogers Island	194.2	Centroid (~center of East and West channels)	Year-round/weekly		May-Nov./bi-weekly	May-Nov./weekly	May-Nov./monthly
Thompson Island	187.5	Transect (6 loc.)	March-Nov./weekly	Weekly (May-June)	May-Nov./bi-weekly	May-Nov./weekly	May-Nov./monthly
Schuylerville ³	181.4	Transect (6 loc.)	Year-round/weekly	Weekly (May-June)	May-Nov./bi-weekly	May-Nov./weekly	May-Nov./monthly
Stillwater	168.4	Transect (5 loc.)	May-Nov./weekly		May-Nov./bi-weekly	May-Nov./weekly	May-Nov./monthly
Lock 1 ⁴	159.5	Transect (5 loc.)	May-Nov./weekly		May-Nov./bi-weekly	May-Nov./weekly	
Waterford	156	Transect (5 loc.)	Year-round/weekly		May-Nov./bi-weekly	May-Nov./weekly	May-Nov./monthly
		Centroid (~center channel)	during high flow				
Mohawk River at Cohoes	NA	Transect (5 loc.)	Year-round/monthly				
Albany/ Troy ⁵	145	Centroid (~center channel)	May-Nov./monthly				
Poughkeepsie ⁵	75	Centroid (~center channel)	May-Nov./monthly				

Note: Water Quality (WQ) measurements including temperature, specific conductivity, pH, turbidity and dissolved oxygen will be taken for each water sample using a probe.

¹ A single composite sample will be generated for each station

² Bakers Falls sampling will be reduced to monthly after the first year if concentrations are uniformly low.

³ The historical single-point sampling locations at TID (TID-PRW2) and Schuylerville will be sampled simultaneously with the transect sampling monthly for the first 12 months of the program.

⁴ If data for Lock 1 for 2004 field season exhibit a strong correlation with the Waterford station, the Lock 1 station will be abandoned.

⁵ Only PCBs and TSS will be measured at the Lower Hudson Stations.

⁶ Sampling for nutrients and dioxins/furans to occur in 2004 only.

Special Studies

- A velocity profile study will be conducted at each routine monitoring station during the first few months of the program to refine the equal discharge interval locations.
- A transect positioned upstream of Lock 1 will be sampled for 7 months (May-Nov.) during the 2004 field season. If the data exhibit a strong correlation with data from the Waterford station, the Lock 1 station will be abandoned.
- Pseudo-TOT sampling will take place monthly at the routine monitoring stations in the Upper Hudson River for 7 months (May-Nov) during 2004.
- A dissolved/particulate phase PCB study will be conducted at Thompson Island and Schuylerville once per month (May to Nov) during 2004.

Table B-3. Description of annual fish sampling locations, procedures, and significance.

Sampling Location	Approx. HRM	Description	Sampling Method	Significance
Feeder Dam Pool	201.1	Above Feeder Dam	Electrofishing/netting/angling	Historical Monitoring Station. Measures background PCB concentrations in fish upstream of the GE facilities, remnant deposits, and PCB-containing sediment.
Thompson Island Pool	188.5 – 195	Griffin Island is the historical sampling station (east channel; forage, west channel; adult); approximately 10 fish samples per species will be collected from this station while additional samples will be collected on a reach average basis, targeting approximately 5 fish samples per species at sub-locations that are approximately evenly distributed (depending on habitat availability) within the pool.	Electrofishing/netting/angling	Historical Monitoring Station. Samples will be collected from the historical sampling station as well as from available habitats in the entire pool to determine whether PCB concentrations measured in fish collected from the historical sampling station provides an accurate representation of the reach-wide average.
Ft. Miller/Northumberland Pools	183.4 – 188.5	Samples will be collected on a reach average basis targeting approximately 5 fish samples per species per river mile.	Electrofishing/netting/angling	Samples will be collected from available habitats within these pools to establish a baseline for comparison to construction and post-construction conditions.
Stillwater Pool	168.2 – 183.2	Historical sampling stations include the east side of the Hudson River above Lock C4 (forage fish) and Coveville (adult fish); approximately 10 fish samples per species will be collected from these stations while additional samples will be collected on a reach average basis, targeting approximately 5 fish samples per species at sub-locations that are approximately evenly distributed (depending on habitat availability) within the pool.	Electrofishing/netting/angling	Historical Monitoring Station. Samples will be collected from the historical sampling station as well as from available habitats in the entire pool to determine whether PCB concentrations measured in fish collected from the historical sampling station provides an accurate representation of the reach-wide average.
Albany/Troy	153.2 and 142	Below Federal Dam between Troy and Green Island (adult fish) and South Turning Basin (forage fish)	Electrofishing/netting/angling	Historical Monitoring Station. Provides data at the boundary of Upper Hudson and Lower Hudson River, where the impact of the Upper Hudson River is the greatest.

Table B-4. Hudson River fish monitoring summary.

Sampling Location	Hudson River Miles	Sampling Approach	Species/Station (Spring/Fall) ²	Samples per Species ³	Analytes ⁴
Feeder Dam Pool	201.1	1 location	3/2	20	Total PCBs Lipid content Mercury Dioxins/furans Organochlorine pesticides
Thompson Island Pool	188.5 - 195	Reach Average ¹	3/2	30	
Northumberland/Fort Miller Pools	183.4 - 188.5	Reach Average ¹	3/2	25	
Stillwater pool	168.2 - 183.2	Reach Average ¹	3/2	30	
Albany/Troy	153.2 and 142	1 location	3/2	20	

- ¹ Approximately 5 fish samples per species will be targeted at sub-locations approximately evenly distributed (depending on habitat availability) within the pool, with a maximum of 25 of each species at Northumberland/Ft. Miller, and a maximum of 30 of each species at Thompson Island and Stillwater Pools.
- ² Spring collections include black bass (smallmouth or largemouth), bullhead (yellow or brown), and yellow perch. Fall collections include pumpkinseed and spottail shiner (or substitute forage species if spottail shiner are not available). One exception is the Albany Troy station where both yellow and white perch will be collected (10 samples each).
- ³ Sample numbers are for bass, bullhead, perch, and pumpkinseed individual fish. For forage fish, 10 composite samples will be collected from each reach or location.
- ⁴ Total PCBs and lipid contents will be measured in all fish. Total PCBs will be measured as Aroclor totals. On 10% of the annual fish sample collections, the Green Bay Congener method will be performed. Mercury, dioxins/furans and organochlorine pesticides will be analyzed one time during the program on 10% of the total number of adult fish sample annual collections.

Table B-5. Required Containers, Preservatives, and Holding Times

Matrix	Parameter	Container Specifications	Preservation	Holding Times¹
Water	PCBs (congener specific)	1 L or 4 L amber glass bottle w/Teflon® lined cap	Cool, 4°±2°C	7 days to extraction, 40 days to analysis
	Total Phosphorus, Total Kjeldahl Nitrogen	1L HDP bottle	Cool, 4°±2°C H ₂ SO ₄ to pH<2	28 days
	Nitrate-Nitrogen, Nitrite Nitrogen	1L HDP bottle	Cool, 4°±2°C	48 hours
	Total TAL Metals	500 ml HDP bottle (no liner)	HNO ₃ to pH< 2	28 days for mercury and 6 months for all other metals
	Dissolved TAL Metals	500 ml HDP bottle (no liner)	Field Filter then HNO ₃ to pH< 2	28 days for mercury and 6 months for all other metals
	Total Suspended Solids	1 L HDP bottle	Cool, 4°±2°C	7 days to analysis
	Dissolved and Particulate Organic Carbon	250 ml glass bottle	Cool, 4°±2°C No preservative	14 days
	Dioxins/Furans	1 L amber glass bottle w/Teflon® lined cap	Cool, 4°±2°C	30 days to extraction, 45 days to analysis
Fish	PCBs (sum of Aroclors)	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to extraction, 40 days to analysis
	PCBs (congener specific)	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to extraction, 40 days to analysis
	Percent Lipid	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to extraction, 40 days to analysis
	Mercury	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to analysis
	Dioxins/furans	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to extraction, 40 days to analysis
	Organochlorine pesticides	appropriate-sized wide mouth glass jar w/Teflon® lined cap ^{2,3}	Cool, 4°±2°C until homogenized and sub-sampled and then freeze to <-18°C	1 year to extraction, 40 days to analysis

¹ Holding times are from the date/time of sample collection unless otherwise stated.

² Separate container not required for each parameter.

³ After homogenization of sample.

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits
Water	PCBs (1 liter) DB-1 Peak:	Total PCB (sum of congeners)	1336-36-3	Modified Green Bay Mass Balance Method (NEA 207_03)	ng/L	9.34	32.3
		02		NEA 207_03	ng/L	0.582	2.19
		03		NEA 207_03	ng/L	25.1	1000
		04		NEA 207_03	ng/L	0.318	1.28
		05		NEA 207_03	ng/L	0.363	0.621
		06		NEA 207_03	ng/L	0.0894	0.219
		07		NEA 207_03	ng/L	0.188	0.347
		08		NEA 207_03	ng/L	0.528	2.56
		09		NEA 207_03	ng/L	0.845	25
		10		NEA 207_03	ng/L	0.0660	0.102
		11		NEA 207_03	ng/L	0.247	25
		12		NEA 207_03	ng/L	0.347	25
		13		NEA 207_03	ng/L	0.0617	0.0975
		14		NEA 207_03	ng/L	0.0978	0.676
		15		NEA 207_03	ng/L	0.142	0.676
		16		NEA 207_03	ng/L	0.0762	0.0762
		17		NEA 207_03	ng/L	0.165	0.713
		19		NEA 207_03	ng/L	0.212	25
		20		NEA 207_03	ng/L	0.0190	0.0194
		21		NEA 207_03	ng/L	0.0674	0.132
		22		NEA 207_03	ng/L	0.101	0.101
		23		NEA 207_03	ng/L	0.144	0.753
		24		NEA 207_03	ng/L	0.184	0.964
		25		NEA 207_03	ng/L	0.121	0.726
		26		NEA 207_03	ng/L	0.197	0.530
		27		NEA 207_03	ng/L	0.105	0.163
		28		NEA 207_03	ng/L	0.181	25
		29		NEA 207_03	ng/L	0.117	0.117
		30		NEA 207_03	ng/L	0.481	25
		31		NEA 207_03	ng/L	0.180	0.872
		32		NEA 207_03	ng/L	0.0508	0.420
		33		NEA 207_03	ng/L	0.0671	0.183
		34		NEA 207_03	ng/L	0.0562	0.183
		35		NEA 207_03	ng/L	0.167	25
		36		NEA 207_03	ng/L	0.2	25
		37		NEA 207_03	ng/L	0.122	0.786
		38		NEA 207_03	ng/L	0.126	0.475
		39		NEA 207_03	ng/L	0.148	0.749
		41		NEA 207_03	ng/L	0.209	25
		42		NEA 207_03	ng/L	0.0990	0.172
		43		NEA 207_03	ng/L	0.221	25
		44		NEA 207_03	ng/L	0.0232	0.0402
		45		NEA 207_03	ng/L	0.115	0.115
		46		NEA 207_03	ng/L	0.0478	0.347
		47		NEA 207_03	ng/L	0.108	0.621
		48		NEA 207_03	ng/L	0.267	1.32
		49		NEA 207_03	ng/L	0.0278	0.0932
		50		NEA 207_03	ng/L	0.0865	0.640
		51		NEA 207_03	ng/L	0.209	0.329
		52		NEA 207_03	ng/L	0.0328	0.0366
		53		NEA 207_03	ng/L	0.0548	0.329
		54		NEA 207_03	ng/L	0.0573	0.135
		55		NEA 207_03	ng/L	0.0150	0.0150
		56		NEA 207_03	ng/L	0.0373	0.0548
		57		NEA 207_03	ng/L	0.0426	0.102
		58		NEA 207_03	ng/L	0.0799	0.212
		59		NEA 207_03	ng/L	0.0525	0.128
		60		NEA 207_03	ng/L	0.0511	0.137
		61		NEA 207_03	ng/L	0.0768	0.389
		62		NEA 207_03	ng/L	0.373	25
		63		NEA 207_03	ng/L	0.0978	0.0978
		64		NEA 207_03	ng/L	0.0502	0.311
		65		NEA 207_03	ng/L	0.0150	0.0530
		66		NEA 207_03	ng/L	0.0591	0.110
		67		NEA 207_03	ng/L	0.0396	0.0475
		68		NEA 207_03	ng/L	0.135	25
		69		NEA 207_03	ng/L	0.106	0.731
		70		NEA 207_03	ng/L	0.156	25
		71		NEA 207_03	ng/L	0.0750	0.0750

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits		
Water (Cont'd)	PCBs (1 liter) (Cont'd)	72		NEA 207_03	ng/L	0.0129	0.0129		
		73		NEA 207_03	ng/L	0.0260	0.0713		
		74		NEA 207_03	ng/L	0.0802	0.248		
		75		NEA 207_03	ng/L	0.126	0.538		
		76		NEA 207_03	ng/L	0.147	25		
		77		NEA 207_03	ng/L	0.0767	0.311		
		78		NEA 207_03	ng/L	0.0351	0.267		
		79		NEA 207_03	ng/L	0.0249	0.0274		
		80		NEA 207_03	ng/L	0.0169	0.0475		
		82		NEA 207_03	ng/L	0.118	0.493		
		83		NEA 207_03	ng/L	0.0611	0.0611		
		84		NEA 207_03	ng/L	0.0067	0.0067		
		85		NEA 207_03	ng/L	0.0903	0.201		
		87		NEA 207_03	ng/L	0.0165	0.0731		
		88		NEA 207_03	ng/L	0.0926	0.658		
		89		NEA 207_03	ng/L	0.0335	0.0366		
		90		NEA 207_03	ng/L	0.0768	0.311		
		91		NEA 207_03	ng/L	0.0493	0.0493		
		92		NEA 207_03	ng/L	0.0265	0.0859		
		93		NEA 207_03	ng/L	0.130	0.585		
		94		NEA 207_03	ng/L	0.125	0.311		
		95		NEA 207_03	ng/L	0.0934	0.144		
		96		NEA 207_03	ng/L	0.00417	0.0121		
		98		NEA 207_03	ng/L	0.0799	0.0799		
		99		NEA 207_03	ng/L	0.0547	0.0713		
		100		NEA 207_03	ng/L	0.0563	0.102		
		101		NEA 207_03	ng/L	0.110	0.110		
		102		NEA 207_03	ng/L	0.163	1.11		
		103		NEA 207_03	ng/L	0.0731	0.0768		
		104		NEA 207_03	ng/L	0.0395	0.0438		
		105		NEA 207_03	ng/L	0.0247	0.0786		
		106		NEA 207_03	ng/L	0.0576	0.234		
		107		NEA 207_03	ng/L	0.0426	0.0768		
		108		NEA 207_03	ng/L	0.0443	0.0443		
		109		NEA 207_03	ng/L	0.244	0.768		
		110		NEA 207_03	ng/L	0.185	0.786		
		111		NEA 207_03	ng/L	0.0340	0.0340		
		112		NEA 207_03	ng/L	0.0492	0.101		
		113		NEA 207_03	ng/L	0.0532	0.0902		
		114 (surrogate)		NEA 207_03	ng/L	0.0236	0.0340		
		115		NEA 207_03	ng/L	0.108	0.329		
		116		NEA 207_03	ng/L	0.0623	0.0623		
		117		NEA 207_03	ng/L	0.118	0.124		
		118		NEA 207_03	ng/L	0.0236	0.0236		
			PCBs (8 liter)	Total PCB (sum of congeners)	1336-36-3	Modified Green Bay Mass Balance Method (NEA 207_03)	ng/L	1.06	4.00
				DB-1 Peak:					
				02		NEA 207_03	ng/L	0.100	0.274
				03		NEA 207_03	ng/L	1.17	125
				04		NEA 207_03	ng/L	0.100	0.160
				05		NEA 207_03	ng/L	0.0250	0.0777
				06		NEA 207_03	ng/L	0.0140	0.0274
				07		NEA 207_03	ng/L	0.0466	0.0466
				08		NEA 207_03	ng/L	0.0355	0.320
				09		NEA 207_03	ng/L	0.0275	3.13
				10		NEA 207_03	ng/L	0.00518	0.0128
				11		NEA 207_03	ng/L	0.0162	3.13
				12		NEA 207_03	ng/L	0.0365	3.13
				13		NEA 207_03	ng/L	0.00839	0.0122
				14		NEA 207_03	ng/L	0.0134	0.0845
				15		NEA 207_03	ng/L	0.0179	0.0845
				16		NEA 207_03	ng/L	0.00133	0.00594
				17		NEA 207_03	ng/L	0.0105	0.0891
				19		NEA 207_03	ng/L	0.0159	3.13
				20		NEA 207_03	ng/L	0.00107	0.00243
		21		NEA 207_03	ng/L	0.0141	0.0164		
		22		NEA 207_03	ng/L	0.00793	0.00793		
		23		NEA 207_03	ng/L	0.0190	0.0942		
		24		NEA 207_03	ng/L	0.0134	0.121		
		25		NEA 207_03	ng/L	0.0157	0.0907		
		26		NEA 207_03	ng/L	0.0446	0.0662		
		27		NEA 207_03	ng/L	0.00949	0.0203		
		28		NEA 207_03	ng/L	0.0202	3.13		

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits
Water (Cont'd)	PCBs (8 liter) (Cont'd)	29		NEA 207_03	ng/L	0.00616	0.00914
		30		NEA 207_03	ng/L	0.0225	3.13
		31		NEA 207_03	ng/L	0.0114	0.109
		32		NEA 207_03	ng/L	0.00511	0.0525
		33		NEA 207_03	ng/L	0.0283	0.0283
		34		NEA 207_03	ng/L	0.00966	0.0228
		35		NEA 207_03	ng/L	0.0231	3.13
		36		NEA 207_03	ng/L	0.0244	3.13
		37		NEA 207_03	ng/L	0.0162	0.0982
		38		NEA 207_03	ng/L	0.0160	0.0594
		39		NEA 207_03	ng/L	0.0159	0.0937
		41		NEA 207_03	ng/L	0.0270	3.13
		42		NEA 207_03	ng/L	0.0149	0.0215
		43		NEA 207_03	ng/L	0.0185	3.13
		44		NEA 207_03	ng/L	0.00337	0.00502
		45		NEA 207_03	ng/L	0.00550	0.00550
		46		NEA 207_03	ng/L	0.00723	0.0434
		47		NEA 207_03	ng/L	0.0138	0.0777
		48		NEA 207_03	ng/L	0.0266	0.164
		49		NEA 207_03	ng/L	0.00197	0.0117
		50		NEA 207_03	ng/L	0.0141	0.0799
		51		NEA 207_03	ng/L	0.0163	0.0411
		52		NEA 207_03	ng/L	0.0031	0.00457
		53		NEA 207_03	ng/L	0.00662	0.0411
		54		NEA 207_03	ng/L	0.00244	0.0169
		55		NEA 207_03	ng/L	0.00111	0.00128
		56		NEA 207_03	ng/L	0.00304	0.00685
		57		NEA 207_03	ng/L	0.00356	0.0128
		58		NEA 207_03	ng/L	0.00759	0.0265
		59		NEA 207_03	ng/L	0.00343	0.0160
		60		NEA 207_03	ng/L	0.00748	0.0171
		61		NEA 207_03	ng/L	0.00740	0.0487
		62		NEA 207_03	ng/L	0.0323	3.13
		63		NEA 207_03	ng/L	0.00664	0.0100
		64		NEA 207_03	ng/L	0.00706	0.0388
		65		NEA 207_03	ng/L	0.00210	0.00663
		66		NEA 207_03	ng/L	0.00293	0.0137
		67		NEA 207_03	ng/L	0.00543	0.00594
		68		NEA 207_03	ng/L	0.0180	3.13
		69		NEA 207_03	ng/L	0.0137	0.0914
		70		NEA 207_03	ng/L	0.0195	3.13
		71		NEA 207_03	ng/L	0.00738	0.00738
		72		NEA 207_03	ng/L	0.00073	0.00133
		73		NEA 207_03	ng/L	0.00493	0.00891
		74		NEA 207_03	ng/L	0.00568	0.0309
		75		NEA 207_03	ng/L	0.0103	0.0673
		76		NEA 207_03	ng/L	0.0271	3.13
		77		NEA 207_03	ng/L	0.0124	0.0388
		78		NEA 207_03	ng/L	0.0104	0.0334
		79		NEA 207_03	ng/L	0.00863	0.00863
		80		NEA 207_03	ng/L	0.00113	0.00594
		82		NEA 207_03	ng/L	0.00555	0.0617
		83		NEA 207_03	ng/L	0.00368	0.00571
		84		NEA 207_03	ng/L	0.000271	0.000591
		85		NEA 207_03	ng/L	0.00808	0.0251
		87		NEA 207_03	ng/L	0.00466	0.00914
		88		NEA 207_03	ng/L	0.0109	0.0822
		89		NEA 207_03	ng/L	0.00322	0.00457
		90		NEA 207_03	ng/L	0.00674	0.0388
		91		NEA 207_03	ng/L	0.00216	0.00224
		92		NEA 207_03	ng/L	0.00246	0.0107
		93		NEA 207_03	ng/L	0.00935	0.0731
		94		NEA 207_03	ng/L	0.00787	0.0388
		95		NEA 207_03	ng/L	0.00347	0.0180
		96		NEA 207_03	ng/L	0.00117	0.00151
		98		NEA 207_03	ng/L	0.00586	0.00586
		99		NEA 207_03	ng/L	0.00169	0.00891
		100		NEA 207_03	ng/L	0.00985	0.0128
		101		NEA 207_03	ng/L	0.00306	0.00502
		102		NEA 207_03	ng/L	0.0169	0.139
		103		NEA 207_03	ng/L	0.0100	0.00959
		104		NEA 207_03	ng/L	0.00239	0.00548
		105		NEA 207_03	ng/L	0.00281	0.00982

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits	
Water (Cont'd)	PCBs (8 liter) (Cont'd)	106		NEA 207_03	ng/L	0.00404	0.0292	
		107		NEA 207_03	ng/L	0.00175	0.00959	
		108		NEA 207_03	ng/L	0.00249	0.00548	
		109		NEA 207_03	ng/L	0.0148	0.0959	
		110		NEA 207_03	ng/L	0.0122	0.0982	
		111		NEA 207_03	ng/L	0.00336	0.00336	
		112		NEA 207_03	ng/L	0.00306	0.0126	
		113		NEA 207_03	ng/L	0.00816	0.0113	
		114 (surrogate)		NEA 207_03	ng/L	0.00279	0.00425	
		115		NEA 207_03	ng/L	0.00610	0.0411	
		116		NEA 207_03	ng/L	0.00933	0.00933	
	117		NEA 207_03	ng/L	0.00319	0.0155		
	118		NEA 207_03	ng/L	0.00133	0.00133		
		Select Nutrients	Nitrate	NT001	EPA 353.3	mg/L	NA	0.20
			Nitrite	NT002	EPA 354.1	mg/L	NA	0.010
			TKN	NT003	EPA 351.3	mg/L	NA	0.10
			Total Phosphorus	NT004	EPA 365.2	mg/L	NA	0.050
		TAL Metals	Ag (Silver)	7440-22-4	EPA 200.8	ug/L	0.13	1.0
			Al (Aluminum)	7429-90-5	EPA 200.8	ug/L	1.1	30.0
			As (Arsenic)	7440-38-2	EPA 200.8	ug/L	0.25	1.0
			Ba (Barium)	7440-39-3	EPA 200.8	ug/L	2.7	10.0
			Be (Beryllium)	7440-41-7	EPA 200.8	ug/L	0.066	1.0
			Ca (Calcium)	7440-70-2	EPA 200.8	ug/L	13.6	100
			Cd (Cadmium)	7440-43-9	EPA 200.8	ug/L	0.072	0.50
			Co (Cobalt)	7440-48-4	EPA 200.8	ug/L	0.031	0.50
			Cr (Chromium)	7440-47-3	EPA 200.8	ug/L	0.31	2.0
			Cu (Copper)	7440-50-8	EPA 200.8	ug/L	0.19	2.0
			Fe (Iron)	7439-89-6	EPA 200.8	ug/L	15.3	50
			Hg (Mercury)	7439-97-6	EPA 245.1	ug/L	0.076	0.20
			K (Potassium)	7440-09-7	EPA 200.8	ug/L	2.8	100
			Mg (Magnesium)	7439-95-4	EPA 200.8	ug/L	0.045	100
			Mn (Manganese)	7439-96-5	EPA 200.8	ug/L	1.2	0.50
			Na (Sodium)	7440-23-5	EPA 200.8	ug/L	4.1	100
			Ni (Nickel)	7440-02-0	EPA 200.8	ug/L	0.18	1.0
			Pb (Lead)	7439-92-1	EPA 200.8	ug/L	0.075	1.0
			Sb (Antimony)	7440-36-0	EPA 200.8	ug/L	0.071	2.0
			Se (Selenium)	7782-49-2	EPA 200.8	ug/L	0.40	5.0
			Tl (Thallium)	7440-28-0	EPA 200.8	ug/L	0.083	1.0
		V (Vanadium)	7440-62-2	EPA 200.8	ug/L	0.50	1.0	
		Zn (Zinc)	7440-66-6	EPA 200.8	ug/L	0.77	5.0	
		Other	TSS	WQ001	EPA 160.2	mg/L	NA	1.0
			DOC and POC	OC001/OC002	NE128_03	mg/L	NA	1.0 & 0.0625
		Dioxins and Furans	Total-TCDD	41903-57-5	USEPA 1613	pg/L	*	-
			2378-TCDD	1746-01-6	USEPA 1613	pg/L	*	10
			Total-TCDF	55722-27-5	USEPA 1613	pg/L	*	-
			2378-TCDF	51207-31-9	USEPA 1613	pg/L	*	10
			Total PeCDD	36088-22-9	USEPA 1613	pg/L	*	-
			12378-PeCDD	40321-76-4	USEPA 1613	pg/L	*	50
			Total PeCDF	30402-15-4	USEPA 1613	pg/L	*	-
			12378-PeCDF	57117-41-6	USEPA 1613	pg/L	*	50
			23478-PeCDF	57117-31-4	USEPA 1613	pg/L	*	50
			Total-HxCDD	34465-46-8	USEPA 1613	pg/L	*	-
			123478-HxCDD	39227-28-6	USEPA 1613	pg/L	*	50
			123678-HxCDD	57653-85-7	USEPA 1613	pg/L	*	50
			123789-HxCDD	19408-74-3	USEPA 1613	pg/L	*	50
			Total HxCDF	55684-94-1	USEPA 1613	pg/L	*	-
			123478-HxCDF	70648-26-9	USEPA 1613	pg/L	*	50
			123678-HxCDF	57117-44-9	USEPA 1613	pg/L	*	50
			123789-HxCDF	72918-21-9	USEPA 1613	pg/L	*	50
			234678-HxCDF	60851-34-5	USEPA 1613	pg/L	*	50
			Total HpCDD	37871-00-4	USEPA 1613	pg/L	*	-
			1234678-HpCDD	35822-46-9	USEPA 1613	pg/L	*	50
			Total HpCDF	38998-75-3	USEPA 1613	pg/L	*	-
			1234678-HpCDF	67562-39-4	USEPA 1613	pg/L	*	50
			1234789-HpCDF	55673-89-7	USEPA 1613	pg/L	*	50
			OCDD	3268-87-9	USEPA 1613	pg/L	*	100
			OCDF	39001-02-0	USEPA 1613	pg/L	*	100

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits
Fish	PCBs (Congeners) DB-1 Peak:	Total PCB (sum of congeners)	1336-36-3	Green Bay Mass Balance Method (NEA 013_07)	ug/g	0.0123	0.313
		02		NEA 013_07	ug/g	0.000667	0.0219
		03		NEA 013_07	ug/g	0.0292	5.00
		04		NEA 013_07	ug/g	0.000795	0.0128
		05		NEA 013_07	ug/g	0.000196	0.00621
Fish (Cont'd)	PCBs (Congeners) (Cont'd)	06		NEA 013_07	ug/g	0.0000653	0.00219
		07		NEA 013_07	ug/g	0.000116	0.00347
		08		NEA 013_07	ug/g	0.000814	0.0256
		09		NEA 013_07	ug/g	0.000847	0.125
		10		NEA 013_07	ug/g	0.0000287	0.000512
		11		NEA 013_07	ug/g	0.000564	0.125
		12		NEA 013_07	ug/g	0.000663	0.125
		13		NEA 013_07	ug/g	0.000189	0.000488
		14		NEA 013_07	ug/g	0.000317	0.00676
		15		NEA 013_07	ug/g	0.000219	0.00676
		16		NEA 013_07	ug/g	0.0000157	0.000475
		17		NEA 013_07	ug/g	0.000203	0.00713
		19		NEA 013_07	ug/g	0.000436	0.125
		20		NEA 013_07	ug/g	0.0000869	0.0000970
		21		NEA 013_07	ug/g	0.0000590	0.00132
		22		NEA 013_07	ug/g	0.0000364	0.000585
		23		NEA 013_07	ug/g	0.000209	0.00753
		24		NEA 013_07	ug/g	0.000259	0.00964
		25		NEA 013_07	ug/g	0.000206	0.00726
		26		NEA 013_07	ug/g	0.000210	0.00530
		27		NEA 013_07	ug/g	0.0000564	0.00163
		28		NEA 013_07	ug/g	0.000822	0.125
		29		NEA 013_07	ug/g	0.0000208	0.000731
		30		NEA 013_07	ug/g	0.000570	0.125
		31		NEA 013_07	ug/g	0.000623	0.00872
		32		NEA 013_07	ug/g	0.000136	0.00420
		33		NEA 013_07	ug/g	0.0000613	0.00183
		34		NEA 013_07	ug/g	0.0000917	0.00183
		35		NEA 013_07	ug/g	0.000442	0.125
		36		NEA 013_07	ug/g	0.000658	0.125
		37		NEA 013_07	ug/g	0.000250	0.00786
		38		NEA 013_07	ug/g	0.000131	0.00475
		39		NEA 013_07	ug/g	0.000256	0.00749
		41		NEA 013_07	ug/g	0.000828	0.125
		42		NEA 013_07	ug/g	0.0000698	0.00172
		43		NEA 013_07	ug/g	0.000443	0.125
		44		NEA 013_07	ug/g	0.00000693	0.000201
		45		NEA 013_07	ug/g	0.0000152	0.000384
		46		NEA 013_07	ug/g	0.000132	0.00347
		47		NEA 013_07	ug/g	0.000191	0.00621
		48		NEA 013_07	ug/g	0.000367	0.0132
		49		NEA 013_07	ug/g	0.0000436	0.000932
		50		NEA 013_07	ug/g	0.000174	0.00640
		51		NEA 013_07	ug/g	0.000109	0.00329
		52		NEA 013_07	ug/g	0.000161	0.000183
		53		NEA 013_07	ug/g	0.000190	0.00329
		54		NEA 013_07	ug/g	0.0000542	0.00135
		55		NEA 013_07	ug/g	0.0000254	0.0000512
		56		NEA 013_07	ug/g	0.000177	0.000274
		57		NEA 013_07	ug/g	0.0000294	0.00102
		58		NEA 013_07	ug/g	0.0000777	0.00212
		59		NEA 013_07	ug/g	0.0000386	0.00128
		60		NEA 013_07	ug/g	0.0000486	0.00137
		61		NEA 013_07	ug/g	0.000123	0.00389
		62		NEA 013_07	ug/g	0.000612	0.125
		63		NEA 013_07	ug/g	0.0000282	0.000804
		64		NEA 013_07	ug/g	0.000101	0.00311
		65		NEA 013_07	ug/g	0.0000386	0.000530
		66		NEA 013_07	ug/g	0.0000372	0.00110
		67		NEA 013_07	ug/g	0.00000918	0.000237
		68		NEA 013_07	ug/g	0.000531	0.125
		69		NEA 013_07	ug/g	0.000243	0.00731
		70		NEA 013_07	ug/g	0.000547	0.125
		71		NEA 013_07	ug/g	0.000230	0.000369
		72		NEA 013_07	ug/g	0.0000569	0.0000569
		73		NEA 013_07	ug/g	0.0000259	0.000713
		74		NEA 013_07	ug/g	0.0000827	0.00248
		75		NEA 013_07	ug/g	0.000157	0.00538

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits
		76		NEA 013_07	ug/g	0.000668	0.125
		77		NEA 013_07	ug/g	0.000122	0.00311
		78		NEA 013_07	ug/g	0.000148	0.00267
		79		NEA 013_07	ug/g	0.0000928	0.000137
		80		NEA 013_07	ug/g	0.0000169	0.000475
Fish (Cont'd)	PCBs (Congeners) (Cont'd)	82		NEA 013_07	ug/g	0.000143	0.00493
		83		NEA 013_07	ug/g	0.0000213	0.000457
		84		NEA 013_07	ug/g	0.0000165	0.0000236
		85		NEA 013_07	ug/g	0.0000930	0.00201
		87		NEA 013_07	ug/g	0.0000100	0.000366
		88		NEA 013_07	ug/g	0.000208	0.00658
		89		NEA 013_07	ug/g	0.0000122	0.000183
		90		NEA 013_07	ug/g	0.0000863	0.00311
		91		NEA 013_07	ug/g	0.0000101	0.0000897
		92		NEA 013_07	ug/g	0.0000277	0.000859
		93		NEA 013_07	ug/g	0.000158	0.00585
		94		NEA 013_07	ug/g	0.0000888	0.00311
		95		NEA 013_07	ug/g	0.0000731	0.00144
		96		NEA 013_07	ug/g	0.00000475	0.000121
		98		NEA 013_07	ug/g	0.0000568	0.0000695
		99		NEA 013_07	ug/g	0.0000297	0.000713
		100		NEA 013_07	ug/g	0.0000374	0.00102
		101		NEA 013_07	ug/g	0.0000121	0.000201
102		NEA 013_07	ug/g	0.000317	0.0111		
103		NEA 013_07	ug/g	0.0000328	0.000768		
104		NEA 013_07	ug/g	0.000148	0.000219		
105		NEA 013_07	ug/g	0.0000293	0.000786		
106		NEA 013_07	ug/g	0.0000726	0.00234		
107		NEA 013_07	ug/g	0.0000260	0.000768		
108		NEA 013_07	ug/g	0.00000968	0.000219		
109		NEA 013_07	ug/g	0.000251	0.00768		
110		NEA 013_07	ug/g	0.000218	0.00786		
111		NEA 013_07	ug/g	0.0000798	0.0000798		
112		NEA 013_07	ug/g	0.0000605	0.00101		
113		NEA 013_07	ug/g	0.0000203	0.000451		
114		NEA 013_07	ug/g	surrogate**	surrogate**		
115		NEA 013_07	ug/g	0.0000881	0.00329		
116		NEA 013_07	ug/g	0.0000833	0.000201		
117		NEA 013_07	ug/g	0.0000453	0.00124		
118		NEA 013_07	ug/g	0.0000187	0.0000222		
	PCBs (Aroclors)	Aroclor 1016	12674-11-2	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1221	11104-28-2	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1232	11141-16-5	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1242	53469-21-9	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1248	12672-29-6	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1254	11097-69-1	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Aroclor 1260	11096-82-5	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
		Total PCBs (sum of Aroclors)	1336-36-3	SW846 8082 (NE148_04)	ug/g	0.0104	0.050
	Other	Percent Lipid	LP001	NE158_03	%	NA	0.01
		Mercury	7439-97-6	SW-846 7471A	ug/g	0.00115	0.020
	Dioxins and Furans	Total-TCDD	41903-57-5	USEPA 1613	pg/g	*	-
		2378-TCDD	1746-01-6	USEPA 1613	pg/g	*	1
		Total-TCDF	55722-27-5	USEPA 1613	pg/g	*	-
		2378-TCDF	51207-31-9	USEPA 1613	pg/g	*	1
		Total PeCDD	36088-22-9	USEPA 1613	pg/g	*	-
		12378-PeCDD	40321-76-4	USEPA 1613	pg/g	*	5
		Total PeCDF	30402-15-4	USEPA 1613	pg/g	*	-
		12378-PeCDF	57117-41-6	USEPA 1613	pg/g	*	5
		23478-PeCDF	57117-31-4	USEPA 1613	pg/g	*	5
		Total-HxCDD	34465-46-8	USEPA 1613	pg/g	*	-
		123478-HxCDD	39227-28-6	USEPA 1613	pg/g	*	5
		123678-HxCDD	57653-85-7	USEPA 1613	pg/g	*	5
		123789-HxCDD	19408-74-3	USEPA 1613	pg/g	*	5
		Total HxCDF	55684-94-1	USEPA 1613	pg/g	*	-
		123478-HxCDF	70648-26-9	USEPA 1613	pg/g	*	5
		123678-HxCDF	57117-44-9	USEPA 1613	pg/g	*	5
		123789-HxCDF	72918-21-9	USEPA 1613	pg/g	*	5
		234678-HxCDF	60851-34-5	USEPA 1613	pg/g	*	5
		Total HpCDD	37871-00-4	USEPA 1613	pg/g	*	-
		1234678-HpCDD	35822-46-9	USEPA 1613	pg/g	*	5
		Total HpCDF	38998-75-3	USEPA 1613	pg/g	*	-
	1234678-HpCDF	67562-39-4	USEPA 1613	pg/g	*	5	
	1234789-HpCDF	55673-89-7	USEPA 1613	pg/g	*	5	

Table B-6. Reference limit and evaluation table.

Matrix	Category	Analyte Name	CAS number	Analytical Method	Units	Laboratory Method Detection Limits	Laboratory Reporting Limits
		OCDD	3268-87-9	USEPA 1613	pg/g	*	10
		OCDF	39001-02-0	USEPA 1613	pg/g	*	10
	Organochlorine Pesticides	Aldrin	309-00-2	SW846 8081A (NE131_03)	ug/g	0.000931	0.00250
		α-BHC	319-84-6	SW846 8081A (NE131_03)	ug/g	0.00134	0.00250
Fish (Cont'd)	Organochlorine Pesticides (Cont'd)	β-BHC	319-85-7	SW846 8081A (NE131_03)	ug/g	0.000373	0.00250
		γ-BHC (Lindane)	58-89-9	SW846 8081A (NE131_03)	ug/g	0.000996	0.00250
		δ-BHC	319-86-8	SW846 8081A (NE131_03)	ug/g	0.000880	0.00250
		α-Chlordane	5103-71-9	SW846 8081A (NE131_03)	ug/g	0.000502	0.00250
		γ-Chlordane	5103-74-2	SW846 8081A (NE131_03)	ug/g	0.000819	0.00250
		Chlordane	57-74-9	SW846 8081A (NE131_03)	ug/g	0.00632	0.125
		4,4'-DDD	72-54-8	SW846 8081A (NE131_03)	ug/g	0.000818	0.00250
		4,4'-DDE	72-55-9	SW846 8081A (NE131_03)	ug/g	0.000875	0.00250
		4,4'-DDT	50-29-3	SW846 8081A (NE131_03)	ug/g	0.000737	0.00250
		Dieldrin	60-57-1	SW846 8081A (NE131_03)	ug/g	0.000835	0.00250
		Endosulfan I	959-98-8	SW846 8081A (NE131_03)	ug/g	0.000500	0.00250
		Endosulfan II	33213-65-9	SW846 8081A (NE131_03)	ug/g	0.00108	0.00250
		Endosulfan sulfate	1031-07-8	SW846 8081A (NE131_03)	ug/g	0.00179	0.00250
		Endrin	72-20-8	SW846 8081A (NE131_03)	ug/g	0.00136	0.00250
		Endrin aldehyde	7421-93-4	SW846 8081A (NE131_03)	ug/g	0.000920	0.00250
		Endrin ketone	53494-70-5	SW846 8081A (NE131_03)	ug/g	0.00108	0.00250
		Heptachlor	76-44-8	SW846 8081A (NE131_03)	ug/g	0.000391	0.00250
		Heptachlor epoxide	1024-57-3	SW846 8081A (NE131_03)	ug/g	0.00154	0.00250
		Methoxychlor	72-43-5	SW846 8081A (NE131_03)	ug/g	0.000523	0.00250
		Toxaphene	8001-35-2	SW846 8081A (NE131_03)	ug/g	0.0106	0.250

Note:
 NA - Not Applicable. Method detection limit (MDL) reporting will not be used for this analyte. The analyte will be reported to the Reporting Limit (RL).
 * - Dioxins and Furans will be reported to estimated detection limits (EDLs) that are calculated on an analyte- and sample-specific basis as addressed in the analytical SOP.
 ** - Peak 114 corresponds to IUPAC 207, which is the surrogate. The surrogate was not included in the Green Bay MDL study performed for fish.

Table B-7a. Measurement Performance Criteria.

Matrix: Water
 Analytical Parameter: Total PCBs as Congeners
 Concentration Level: Low to High
 Method: Green Bay Congener Method (NEA 207_03, QAPP Appendix 9)

Data Quality Indicators (DQIs) ¹	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL, or associated samples >5x blank values	Laboratory or Equipment Blank	S&A
	60-140% R (NE207_03)	Laboratory Control Sample	A
	60-140% R (NE207_03)	Matrix Spike	S&A
	60-140% R (NE207_03)	Surrogates	A
Precision	The RPD for water field duplicates should be ≤ 35% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Field Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7b. Measurement Performance Criteria.

Matrix: Water
 Analytical Parameter: Select nutrients
 Concentration Level: Low to High
 Method: EPA 353.3, EPA 354.1, 351.3, 365.2 (QAPP Appendices 10-13)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	<Sample-specific RL, or associated samples >5x blank values	Laboratory or Equipment Blank	A
	Laboratory-specified control limits	Laboratory Control Sample	A
	Laboratory-specified control limits	Matrix Spike	S&A
Precision	Laboratory-specified control limits	Laboratory Duplicates	A
	The RPD for water field duplicates should be ≤20% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Field Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7c. Measurement Performance Criteria.

Matrix: Water
 Analytical Parameter: TAL Metals (except Mercury)
 Concentration Level: Low to High
 Method: EPA 200.8 (QAPP Appendix 15)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL or associated samples >5X blank values	Laboratory or Equipment Blank	S&A
	85-115% R	Laboratory Control Sample	A
	70-130% R (NA if sample concentration is >4X spike added)	Matrix Spike	S&A
Precision	The RPD for water lab duplicates should be ≤20% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Laboratory Duplicates	A
	The RPD for water field duplicates should be ≤20% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Field Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7d. Measurement Performance Criteria.

Matrix: Fish and Water
 Analytical Parameter: Mercury
 Concentration Level: Low to High
 Method: EPA 245.1, SW-846 7470A/ 7471A (QAPP Appendices 16 and 17)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL or associated samples >5X blank values	Laboratory or Equipment (water only) Blank	S&A
	80-120% R (water) 70-130% R (solid)	Laboratory Control Sample	A
	75-125% R (NA if sample concentration is >4X spike added)	Matrix Spike	S&A
Precision	The RPD for fish lab duplicates should be ≤40 for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Laboratory Duplicates	A
	The RPD for water field duplicates should be ≤20% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Field Duplicates (water only)	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7e. Measurement Performance Criteria.

Matrix: Water
 Analytical Parameter: Total Suspended Solids
 Concentration Level: Low to High
 Method: EPA 160.2 (QAPP Appendix 18)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL or associated samples >5X blank values	Laboratory or Equipment Blank	S&A
	Recovery Limit supplied with Control	Laboratory Control Sample	A
Precision	The RPD for water lab duplicates should be $\leq 20\%$ for results $>5x$ the RL. The difference between results should be $\pm RL$ when at least one result is $\leq 5x$ the RL	Laboratory Duplicates	A
	The RPD for water field duplicates should be $\leq 20\%$ for results $>5x$ the RL. The difference between results should be $\pm RL$ when at least one result is $\leq 5x$ the RL	Field Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7f. Measurement Performance Criteria.

Matrix: Water
 Analytical Parameter: Total and Particulate Organic Carbon
 Concentration Level: Low to High
 Method: UV Promoted Persulfate Infrared Method (NE128_03, QAPP Appendix 19)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL or associated samples >5X blank values	Laboratory or Equipment Blank	S&A
	Laboratory Control Limit	Matrix Spike	S&A
Precision	The RPD for water lab duplicates should be ≤20% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Laboratory Duplicates	A
	The RPD for water field duplicates should be ≤20% for results >5x the RL. The difference between results should be ±RL when at least one result is ≤5x the RL	Field Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7g. Measurement Performance Criteria.

Matrix: Fish and Water
 Analytical Parameter: Dioxins/Furans
 Concentration Level: Low to High
 Method: EPA 1613B (QAPP Appendix 20)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	<RL or associated samples >5X blank value (or >10X for OCDD)	Laboratory or Equipment (water only) Blank	S&A
	Method 1613B-specified limits (see SOP, Appendix 20)	Laboratory Control Sample/On-going Precision Recovery (OPR) sample	A
Precision	The RPD for fish lab duplicates should be ≤40% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Lab Duplicates (fish only)	A
	The RPD for water field duplicates should be ≤20% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Field Duplicates (water only)	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7h. Measurement Performance Criteria.

Matrix: Fish
 Analytical Parameter: Total PCBs as Aroclors
 Concentration Level: Low to High
 Method: SW-846 8082 (NE148_04, QAPP Appendix 25)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	< RL or associated samples >5X blank values	Laboratory Blank	A
	laboratory-specified limits	Laboratory Control Sample	A
	laboratory-specified limits	Standard Reference Material	A
	laboratory-specified limits	Matrix Spike/Matrix Spike Duplicate	
	laboratory-specified limits	Surrogates	A
Precision	The RPD for fish lab duplicates should be $\leq 40\%$ for results $> 5x$ the RL. The difference between results should be $< 2x$ the RL when at least one result is $\leq 5x$ the RL	Laboratory Duplicates	A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7i. Measurement Performance Criteria.

Matrix: Fish
 Analytical Parameter: Total PCBs as Congeners
 Concentration Level: Low to High
 Method: NEA 013_07 (QAPP Appendix 26)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	<Sample-specific RL, or associated samples >5x blank values	Laboratory Blank	A
	70-130% R	Laboratory Control Sample	A
	70-130% R	Matrix Spike/Matrix Spike Duplicate	S&A
	70-130% R	Surrogates	A
Precision	The RPD for fish lab duplicates should be ≤40% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Laboratory Duplicates	A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7j. Measurement performance criteria.

Matrix: Fish
 Analytical Parameter: Percent Lipid
 Concentration Level: Low to High
 Method: NE158_03 (QAPP Appendix 24)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	<RL, or associated samples >5x blank values	Laboratory Blank	A
Precision	The RPD for lab duplicates should be ≤20% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Laboratory Duplicates	S&A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-7k. Measurement Performance Criteria.

Matrix: Fish
 Analytical Parameter: Organochlorine Pesticides
 Concentration Level: Low to High
 Method: SW-846-8081A (NE131_03, QAPP Appendix 27)

Data Quality Indicators (DQIs)	Measurement Performance Criteria	QC Sample and/or activity used to assess measurement performance	QC Sample Assesses Error for Sampling (S), Analytical (A) or both (S&A)
Accuracy	<Sample-specific RL, or associated samples >5x blank values	Laboratory Blank	A
	laboratory-specified limits	Laboratory Control Sample	A
	laboratory-specified limits	Matrix Spike/Matrix Spike Duplicate	S&A
	laboratory-specified limits	Surrogates	A
Precision	The RPD for fish lab duplicates should be ≤40% for results >5x the RL. The difference between results should be <2x the RL when at least one result is ≤5x the RL	Laboratory Duplicates	A
Sensitivity	See Table B-6	Reporting Limits	A
Representativeness	Use of standardized collection and analytical methods	Field audits and laboratory audits. See QAPP Section A7.3.3	S&A
Completeness	95%	See QAPP Section A7.3.5	S&A
Comparability	Based on accuracy and media comparison	Use of standardized SOPs by field and analytical contractors	S&A

RL = Reporting Limit; R = Recovery; RPD = Relative Percent Difference; LWL = Lower Warning Limit; UWL = Upper Warning Limit; LCL = Lower Control Limit; UCL = Upper Control Limit

¹ Data Quality Indicators (a.k.a. PASRCC parameters, *i.e.*, precision, accuracy/bias, sensitivity, representativeness, data completeness, comparability).

Table B-8. Field information recorded during water sampling.

Data Field	Valid Values	Data Entry Type
Program ID	RTN (routine sampling) TOT (time-of-travel sampling) HFL (high flow sampling)	Drop-down selection list
Location ID	BF (Bakers Falls) RI (Rogers Island) TI (Thompson Island) SV (Schuylerville) SW (Stillwater) WF (Waterford) MR (Mohawk River at Cohoes) L1 (Lock1)	Drop-down selection list
Date collected	MM/DD/YYYY	Automatic (based on current computer time)
Sample ID		Automatic (based on program, location, and date)
General description		Manual
Sample Type	center channel E-W composite transect composite	Drop-down selection list
Sampler Initials		Manual
Crew ID		Automatic (value loaded on computer)
Time ¹	HH:MM	Automatic (based on current computer time)
Specific Conductivity ¹	≥ 0	Manual or automatic (using data logger)
Temperature ¹	≥ 0	Manual or automatic (using data logger)
Dissolved Oxygen ¹	≥ 0	Manual or automatic (using data logger)
pH ¹	≥ 0	Manual or automatic (using data logger)
Turbidity ¹	≥ 0	Manual or automatic (using data logger)
Depth ¹	≥ 0	Manual or automatic (using data logger)

¹ One entry for each transect point

Table B-9. Field information recorded during fish sampling.

Data Field	Valid Values	Data Entry Type
Location ID		Drop-down selection list
Sample Collection Method	NET (netting) ELT (electroshocking) ANG (angling)	Drop-down selection list
Date collected	MM/DD/YYYY	Automatic (based on current computer time)
Start Time	HH:MM	Automatic (based on current computer time)
End Time	HH:MM	Automatic (based on current computer time)
Temperature	≥ 0	Manual
Turbidity	≥ 0	Manual
Northing	≥ 0	Manual
Easting	≥ 0	Manual
Distance	≥ 0	Manual
Weather Conditions		Manual
Sampling Event ID		Automatic
Sample ID ¹		Automatic
Species ¹		Drop-down selection list
Sample Type	Individual Composite	Drop-down selection list
Sample Prep	fillet, offal, or whole body	Drop-down selection list
Weight ¹	≥ 0	Manual
Length ¹	≥ 0	Manual
Sex ¹	M (male) F (female) ND (undetermined)	Drop-down selection list
General description ¹		Manual
Sampler Initials		Manual

¹ One entry for individual. Individuals in a composite will be recorded separately.

Table B-10 Valid values for Baseline Monitoring Program.

Analytic - Water

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
PCB Congeners	NE207_03	PK001	Peak 1	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK002	Peak 2	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK003	Peak 3	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK004	Peak 4	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK005	Peak 5	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK006	Peak 6	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK007	Peak 7	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK008	Peak 8	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK009	Peak 9	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK010	Peak 10	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK011	Peak 11	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK012	Peak 12	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK013	Peak 13	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK014	Peak 14	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK015	Peak 15	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK016	Peak 16	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK017	Peak 17	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK018	Peak 18	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK019	Peak 19	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK020	Peak 20	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK021	Peak 21	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK022	Peak 22	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK023	Peak 23	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK024	Peak 24	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK025	Peak 25	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK026	Peak 26	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK027	Peak 27	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK028	Peak 28	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK029	Peak 29	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK030	Peak 30	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK031	Peak 31	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK032	Peak 32	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK033	Peak 33	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK034	Peak 34	ng/L	SW846 (3535/3540C/3545)

Table B-10 Valid values for Baseline Monitoring Program.*Analytic - Water*

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE207_03	PK035	Peak 35	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK036	Peak 36	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK037	Peak 37	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK038	Peak 38	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK039	Peak 39	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK040	Peak 40	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK041	Peak 41	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK042	Peak 42	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK043	Peak 43	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK044	Peak 44	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK045	Peak 45	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK046	Peak 46	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK047	Peak 47	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK048	Peak 48	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK049	Peak 49	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK050	Peak 50	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK051	Peak 51	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK052	Peak 52	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK053	Peak 53	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK054	Peak 54	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK055	Peak 55	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK056	Peak 56	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK057	Peak 57	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK058	Peak 58	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK059	Peak 59	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK060	Peak 60	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK061	Peak 61	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK062	Peak 62	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK063	Peak 63	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK064	Peak 64	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK065	Peak 65	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK066	Peak 66	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK067	Peak 67	ng/L	SW846 (3535/3540C/3545)

Table B-10 Valid values for Baseline Monitoring Program.

Analytic - Water

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE207_03	PK068	Peak 68	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK069	Peak 69	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK070	Peak 70	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK071	Peak 71	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK072	Peak 72	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK073	Peak 73	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK074	Peak 74	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK075	Peak 75	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK076	Peak 76	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK077	Peak 77	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK078	Peak 78	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK079	Peak 79	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK080	Peak 80	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK081	Peak 81	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK082	Peak 82	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK083	Peak 83	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK084	Peak 84	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK085	Peak 85	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK086	Peak 86	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK087	Peak 87	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK088	Peak 88	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK089	Peak 89	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK090	Peak 90	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK091	Peak 91	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK092	Peak 92	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK093	Peak 93	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK094	Peak 94	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK095	Peak 95	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK096	Peak 96	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK097	Peak 97	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK098	Peak 98	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK099	Peak 99	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK100	Peak 100	ng/L	SW846 (3535/3540C/3545)

Table B-10 Valid values for Baseline Monitoring Program.

Analytic - Water

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE207_03	PK101	Peak 101	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK102	Peak 102	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK103	Peak 103	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK104	Peak 104	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK105	Peak 105	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK106	Peak 106	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK107	Peak 107	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK108	Peak 108	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK109	Peak 109	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK110	Peak 110	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK111	Peak 111	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK112	Peak 112	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK113	Peak 113	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK114	Peak 114	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK115	Peak 115	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK116	Peak 116	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK117	Peak 117	ng/L	SW846 (3535/3540C/3545)
	NE207_03	PK118	Peak 118	ng/L	SW846 (3535/3540C/3545)
	NE207_03	1336-36-3	Total PCB	ng/L	SW846 (3535/3540C/3545)
Nutrients	EPA 353.3	NT001	Nitrate	mg/L	EPA 353.3
	EPA 354.1	NT002	Nitrite	mg/L	EPA 354.1
	EPA 351.3	NT003	TKN	mg/L	EPA 351.3
	EPA 365.2	NT004	Total Phosphorous	mg/L	EPA 365.2
TAL Metals	EPA 200.8	7440-22-4	TAL - Silver	mg/L	SW846 3005A/200_7
	EPA 200.8	7429-90-5	TAL - Aluminum	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-36-0	TAL - Antimony	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-38-2	TAL - Arsenic	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-39-3	TAL - Barium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-41-7	TAL - Beryllium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-70-2	TAL - Calcium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-43-9	TAL - Cadmium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-48-4	TAL - Cobalt	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-47-3	TAL - Chromium	mg/L	SW846 3005A/200_7

Table B-10 Valid values for Baseline Monitoring Program.

Analytic - Water

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	EPA 200.8	7440-50-8	TAL - Copper	mg/L	SW846 3005A/200_7
	EPA 200.8	7439-89-6	TAL - Iron	mg/L	SW846 3005A/200_7
	EPA 245.1	7439-97-6	TAL - Mercury	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-09-7	TAL - Potassium	mg/L	SW846 3005A/200_7
	EPA 200.8	7439-95-4	TAL - Magnesium	mg/L	SW846 3005A/200_7
	EPA 200.8	7439-96-5	TAL - Manganese	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-23-5	TAL - Sodium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-02-0	TAL - Nickel	mg/L	SW846 3005A/200_7
	EPA 200.8	7439-92-1	TAL - Lead	mg/L	SW846 3005A/200_7
	EPA 200.8	7782-49-2	TAL - Selenium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-28-0	TAL - Thallium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-62-2	TAL - Vanadium	mg/L	SW846 3005A/200_7
	EPA 200.8	7440-66-6	TAL - Zinc	mg/L	SW846 3005A/200_7
TSS	EPA 160.2	WQ001	Total Suspended Solids	mg/L	EPA 160.2
Organic Carbon	NE128_03	OC001	Particulate OC	mg/L	NE128_03
	NE128_03	OC002	Dissolved OC	mg/L	NE128_03
	NE128_03	OC003	Total OC	mg/L	NE128_03
Dioxins and Furans	EPA 1613B	41903-57-5	Total-TCDD	pg/uL	EPA 1613B
	EPA 1613B	1746-01-6	2,3,7,8-TCDD	pg/uL	EPA 1613B
	EPA 1613B	55722-27-5	Total-TCDF	pg/uL	EPA 1613B
	EPA 1613B	51207-31-9	2,3,7,8-TCDF	pg/uL	EPA 1613B
	EPA 1613B	36088-22-9	Total PeCDD	pg/uL	EPA 1613B
	EPA 1613B	40321-76-4	1,2,3,7,8-PeCDD	pg/uL	EPA 1613B
	EPA 1613B	30402-15-4	Total PeCDF	pg/uL	EPA 1613B
	EPA 1613B	57117-41-6	1,2,3,7,8-PeCDF	pg/uL	EPA 1613B
	EPA 1613B	57117-31-4	2,3,4,7,8-PeCDF	pg/uL	EPA 1613B
	EPA 1613B	34465-46-8	Total HxCDD	pg/uL	EPA 1613B
	EPA 1613B	39227-28-6	1,2,3,4,7,8-HxCDD	pg/uL	EPA 1613B
	EPA 1613B	57653-85-7	1,2,3,6,7,8-HxCDD	pg/uL	EPA 1613B
	EPA 1613B	19408-74-3	1,2,3,7,8,9-HxCDD	pg/uL	EPA 1613B
	EPA 1613B	55684-94-1	Total HxCDF	pg/uL	EPA 1613B
	EPA 1613B	70648-26-9	1,2,3,4,7,8-HxCDF	pg/uL	EPA 1613B
	EPA 1613B	57117-44-9	1,2,3,6,7,8-HxCDF	pg/uL	EPA 1613B

Table B-10 Valid values for Baseline Monitoring Program.*Analytic - Water*

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	EPA 1613B	72918-21-9	1,2,3,7,8,9-HxCDF	pg/uL	EPA 1613B
	EPA 1613B	60851-34-5	2,3,4,6,7,8-HxCDF	pg/uL	EPA 1613B
	EPA 1613B	37871-00-4	Total HpCDD	pg/uL	EPA 1613B
	EPA 1613B	35822-46-9	1,2,3,4,6,7,8-HpCDD	pg/uL	EPA 1613B
	EPA 1613B	38998-75-3	Total HpCDF	pg/uL	EPA 1613B
	EPA 1613B	67562-39-4	1,2,3,4,6,7,8-HpCDF	pg/uL	EPA 1613B
	EPA 1613B	55673-89-7	1,2,3,4,7,8,9-HpCDF	pg/uL	EPA 1613B
	EPA 1613B	3268-87-9	OCDD	pg/uL	EPA 1613B
	EPA 1613B	39001-02-0	OCDF	pg/uL	EPA 1613B

Table B-10 Valid values for Baseline Monitoring Program.

Analytical - Fish

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
PCB Aroclors	SW846 8082 (NE148_04)	12674-11-2	Aroclor 1016	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	11104-28-2	Aroclor 1221	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	11141-16-5	Aroclor 1232	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	53469-21-9	Aroclor 1242	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	12672-29-6	Aroclor 1248	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	11097-69-1	Aroclor 1254	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	11096-82-5	Aroclor 1260	mg/Kg	NE17_07
	SW846 8082 (NE148_04)	1336-36-3	Total PCBs	mg/Kg	NE17_07
PCB Congeners	NE013_07	PK001	Peak 1	mg/Kg	NE17_07
	NE013_07	PK002	Peak 2	mg/Kg	NE17_07
	NE013_07	PK003	Peak 3	mg/Kg	NE17_07
	NE013_07	PK004	Peak 4	mg/Kg	NE17_07
	NE013_07	PK005	Peak 5	mg/Kg	NE17_07
	NE013_07	PK006	Peak 6	mg/Kg	NE17_07
	NE013_07	PK007	Peak 7	mg/Kg	NE17_07
	NE013_07	PK008	Peak 8	mg/Kg	NE17_07
	NE013_07	PK009	Peak 9	mg/Kg	NE17_07
	NE013_07	PK010	Peak 10	mg/Kg	NE17_07
	NE013_07	PK011	Peak 11	mg/Kg	NE17_07
	NE013_07	PK012	Peak 12	mg/Kg	NE17_07
	NE013_07	PK013	Peak 13	mg/Kg	NE17_07
	NE013_07	PK014	Peak 14	mg/Kg	NE17_07
	NE013_07	PK015	Peak 15	mg/Kg	NE17_07
	NE013_07	PK016	Peak 16	mg/Kg	NE17_07
	NE013_07	PK017	Peak 17	mg/Kg	NE17_07
	NE013_07	PK018	Peak 18	mg/Kg	NE17_07
	NE013_07	PK019	Peak 19	mg/Kg	NE17_07
	NE013_07	PK020	Peak 20	mg/Kg	NE17_07
	NE013_07	PK021	Peak 21	mg/Kg	NE17_07
	NE013_07	PK022	Peak 22	mg/Kg	NE17_07
	NE013_07	PK023	Peak 23	mg/Kg	NE17_07
	NE013_07	PK024	Peak 24	mg/Kg	NE17_07
	NE013_07	PK025	Peak 25	mg/Kg	NE17_07

Table B-10 Valid values for Baseline Monitoring Program.

Analytical - Fish

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE013_07	PK026	Peak 26	mg/Kg	NE17_07
	NE013_07	PK027	Peak 27	mg/Kg	NE17_07
	NE013_07	PK028	Peak 28	mg/Kg	NE17_07
	NE013_07	PK029	Peak 29	mg/Kg	NE17_07
	NE013_07	PK030	Peak 30	mg/Kg	NE17_07
	NE013_07	PK031	Peak 31	mg/Kg	NE17_07
	NE013_07	PK032	Peak 32	mg/Kg	NE17_07
	NE013_07	PK033	Peak 33	mg/Kg	NE17_07
	NE013_07	PK034	Peak 34	mg/Kg	NE17_07
	NE013_07	PK035	Peak 35	mg/Kg	NE17_07
	NE013_07	PK036	Peak 36	mg/Kg	NE17_07
	NE013_07	PK037	Peak 37	mg/Kg	NE17_07
	NE013_07	PK038	Peak 38	mg/Kg	NE17_07
	NE013_07	PK039	Peak 39	mg/Kg	NE17_07
	NE013_07	PK040	Peak 40	mg/Kg	NE17_07
	NE013_07	PK041	Peak 41	mg/Kg	NE17_07
	NE013_07	PK042	Peak 42	mg/Kg	NE17_07
	NE013_07	PK043	Peak 43	mg/Kg	NE17_07
	NE013_07	PK044	Peak 44	mg/Kg	NE17_07
	NE013_07	PK045	Peak 45	mg/Kg	NE17_07
	NE013_07	PK046	Peak 46	mg/Kg	NE17_07
	NE013_07	PK047	Peak 47	mg/Kg	NE17_07
	NE013_07	PK048	Peak 48	mg/Kg	NE17_07
	NE013_07	PK049	Peak 49	mg/Kg	NE17_07
	NE013_07	PK050	Peak 50	mg/Kg	NE17_07
	NE013_07	PK051	Peak 51	mg/Kg	NE17_07
	NE013_07	PK052	Peak 52	mg/Kg	NE17_07
	NE013_07	PK053	Peak 53	mg/Kg	NE17_07
	NE013_07	PK054	Peak 54	mg/Kg	NE17_07
	NE013_07	PK055	Peak 55	mg/Kg	NE17_07
	NE013_07	PK056	Peak 56	mg/Kg	NE17_07
	NE013_07	PK057	Peak 57	mg/Kg	NE17_07
	NE013_07	PK058	Peak 58	mg/Kg	NE17_07

Table B-10 Valid values for Baseline Monitoring Program.*Analytical - Fish*

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE013_07	PK059	Peak 59	mg/Kg	NE17_07
	NE013_07	PK060	Peak 60	mg/Kg	NE17_07
	NE013_07	PK061	Peak 61	mg/Kg	NE17_07
	NE013_07	PK062	Peak 62	mg/Kg	NE17_07
	NE013_07	PK063	Peak 63	mg/Kg	NE17_07
	NE013_07	PK064	Peak 64	mg/Kg	NE17_07
	NE013_07	PK065	Peak 65	mg/Kg	NE17_07
	NE013_07	PK066	Peak 66	mg/Kg	NE17_07
	NE013_07	PK067	Peak 67	mg/Kg	NE17_07
	NE013_07	PK068	Peak 68	mg/Kg	NE17_07
	NE013_07	PK069	Peak 69	mg/Kg	NE17_07
	NE013_07	PK070	Peak 70	mg/Kg	NE17_07
	NE013_07	PK071	Peak 71	mg/Kg	NE17_07
	NE013_07	PK072	Peak 72	mg/Kg	NE17_07
	NE013_07	PK073	Peak 73	mg/Kg	NE17_07
	NE013_07	PK074	Peak 74	mg/Kg	NE17_07
	NE013_07	PK075	Peak 75	mg/Kg	NE17_07
	NE013_07	PK076	Peak 76	mg/Kg	NE17_07
	NE013_07	PK077	Peak 77	mg/Kg	NE17_07
	NE013_07	PK078	Peak 78	mg/Kg	NE17_07
	NE013_07	PK079	Peak 79	mg/Kg	NE17_07
	NE013_07	PK080	Peak 80	mg/Kg	NE17_07
	NE013_07	PK081	Peak 81	mg/Kg	NE17_07
	NE013_07	PK082	Peak 82	mg/Kg	NE17_07
	NE013_07	PK083	Peak 83	mg/Kg	NE17_07
	NE013_07	PK084	Peak 84	mg/Kg	NE17_07
	NE013_07	PK085	Peak 85	mg/Kg	NE17_07
	NE013_07	PK086	Peak 86	mg/Kg	NE17_07
	NE013_07	PK087	Peak 87	mg/Kg	NE17_07
	NE013_07	PK088	Peak 88	mg/Kg	NE17_07
	NE013_07	PK089	Peak 89	mg/Kg	NE17_07
	NE013_07	PK090	Peak 90	mg/Kg	NE17_07
	NE013_07	PK091	Peak 91	mg/Kg	NE17_07

Table B-10 Valid values for Baseline Monitoring Program.

Analytical - Fish

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	NE013_07	PK092	Peak 92	mg/Kg	NE17_07
	NE013_07	PK093	Peak 93	mg/Kg	NE17_07
	NE013_07	PK094	Peak 94	mg/Kg	NE17_07
	NE013_07	PK095	Peak 95	mg/Kg	NE17_07
	NE013_07	PK096	Peak 96	mg/Kg	NE17_07
	NE013_07	PK097	Peak 97	mg/Kg	NE17_07
	NE013_07	PK098	Peak 98	mg/Kg	NE17_07
	NE013_07	PK099	Peak 99	mg/Kg	NE17_07
	NE013_07	PK100	Peak 100	mg/Kg	NE17_07
	NE013_07	PK101	Peak 101	mg/Kg	NE17_07
	NE013_07	PK102	Peak 102	mg/Kg	NE17_07
	NE013_07	PK103	Peak 103	mg/Kg	NE17_07
	NE013_07	PK104	Peak 104	mg/Kg	NE17_07
	NE013_07	PK105	Peak 105	mg/Kg	NE17_07
	NE013_07	PK106	Peak 106	mg/Kg	NE17_07
	NE013_07	PK107	Peak 107	mg/Kg	NE17_07
	NE013_07	PK108	Peak 108	mg/Kg	NE17_07
	NE013_07	PK109	Peak 109	mg/Kg	NE17_07
	NE013_07	PK110	Peak 110	mg/Kg	NE17_07
	NE013_07	PK111	Peak 111	mg/Kg	NE17_07
	NE013_07	PK112	Peak 112	mg/Kg	NE17_07
	NE013_07	PK113	Peak 113	mg/Kg	NE17_07
	NE013_07	PK114	Peak 114	mg/Kg	NE17_07
	NE013_07	PK115	Peak 115	mg/Kg	NE17_07
	NE013_07	PK116	Peak 116	mg/Kg	NE17_07
	NE013_07	PK117	Peak 117	mg/Kg	NE17_07
	NE013_07	PK118	Peak 118	mg/Kg	NE17_07
	NE013_07	1336-36-3	Total PCB	mg/Kg	NE17_07
Percent Lipid	NE158_03	LP001	Percent Lipid	%	NE158_03
Mercury	NE025_04	7439-97-6	Mercury	mg/Kg	NE025_04
Dioxins and Furans	EPA 1613B	41903-57-5	Total-TCDD	pg/ug	EPA 1613B
	EPA 1613B	1746-01-6	2,3,7,8-TCDD	pg/ug	EPA 1613B
	EPA 1613B	55722-27-5	Total-TCDF	pg/ug	EPA 1613B

Table B-10 Valid values for Baseline Monitoring Program.

Analytical - Fish

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	EPA 1613B	51207-31-9	2,3,7,8-TCDF	pg/ug	EPA 1613B
	EPA 1613B	36088-22-9	Total PeCDD	pg/ug	EPA 1613B
	EPA 1613B	40321-76-4	1,2,3,7,8-PeCDD	pg/ug	EPA 1613B
	EPA 1613B	30402-15-4	Total PeCDF	pg/ug	EPA 1613B
	EPA 1613B	57117-41-6	1,2,3,7,8-PeCDF	pg/ug	EPA 1613B
	EPA 1613B	57117-31-4	2,3,4,7,8-PeCDF	pg/ug	EPA 1613B
	EPA 1613B	34465-46-8	Total HxCDD	pg/ug	EPA 1613B
	EPA 1613B	39227-28-6	1,2,3,4,7,8-HxCDD	pg/ug	EPA 1613B
	EPA 1613B	57653-85-7	1,2,3,6,7,8-HxCDD	pg/ug	EPA 1613B
	EPA 1613B	19408-74-3	1,2,3,7,8,9-HxCDD	pg/ug	EPA 1613B
	EPA 1613B	55684-94-1	Total HxCDF	pg/ug	EPA 1613B
	EPA 1613B	70648-26-9	1,2,3,4,7,8-HxCDF	pg/ug	EPA 1613B
	EPA 1613B	57117-44-9	1,2,3,6,7,8-HxCDF	pg/ug	EPA 1613B
	EPA 1613B	72918-21-9	1,2,3,7,8,9-HxCDF	pg/ug	EPA 1613B
	EPA 1613B	60851-34-5	2,3,4,6,7,8-HxCDF	pg/ug	EPA 1613B
	EPA 1613B	37871-00-4	Total HpCDD	pg/ug	EPA 1613B
	EPA 1613B	35822-46-9	1,2,3,4,6,7,8-HpCDD	pg/ug	EPA 1613B
	EPA 1613B	38998-75-3	Total HpCDF	pg/ug	EPA 1613B
	EPA 1613B	67562-39-4	1,2,3,4,6,7,8-HpCDF	pg/ug	EPA 1613B
	EPA 1613B	55673-89-7	1,2,3,4,7,8,9-HpCDF	pg/ug	EPA 1613B
	EPA 1613B	3268-87-9	OCDD	pg/ug	EPA 1613B
	EPA 1613B	39001-02-0	OCDF	pg/ug	EPA 1613B
Organochlorine Pesticides	SW846 8081A (NE131_03)	309-00-2	Aldrin	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	319-84-6	alpha-BHC	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	319-85-7	beta-BHC	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	58-89-9	gamma-BHC (Lindane)	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	319-86-8	delta-BHC	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	510-15-6	Chlorobenzilate	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	5103-71-9	alpha-Chlordane	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	5103-74-2	gamma-Chlordane	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	57-74-9	Chlordane	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	96-12-8	DBCP	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	72-54-8	4,4'-DDD	mg/Kg	SW846 3540

Table B-10 Valid values for Baseline Monitoring Program.*Analytical - Fish*

	lab_anl_method_name	cas_rn	chemical_name	result_unit	prep_method
	SW846 8081A (NE131_03)	72-55-9	4,4'-DDE	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	50-29-3	4,4'-DDT	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	2303-16-4	Diallate	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	60-57-1	Dieldrin	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	959-98-8	Endosulfan I	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	33213-65-9	Endosulfan II	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	1031-07-8	Endosulfan sulfate	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	72-20-8	Endrin	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	7421-93-4	Endrin aldehyde	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	53494-70-5	Endrin ketone	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	76-44-8	Heptachlor	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	1024-57-3	Heptachlor epoxide	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	118-74-1	Hexachlorobenzene	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	77-47-4	Hexachlorocyclopentadiene	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	465-73-6	Isodrin	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	72-43-5	Methoxychlor	mg/Kg	SW846 3540
	SW846 8081A (NE131_03)	8001-35-2	Toxaphene	mg/Kg	SW846 3540

Table B-10 Valid values for Baseline Monitoring Program.

Surrogates

cas_no	chemical_name	method	prep_method	reporting_unit
53742-07-7	Nonachlorobiphenyl	NE207_03	NE005_04	%
877-09-8	Tetrachloro-meta-xylene	SW846 8082 (NE148_04)	NE17_07	%
S2051-24-3	Decachlorobiphenyl	SW846 8082 (NE148_04)	NE17_07	%
877-09-8	Tetrachloro-meta-xylene	NE013_07	NE17_07	%
S2051-24-3	Decachlorobiphenyl	NE013_07	NE17_07	%
53742-07-7	Nonachlorobiphenyl	NE013_07	NE17_07	%

Other analytical

total_or_dissolved	test_type	result_type_code	reportable_result	detect_flag	lab_qualifiers
T	INITIAL	TRG	YES	Y	U
D	REANALYSIS	SUR	NO	N	U*
N	REEXTRACT				J
					UJ
					R
					UR
					<

organic_yn	lab_matrix_code	analysis_location	basis	data_package_level	sample_type_code	sample_source
Y	S	FI	WET	A	ENV	Lab
N	W	FL	DRY	B	FDBL	Field
	F	LB	NA		DUP	
					LR	
					LCS	
					MS	
					MB	
					HRM	

Table B-10 Valid values for Baseline Monitoring Program.

Field Values - water

sample_matrix_code	sample_type_code	sample_source	test_requested	sample_type	volume
S	ENV	Field	NE207_03	CENTER CHANNEL	ROUTINE
W	FDBL		EPA 353.3	E-W COMPOSITE	HIGH
	DUP		EPA 354.1	TRANSECT COMPOSITE	
			EPA 351.1		
			EPA 365.2		
			EPA 200.8		
			EPA 245.1		
			EPA 160.2		
			NE128_03		
			EPA 1613B		

Field Values - fish

sample_matrix_code	sample_type_code	sample_source	test_requested	sex	sample_prep
F	ENV	Field	SW846 8082 (NE148_04)	M	fillet
	HRM		NE013_07	F	whole body
			NE158_03	U	
			NE025_04		
			EPA 1613B		
			SW846 8081A (NE131_03)		

Table D-1. Format of an Environmental Standards quality assurance review.

**GENERAL ELECTRIC COMPANY
HUDSON RIVER BASELINE MONITORING PROGRAM**

TITLE PAGE

TABLE OF CONTENTS

INTRODUCTION AND SAMPLE LISTING

SECTION 1

1. Introduction

The introduction section will briefly state the number of samples analyzed, the laboratory that analyzed them, the parameters that were analyzed for, and the methods used for analysis.

2. Laboratory Compliance

This section will specify any correctable and/or noncorrectable deficiencies and will make informative comments about issues that were identified relative to the organic, inorganic, and general chemistry requirements specified in the analytical SOPs. Appropriate EPA citations or project citations will be provided for each item listed. This section will also specify discrepancies between the reported data and the raw data.

3. Data Qualifiers

This section will present qualifiers that should be considered for the data to best be utilized. A detailed assessment of the degree to which data have been compromised by any deviation from protocol (i.e., lack of analytical control and QC failure) will be included. For every statement made in this section, there will be a subsequent finding that justifies the qualifying statement. These qualifiers/findings will be presented as bulleted items, in order of importance, relative to their impact on the data set. The data qualifiers will be presented in two subsections; organic data and inorganic and general chemistry data. The qualifiers will be presented in the order of greatest impact to least impact within each subsection.

SECTION 2

This section will include the qualified sample result summaries and a glossary defining the qualifier codes. These qualified spreadsheets will be presented in the following order: volatiles, semivolatiles, pesticides, PCBs, herbicides, metals, and general chemistry parameters.

SECTION 3

The organic quality assurance review is fully supported by a documentation appendix. For every qualifier made in the report, there is a photocopied page of laboratory data that is used in support of the reviewer's comments. All QC summary forms as well as the reviewer's worksheets are presented in the support documentation.

SECTION 4

The inorganic and general chemistry quality assurance review is also fully supported by a documentation appendix in the same format as the organic data. All QC summary forms as well as the reviewer's worksheets are presented in the support documentation.

SECTION 5

This section of the quality assurance review will contain the laboratory case narratives and the field and laboratory Chain-of-Custody Records.

SECTION 6

This section of the quality assurance review will contain any applicable project correspondence.