



Linkage of Exposure and Effects Using Genomics, Proteomics, and Metabolomics in Small Fish Models

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research development

Project IA-1

Agency Needs

- Improved understanding of the linkages in the continuum between the source of a chemical in the environment and adverse outcomes
- Predictive models for screening and testing
- Improvements in quantitative risk assessment

Research Goals

- Identify new molecular markers of effects of exposure to chemicals representing different mechanisms of action (MOA) within the hypothalamic-pituitary-gonadal (HPG) axis of small fish models
- Link these biomarkers to responses that are relevant to ecological risk assessments
- Support development of integrated modeling approaches that utilize MOA as a basis for prediction of adverse outcomes

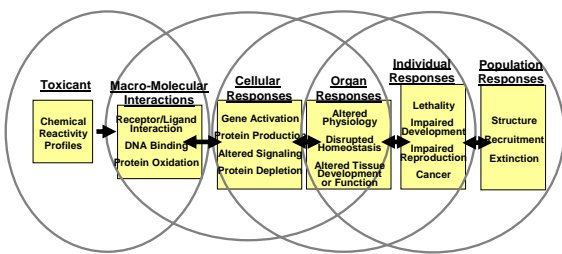
Relevance to Client Offices

- OSCP – Further validation of fathead minnow assay for testing endocrine disrupting chemicals. Impact on international programs for testing EDCs via relationship between OSCP and OECD.
- OPPTS – a) Testing pesticides of direct interest to program office with respect to human health and/or ecological effects; b) helps develop linkages between mechanism-specific data and population level effects.
- Contribution to EPA programs involved in monitoring and/or diagnostic (retrospective) risk assessments [e.g. NPDES program through OW and EPA Regions].

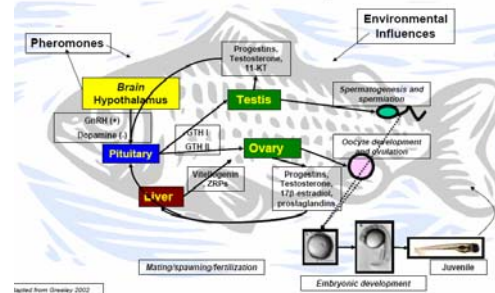
Specific Approach

- Investigate effects of chemicals on the HPG axis using definitive tests in the fathead minnow to establish initial toxicity pathways and ecologically-relevant responses (Phase 1)
- Identify the genomic, proteomic, metabolomic, and other whole-animal responses to the same chemicals using short-term zebrafish exposures (Phase 2)
- Validate zebrafish molecular markers in the fathead minnow using extrapolation of relevant gene and protein markers with a subset of definitive endpoint tests and metabolite profiles for the HPG-active chemicals (Phase 3)
- Concurrently integrate data from above in a systems modeling context, as well as relevant population modeling

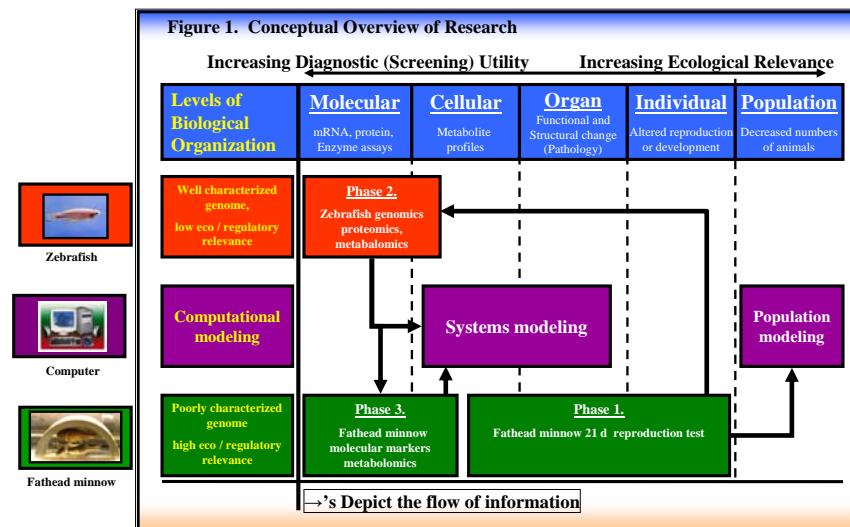
Understanding Toxicity Pathways



Overview of Fish Reproduction



Methods/Approach

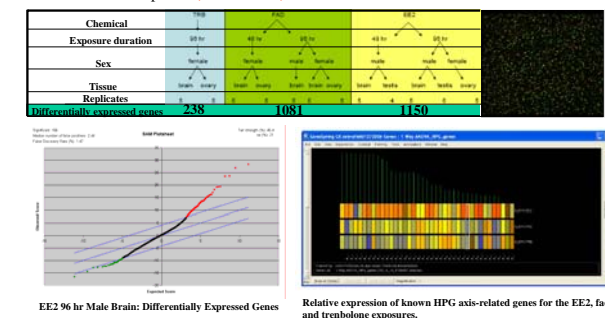


Progress to Date

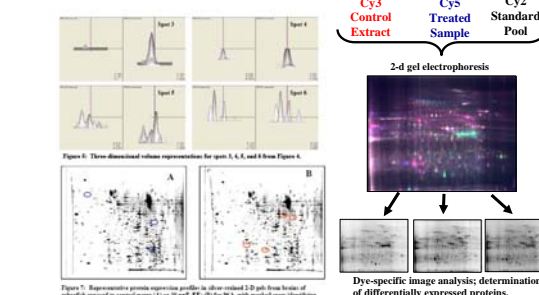
Phase 1 (definitive fathead minnow) assays completed with ketoconazole, fadrozole, prochloraz, flutamide, trenbolone, haloperidol, vinclozolin, apomorphine (preliminary). (EPA-MED)

Phase 2 (short-term zebrafish) assays completed with trenbolone, fadrozole, ethynyl estradiol (EE2), flutamide, and fipronil. (EPA-MED)

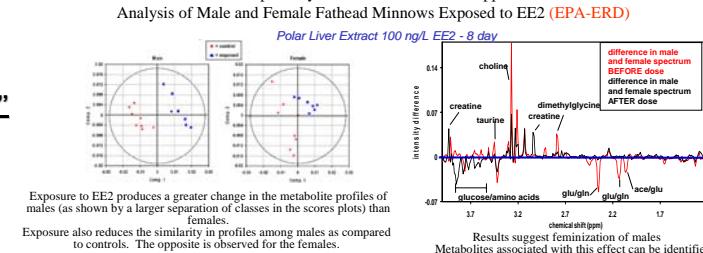
Zebrafish array work completed (Icoria) for EE2, fadrozole, trenbolone, and in progress for flutamide and fipronil (EPA-EERD).



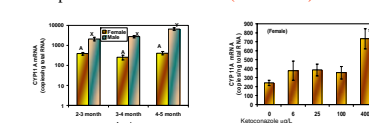
Effects on the proteome of selected zebrafish tissues are being evaluated by Ettan™ DIGE 3-dye system (EPA-EERD)



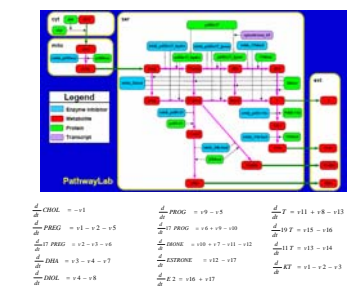
Proof-of-Concept Study for Eco-metabolomics Applied to EDCs: Analysis of Male and Female Fathead Minnows Exposed to EE2 (EPA-ERD)



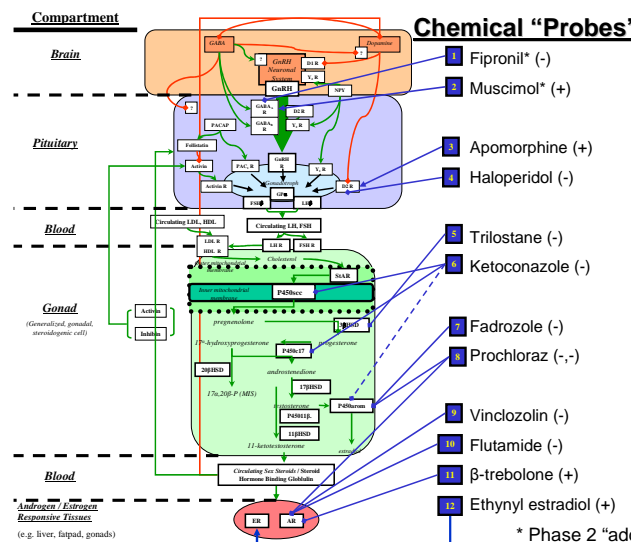
Over a dozen fathead minnow genes sequenced and quantitative RT-PCR assays developed for > 20 key HPG axis genes. Transcript levels measured under "baseline" conditions and after exposure to test chemicals. (EPA-MED)



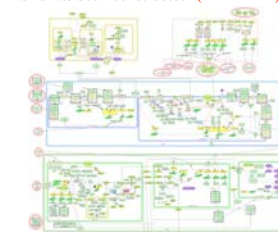
Deterministic computational model of vertebrate steroidogenesis (EPA-NCCT)



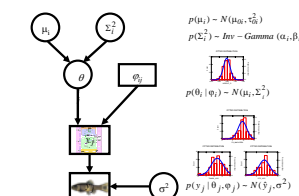
Embracing Collaboration



SBML compatible graphical systems model of the generalized teleost HPG axis has been constructed (EPA-MED)



Physiologically-based computational systems model being calibrated with fathead minnow control data (OHSU)



Population model linking from a biomarker, plasma Vtg levels, to predicted population outcomes. (EPA-MED Grosse Ile)

