

## 4. METAL-SPECIFIC TOPICS AND METHODS

This chapter discusses metal-specific topics and methods to be used in the assessment of risk to humans and ecological entities from exposures to inorganic metals. It applies information and text from the metals issue papers and reflects contributions by EPA scientists and external experts. The final metals issue papers are available on the EPA Web site at <http://cfpub.epa.gov/ncea/raf/recordisplay.cfm?deid=86119>.

Key topics and tools in this section are presented in subsections on environmental chemistry, exposure pathway analysis, human health effects, and ecological effects. The applications and limitations of the various models and methods for conducting metals assessments are presented to inform the reader. Topics and tools related to bioavailability and bioaccumulation are discussed throughout Chapter 4 because they have far reaching impact that crosses many aspects of metals assessment.

### 4.1. ENVIRONMENTAL CHEMISTRY

#### 4.1.1. Introduction and Terminology

A general review of factors pertaining to the chemistry of metals in sediments, soils, waters, and the atmosphere is presented in this chapter in the context of risk assessment. Because the behavior of metals defies simple generalities, it is necessary to understand the chemistry of the particular metal and the environment of concern. However, we can generalize factors that control metal chemistry and environmental characteristics where this generalization allows us to progress with estimates of metal fate and effects.

Metal speciation determines the behavior and toxicity of metals in the environment. Speciation refers to the occurrence of a metal in a variety of chemical forms. These forms may include free metal ions, metal complexes dissolved in solution and sorbed on solid surfaces, and metal species that have been coprecipitated in major metal solids or that occur in their own solids. The speciation of a metal affects not only its toxicity but also its volatilization, photolysis, sorption, atmospheric deposition, acid/base equilibria, polymerization, complexation, electron-transfer reactions, solubility and precipitation equilibria, microbial transformations, and diffusivity (Bodek et al., 1988).

The following sections address the application of hard and soft acid and base (HSAB) concepts to metal behavior, including the formation of metal complexes, and the importance of pH and oxidation-reduction reactions to metal mobility and toxicity. The chapter then examines

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1 the occurrence and interactions of the metals of concern in natural media, including in surface  
2 and ground waters, soils and aquatic sediments, and the atmosphere. Important topics  
3 considered in these sections are metal sorption behavior, aging in soils, metal dissolution and  
4 transformation and transfer to plants, and methods of determining metal speciation in soils and  
5 sediments.

#### 7 **4.1.2. Hard and Soft Acids and Bases: The Stability of Complexes**

8         Complexes are formed between metals (acids) and  
9 ligands (bases), both in solution and at the surfaces  
10 minerals and organisms. The toxic reaction of organisms  
11 to metals can be directly related to the nature of the metal  
12 complexes formed in solution and at the surface of the  
13 organism.

14         A useful concept that helps to explain the strength  
15 of metal complexing and metal toxicity is that of hard and  
16 soft acids and bases (HSAB), which was introduced by  
17 Pearson (1973). In this concept, metal cations are Lewis  
18 acids and ligands are Lewis bases, with the metal cation  
19 and ligand in a complex acting as electron acceptor and  
20 donor, respectively. “Soft” implies that the species’

21 electron cloud is deformable or polarizable and the electrons are mobile and easily moved. Soft  
22 species prefer to participate in covalent bonding. Hard species are comparatively rigid and  
23 nondeformable, have low polarizability, hold their electrons firmly, and prefer to participate in  
24 ionic bonds in complex formation (Langmuir, 1997). Hard acids form strong, chiefly ionic  
25 bonds with hard bases, whereas soft acids and soft bases form strong, chiefly covalent bonds  
26 when they form complexes. In contrast, the bonds formed  
27 between hard-soft or soft-hard acids and bases are weak,  
28 such that their complexes tend to be rare. Table 4-1  
29 summarizes hard and soft acid and base relationships for  
30 the metals of concern. The first text box summarizes the  
31 applicability of hard and soft concepts to the formation of  
32 metal complexes; the second text box defines ligands.

##### **Hard and Soft Acids and Bases**

###### ***Hard acids and hard bases.***

Complexes formed between divalent hard acid cations and monovalent or divalent hard bases are ionic and relatively weak and are often termed “ion pairs.” Complexes formed between Be<sup>2+</sup> or trivalent hard acids and hard bases tend to be ionic and relatively strong.

***Soft acids and soft bases.*** Strong, relatively covalent bonds are formed in complexes between soft and borderline soft acid cations and soft bases. Ligand binding sites on the external or internal surfaces of organisms are often of soft base character and thus bond strongly with soft and borderline soft acid cations.

##### **Ligands**

***Ligands*** are anions or molecules that form complexes with metal ions. Depending on whether a ligand shares one, two, three or more electron pairs with metals, it is called a mono-, bi-, tri- or multidentate ligand.

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1 *Hard metals (hard acids), which are the least toxic, preferentially bind with hard bases*  
 2 *that contain oxygen, forming weaker bonds with soft nitrogen and sulphur species. The strength*  
 3 *of binding between hard metals and hard ligands is usually a function of pH. Many of the hard*  
 4 *metals are macronutrients. Soft metals (acids) bind preferentially with soft S and N ligands,*  
 5 *forming weaker bonds with hard base species such as hydroxide and sulfate. Soft and*  
 6 *borderline metals, and Mn<sup>2+</sup>, which is hard, form bonds of decreasing strength with soft ligands*  
 7 *such as*

**Table 4-1. Hard and soft acids (metal cations) and bases (ligands)**

Hard acids	Al <sup>3+</sup> , Ba <sup>2+</sup> , Be <sup>2+</sup> , Co <sup>3+</sup> , Cr <sup>3+</sup> , Fe <sup>3+</sup> , Mn <sup>2+</sup> , Sr <sup>2+</sup> , U <sup>4+</sup> , UO <sub>2</sub> <sup>2+</sup> , VO <sup>2+</sup>
Borderline acids (between hard and soft)	Co <sup>2+</sup> , Cu <sup>2+</sup> , Fe <sup>2+</sup> , Ni <sup>2+</sup> , Pb <sup>2+</sup> , Zn <sup>2+</sup>
Soft acids	Ag <sup>+</sup> , Cd <sup>2+</sup> , Cu <sup>+</sup> , Hg <sup>2+</sup> , Hg <sup>+</sup> , CH <sub>3</sub> Hg <sup>+</sup> , Tl <sup>3+</sup> , Tl <sup>+</sup>
Hard bases	F <sup>-</sup> , H <sub>2</sub> O, oxyanions: OH <sup>-</sup> , SO <sub>4</sub> <sup>2-</sup> , CO <sub>3</sub> <sup>2-</sup> , HCO <sub>3</sub> <sup>-</sup> , C <sub>2</sub> O <sub>4</sub> <sup>2-</sup> , CrO <sub>4</sub> <sup>2-</sup> , MoO <sub>4</sub> <sup>2-</sup> , H <sub>n</sub> PO <sub>4</sub> <sup>n-3</sup> , H <sub>n</sub> AsO <sub>4</sub> <sup>n-3</sup> , SeO <sub>4</sub> <sup>2-</sup> , H <sub>2</sub> VO <sub>4</sub> <sup>-</sup> , NH <sub>3</sub> , RNH <sub>2</sub> , N <sub>2</sub> H <sub>4</sub> , ROH, RO <sup>-</sup> , R <sub>2</sub> O, CH <sub>3</sub> COO <sup>-</sup> , etc.
Borderline bases (between hard and soft)	Cl <sup>-</sup> , Br <sup>-</sup> , NO <sub>2</sub> <sup>-</sup> , SO <sub>3</sub> <sup>2-</sup> , H <sub>n</sub> AsO <sub>3</sub> <sup>n-3</sup> , C <sub>6</sub> H <sub>5</sub> NH <sub>2</sub> , C <sub>5</sub> H <sub>5</sub> N, N <sub>3</sub> <sup>-</sup> , N <sub>2</sub>
Soft bases	I <sup>-</sup> , HS <sup>-</sup> , S <sup>2-</sup> , CN <sup>-</sup> , SCN <sup>-</sup> , Se <sup>2-</sup> , S <sub>2</sub> O <sub>3</sub> <sup>2-</sup> , -SH, -SCH <sub>3</sub> , -NH <sub>2</sub> , R <sup>-</sup> , C <sub>2</sub> H <sub>4</sub> , C <sub>6</sub> H <sub>6</sub> , RNC, CO, R <sub>3</sub> P, (RO) <sub>3</sub> P, R <sub>3</sub> As, R <sub>3</sub> S, RSH, RS <sup>-</sup>

Source: Modified after Huheey et al. (1993) and Langmuir (1997). "R" refers to an organic molecule.

1 sulfide, generally in the following order: Pb<sup>2+</sup> > Cu<sup>2+</sup> > Cd<sup>2+</sup> > Co<sup>2+</sup> ≈ Fe<sup>2+</sup> > Ni<sup>2+</sup> > Zn<sup>2+</sup> > Mn<sup>2+</sup>.  
 2 The tendency of metals to bind to soft ligands or to organic substrates (which are usually soft) is  
 3 greatest for soft and borderline metals (soft acids), followed by the hard metals (hard acids),  
 4 typically in the order Pb<sup>2+</sup> > Cu<sup>2+</sup> > Cd<sup>2+</sup> > Zn<sup>2+</sup> > Ca<sup>2+</sup> > Mg<sup>2+</sup> >> Na<sup>+</sup> (Pickering, 1986).

5 The tendency of metals to form solid phases, such as sulfides in sediments, is also related  
 6 to their HSAB qualities. For example, extremely insoluble metal sulfides are formed in anoxic  
 7 sediments by soft acid metal cations, such as Hg<sup>2+</sup> (log K<sub>sp</sub> = -57.25) or Ag<sup>+</sup> (log K<sub>sp</sub> = -49.7),  
 8 whereas borderline hard and hard metal cations such as Mn<sup>2+</sup> (log K<sub>sp</sub> = -19.25) or Fe<sup>2+</sup> (log K<sub>sp</sub>  
 9 = -22.39) form slightly more soluble, although still highly insoluble, metal sulfides.<sup>1</sup> These

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<sup>1</sup>Solubility products for all sulfides except Ag<sub>2</sub>S are from Di Toro et al. (1990). The product for silver sulfide is from Stumm and Morgan (1970).

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1 differences in solubilities are fundamental to the method of acid-volatile sulfide (AVS)  
2 normalization of sediment-associated metals (see Section 4.1.5, on Sediment Chemistry).  
3 Finally, according to the Biotic Ligand Model (BLM) (see Sections 3.1.4.2 and 3.4), effects of  
4 metals are related to or correlated with metal interactions with biological ligands, which are  
5 generally soft base species.

### 7 **4.1.3. Aquatic Chemistry**

#### 8 **4.1.3.1. Speciation and Complexes**

9 Metal species dissolved in water may occur as free ions, or aquo-ions, or as complexes.  
10 Free metal cations are generally surrounded by coordinating water molecules and so have been  
11 termed aquocations, although by convention the water molecules are ignored when writing  
12 chemical reactions involving metal cations.

13 The total analytical concentration of a given metal in water is the sum of the  
14 concentrations of its free ion and its complexes and any metal associated with suspended solids,  
15 whether organic or mineral. For example, the total molal concentration of lead,  $\Sigma\text{Pb}$ , in a natural  
16 water might equal:

$$18 \quad \Sigma\text{Pb} = m\text{Pb}^{2+} + m\text{PbOH}^+ + m\text{PbCO}_3^0 + m\text{PbHCO}_3^+ + m\text{PbSO}_4^0 + m\text{Pb}(\text{suspended solids})$$

19  
20 In most natural waters, the concentration of free lead ion,  $m\text{Pb}^{2+}$ , is less than the sum of  
21 the concentrations of its complexes, which in this case are lead complexes with hydroxyl,  
22 carbonate, bicarbonate, and sulfate ions. Other metals that are found in natural waters most often  
23 as complexes and not as free ions include  $\text{Al}^{3+}$ ,  $\text{Ag}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ , and  $\text{Hg}^{2+}$ . The metalloids As and  
24 Se and the metals Cr, Mo, Sb, and V occur most often in aerobic waters and soils as covalently  
25 bonded to oxygen in oxyanions. Under oxidizing conditions, these include arsenate, selenate,  
26 chromate, molybdate, and vanadate, which themselves are complexes.

27 Complexes that incorporate metals play a major role in controlling the availability and  
28 fate of metals in the environment. Increasing the fraction of a metal that is complexed increases  
29 the solubility of minerals of that metal (Langmuir, 1997). For example, the solubility of lead  
30 sulfate is related to the molal concentrations of free lead and sulfate ions through the expression:

$$31 \quad K_{\text{sp}} = (\gamma_{\text{Pb}} \cdot m\text{Pb}^{2+})(\gamma_{\text{SO}_4} \cdot m\text{SO}_4^{2-})$$

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1 where the terms  $\gamma_{\text{Pb}}$  and  $\gamma_{\text{SO}_4}$  are the activity coefficients of the ions. The product of the ion  
2 activity coefficient and the molal concentration of each species equals the activity of the ion.  
3 This equation shows that the activity of free lead ion controls the solubility of lead sulfate. For a  
4 given total lead concentration (see the previous equation), the more of the lead that is  
5 complexed, the lower will be the concentration of free lead ion. This means that as the extent of  
6 lead complexing increases, the total lead concentration must also increase to reach saturation  
7 with lead sulfate. In other words, metal complexing increases total metal solubility.

8 Metal complexing also has a direct influence on metal adsorption to organic matter or  
9 mineral surfaces. For example, metal carbonate, sulfate, and fluoride complexes are usually  
10 poorly adsorbed, whereas metal hydroxide complexes are strongly adsorbed (Langmuir, 1997).  
11 In summary, metal complexing generally increases the solubility and mobility of metals in  
12 surface and ground waters.

13 For many metals, the free metal ion is thought to be the primary metal species that causes  
14 toxicity to aquatic organisms. This is consistent with the free ion activity model (FIAM), which  
15 assumes that the free or aquo-ion is the most biologically active form of the dissolved metal.  
16 Accordingly, the key parameters that can modify the degree of toxicity are those that affect  
17 speciation, such as pH and the amount of inorganic and organic ligands (e.g., dissolved organic  
18 carbon, DOC) that can form metal complexes and so provide alternative binding sites for the  
19 metal ion. Metal toxicity is also affected by other dissolved ions (e.g., Na, Ca) that compete with  
20 metals for binding sites on the gills of fish or on respiratory surfaces of other aquatic organisms.

#### 21 22 **4.1.3.2. Importance of pH and Redox Conditions**

23 The pH is probably the single most important variable that influences the behavior of  
24 metals in the environment. Thus, metal complexes with sulfate, fluoride, chloride, and  
25 phosphate are most stable and important below pH 7, whereas metal carbonate and hydroxide  
26 complexes become increasingly more important above pH 6–8.

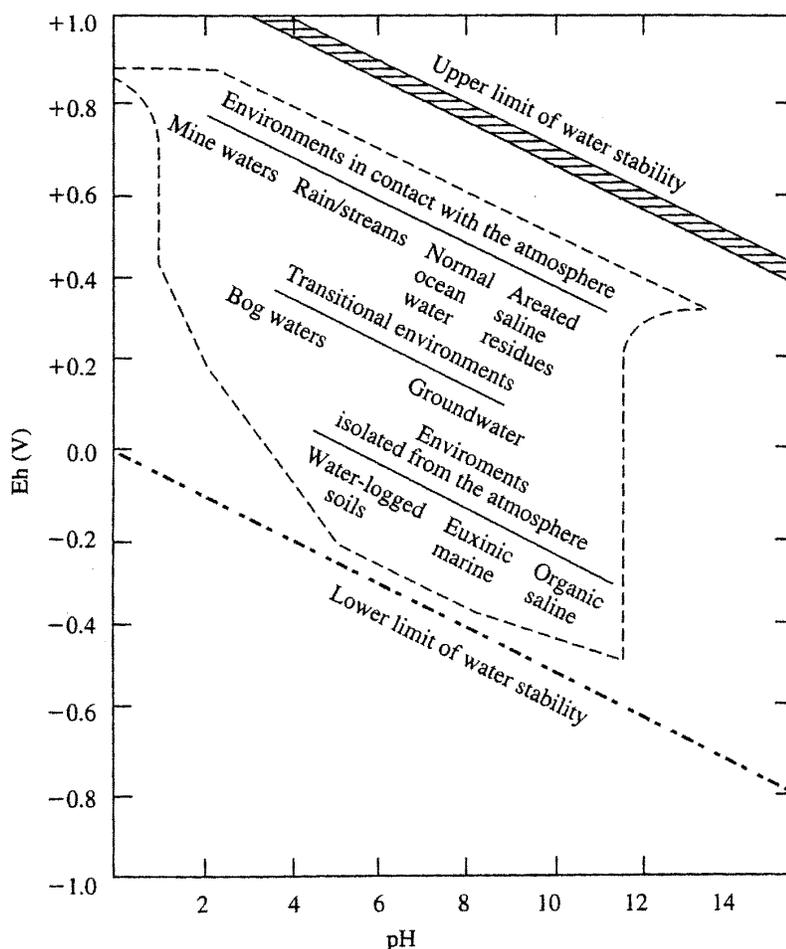
27 Also, as discussed in Section 4.1.4, hydrogen ion competes with metal cations for  
28 adsorption sites, so that adsorption of metal cations by hydrous ferric oxide (HFO), for example,  
29 is low in acid systems but increases with increasing pH. In contrast, oxyanions of As, Mo, Se,  
30 and Cr tend to be desorbed from HFO with increasing pH because of competition between the  
31 oxyanions and  $\text{OH}^-$  ion for sorption sites. Furthermore, the solubility of most metal-containing  
32 minerals is greatest under acid conditions and decreases with increasing pH.

33 Figure 4-1 shows the locus of measured values of oxidation potential (Eh) and pH in  
34 aquatic systems. The principal controls on Eh are atmospheric oxygen and organic matter. High  
35 Eh (oxidizing or aerobic) conditions are maintained in the atmosphere and in most surface waters

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1 and shallow soils in contact with atmospheric oxygen. The lowest Eh values and reducing or  
 2 anaerobic conditions are found in water-logged soils and sediments that contain organic matter,  
 3 and in ground waters that contain a few milligrams per liter or more of dissolved organic carbon  
 4 (DOC). Intermediate Eh conditions are found in waters and sediments that are only partially  
 5 oxidized because of their relative isolation from the atmosphere. Measured Eh values may not  
 6 agree with Eh values computed from the concentrations of redox-sensitive species. The  
 7 difference between measured and computed Eh values is discussed at length by Stumm and  
 8 Morgan (1996) and Langmuir (1997).  
 9

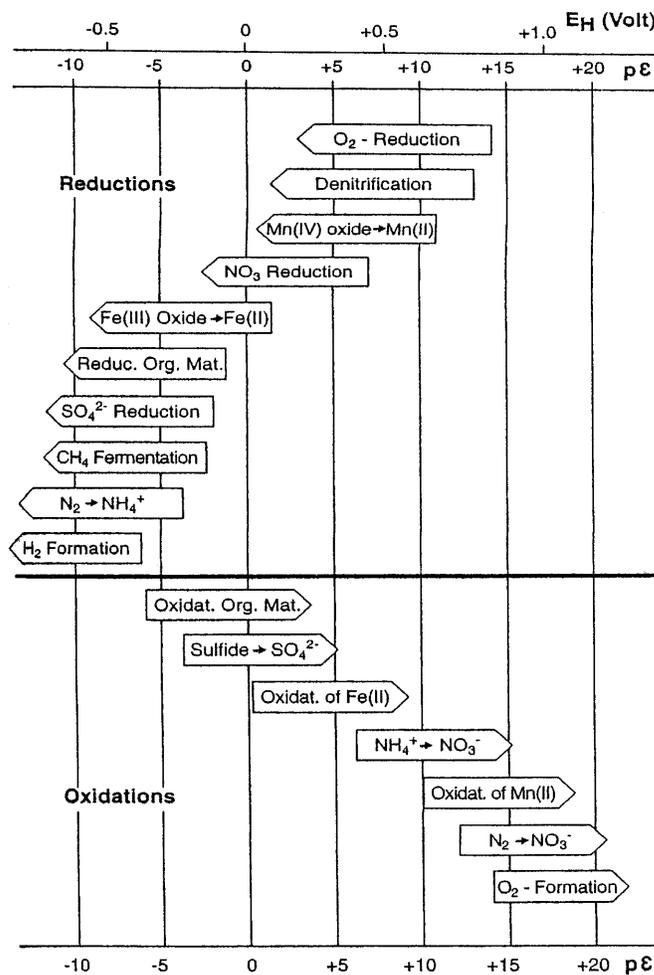


**Figure 4-1. Approximate positions of some natural environments in terms of Eh and pH.** The dashed line represents the limits of measurements in natural environments, as reported by Baas-Becking et al. (1960).

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Microorganisms play an essential role in defining the redox conditions in aquatic systems. The effect of the common sequence of microbially mediated redox reactions on Eh when conditions in a water or sediment become more reducing (depleted in oxygen) or oxidizing is shown in Figure 4-2. These reactions importantly affect or are affected by the redox behavior of the major elements C, S, N, Fe, and Mn and also affect the mobility of most of the metals of concern, which have multiple redox states (Sb, As, Cr, Co, Cu, Mn, Hg, Mo, Ni, Se, and Ag). Thus, Cr(VI) (oxidized) in chromate ion is highly mobile in aqueous environments compared to Cr(III) (reduced). Cr(VI) is considered a known human inhalation carcinogen, whereas Cr(III) is generally considered to have low human toxicity. Among the microbially mediated reactions in Figure 4-2, the one that most affects possible metal toxicity is sulfate reduction, which requires the presence of organic matter. Sulfate reduction produces hydrogen sulfide, which reacts with



**Figure 4-2. Sequence of microbially mediated oxidation-reduction reactions.**

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1 metals such as Sb, As, Cd, Co, Cu, Pb, Hg, Mo, Ni, and Ag and can cause their almost  
2 quantitative precipitation as insoluble metal sulfides. This immobilizes these metals and so  
3 makes them unavailable to plants and animals.

4 Much of the preceding discussion of the importance of metal speciation, hard and soft  
5 acids and bases, oxidation potential, and metal sulfides, is summarized in Table 4-2. The table  
6 shows, for example, that the hard acid metals Al, Ba, Be, and Sr have but one oxidation state and  
7 do not form insoluble sulfides; thus, their solubilities and mobilities are not directly affected by  
8 redox conditions, although they are strongly affected by changes in pH. Also indicated are  
9 forms of the metals that occur in soils and waters, with As, Mo, Se, and sometimes Cr and V  
10 occurring as oxyanions, and most of the other metals occurring usually as metal cations. Eh-pH  
11 diagrams for the redox-sensitive metals of concern and of major elements such as Fe, S, and C  
12 are given in Langmuir et al. (2004). These show the detailed occurrence of the metal species  
13 under oxidizing and reducing conditions. In simplified terms, the Eh-pH diagram and Table 4-2  
14 show that Fe, Mn, and Tl are most mobile under reducing conditions, whereas S and the  
15 remaining metals of concern are usually most mobile under oxidizing conditions.  
16

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**Table 4-2. Important properties of trace metals as they occur in natural waters and mineral systems: oxidation states, redox sensitivity, tendency to form sulfides at low Eh, dominant chemical species of metals in soils and natural waters not considering most (especially weak) metal complexes**

Metal	Symbol	Hard or soft	Oxidation states	Oxidized forms	Reduced forms <sup>a</sup>	Can form sulfides at low Eh	Soils	Waters
Aluminum	Al	H	3+	Al <sup>3+</sup>	Al <sup>3+</sup>	no		
Antimony	Sb	H/S	5+, 3+, 0	Sb(OH) <sub>6</sub> <sup>-</sup>	Sb(OH) <sub>3</sub> <sup>0</sup> /Sb(s)	yes	Sb <sub>III</sub> O <sub>x</sub> ?	Sb(OH) <sub>6</sub> <sup>-</sup>
Arsenic	As	H/S <sup>b</sup>	5+, 3+, 0	HAsO <sub>4</sub> <sup>2-</sup>	H <sub>3</sub> AsO <sub>3</sub> <sup>0</sup> /As(s)	yes	AsO <sub>4</sub> <sup>3-</sup>	AsO <sub>4</sub> <sup>3-</sup> , AsO <sub>3</sub> <sup>3-</sup>
Barium	Ba	H	2+	Ba <sup>2+</sup>	Ba <sup>2+</sup>	no	Ba <sup>2+</sup>	Ba <sup>2+</sup>
Beryllium	Be	H	2+	Be <sup>2+</sup>	Be <sup>2+</sup>	no	Be <sup>2+</sup> , Be <sub>x</sub> O <sub>y</sub> <sup>2x-2y</sup>	Be <sup>2+</sup>
Cadmium	Cd	S	2+	Cd <sup>2+</sup>	Cd <sup>2+</sup>	yes	Cd <sup>2+</sup>	Cd <sup>2+</sup>
Chromium	Cr	H	6+, 3+	CrO <sub>4</sub> <sup>2-</sup>	Cr <sup>3+</sup> , Cr(OH) <sub>3</sub> (s)	no	Cr <sup>3+</sup>	CrO <sub>4</sub> <sup>2-</sup> , Cr <sup>3+</sup>
Cobalt	Co	B	(3+), 2+	(Co <sup>3+</sup> ), Co <sup>2+</sup>	Co <sup>2+</sup>	yes	Co <sup>2+</sup>	Co <sup>2+</sup>
Copper	Cu <sup>a</sup>	B/S <sup>b</sup>	2+, 1+, 0	Cu <sup>2+</sup>	Cu <sup>+</sup> /Cu(s)	yes	Cu <sup>2+</sup> , Cu-OM <sup>b</sup>	Cu <sup>2+</sup> , -fulvate
Lead	Pb	B	(4+), 2+	Pb <sup>2+</sup>	Pb <sup>2+</sup>	yes	Pb <sup>2+</sup>	Pb(OH) <sup>+</sup>
Manganese	Mn	H	(4+), (3+), 2+	(4+), 2+	(3+), 2+	no <sup>b</sup>	Mn <sup>4+</sup> , Mn <sup>2+</sup>	Mn <sup>2+</sup>
Mercury	Hg	S	2+, 1+, 0	Hg <sup>2+</sup>	Hg <sup>+</sup> /Hg(l)	yes	Hg <sup>2+</sup> , CH <sub>3</sub> Hg	Hg(OH) <sub>2</sub> <sup>0</sup> , HgCl <sub>2</sub> <sup>0</sup>
Molybdenum	Mo	H/S	6+, (5.33+), 5+, (4+)	HMoO <sub>4</sub> <sup>-</sup>	MoO <sub>2</sub> <sup>+</sup> /Mo <sub>3</sub> O <sub>8</sub> (s)/MoS <sub>2</sub> (s)	yes	MoO <sub>4</sub> <sup>2-</sup>	MoO <sub>4</sub> <sup>2-</sup>
Nickel	Ni	B	(3+), 2+	(Ni <sup>3+</sup> ), Ni <sup>2+</sup>	Ni <sup>2+</sup>	yes	Ni <sup>2+</sup>	Ni <sup>2+</sup>
Selenium	Se	H/S	6+, 4+, (0), 2-	SeO <sub>4</sub> <sup>2-</sup>	SeO <sub>3</sub> <sup>2-</sup> /Se(s)/HSe <sup>-</sup>	no	SeO <sub>4</sub> <sup>2-</sup> , HSeO <sub>3</sub>	SeO <sub>4</sub> <sup>2-</sup>
Silver	Ag	S	1+, (0)	Ag <sup>+</sup>	Ag <sup>+</sup> /Ag(s)	yes	Ag <sup>+</sup>	Ag <sup>+</sup> , AgCl
Strontium	Sr	H	2+	Sr <sup>2+</sup>	Sr <sup>2+</sup>	no		
Thallium	Tl	S	(4+), (3+), 1+	(Tl <sup>4+</sup> ), (Tl <sup>3+</sup> )	Tl <sub>2</sub> O(s)/Tl <sup>+</sup>	yes		
Vanadium	V	H	5+, 4+, 3+	H <sub>2</sub> VO <sub>4</sub> <sup>-</sup>	VO <sup>2+</sup> , V(OH) <sub>3</sub> <sup>0</sup>	no	V <sub>IV</sub> O <sub>x</sub> ?	HVO <sub>4</sub> <sup>2-</sup>
Zinc	Zn	B	2+	Zn <sup>2+</sup>	Zn <sup>2+</sup>	yes	Zn <sup>2+</sup>	Zn <sup>2+</sup>

Oxidation states in parentheses are found in mineral systems only. Hard (H), soft (S), and borderline soft (B) metals are indicated.

<sup>a</sup>Cu<sup>2+</sup> is borderline soft, and Cu<sup>+</sup> is a soft cation. Typically, much or most of Cu in soils is complexed with organic matter.

<sup>b</sup>B/S or H/S denotes whether the oxidized species is borderline hard or hard, and the reduced species is soft.

<sup>c</sup>Cu-OM denotes copper complexed with organic matter.

Source: Modified after Logan and Traina, 1993.

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1 **4.1.3.3. *Effects Concentration and Bioavailability***

2 The traditional method of predicting effects of metals on aquatic organisms is to estimate  
3 an effects concentration for the water column and the sediment. For the water column, the  
4 EPA's Water Quality Criteria (WQC) (U.S. EPA, 1996a; Stephen et al., 1985) or the Predicted  
5 No-Effect Concentration (PNEC) derived following the European Union's (EU's) technical  
6 guidelines (EU, 1996) are frequently used as default screening values. However, these criteria  
7 make only limited corrections for bioavailability, taking into account only water hardness and  
8 ignoring other modifying factors such as pH and competing ligands. The BLM was developed to  
9 remedy this situation (Paquin et al., 2002a; U.S. EPA, 2000d; Di Toro et al., 2000). It  
10 incorporates the WHAM speciation model and it also models the competitive metal binding at  
11 the toxic site of action (the biotic ligand). BLMs are currently available for copper and silver  
12 (Di Toro et al., 2000; Santore et al., 2000) and zinc (Santore et al., 2002) and are under  
13 development for cadmium, nickel, and lead (see [www.epa.gov/waterscience](http://www.epa.gov/waterscience)). The development  
14 and use of the BLM is covered in detail in Section 4.5, Characterization of Ecological Effects.  
15

16 **4.1.4. Ground Water and Metals Mobility**

17 Site-specific risk assessments for EPA programs often need to predict the rate of  
18 movement of metals through soils and their subsequent movement and concentrations in ground  
19 water. The primary processes governing the environmental fate and transport of metals in the  
20 subsurface are advection, dispersion, matrix diffusion, and retardation (U.S. EPA, 1994c).  
21 Advection and dispersion are functions of the system rather than of the contaminant. Matrix  
22 diffusion, which is a function of the contaminant, is relatively unimportant and is omitted in most  
23 model transport algorithms. Retardation depends on a number of factors (Langmuir, 1997; U.S.  
24 EPA, 1994c) and may involve or be affected by the following:

- 25 • **Sorption.** The attachment of metal species to mineral surfaces or other surfaces.
- 26
- 27 • **Speciation.** The distribution of a given constituent among its possible chemical  
28 forms, including metal complexes, which have differing tendencies to be adsorbed or  
29 desorbed.
- 30
- 31 • **Precipitation.** The process by which dissolved species exceed the solubility limits of  
32 their solids, so that some of the species precipitate from solution. When a metal  
33 species reaches mineral saturation, addition of further amounts of the species to  
34 solution are precipitated, not adsorbed.
- 35
- 36

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- 1 • **Colloid formation.** The process of forming colloids and the association of metal  
2 species with them. The metals may be sorbed or coprecipitated with colloidal-sized  
3 particles.
- 4
- 5 • **Biofixation.** The binding of metals to solid materials due to the interactions of  
6 microorganisms or plants.
- 7
- 8 • Natural organic matter (NOM) interactions.
- 9
- 10 • Other important processes, such as changes in pH, oxidation potential, salinity,  
11 concentrations of competing ions, the nature of sorbent phases and their surface areas,  
12 and surface site densities.
- 13

#### 14 **4.1.4.1. Metals Sorption**

15 **4.1.4.1.1. Introduction.** The ability of a trace or minor metal such as a metal of concern to sorb  
16 to a substrate is usually the determining factor in its mobility. Physical adsorption, which is  
17 important for molecular organic compounds, is largely inapplicable to the sorption of the trace  
18 metals, which are usually adsorbed as ionic species. In fact, trace metal adsorption is often  
19 relatively independent of the concentrations of the major ions. *Trace metal adsorption is*  
20 *strongly pH-dependent and a function of ionic strength, metal complex formation, competitive*  
21 *ion sorption, redox conditions, and the amounts and reactive surface areas of specific sorbing*  
22 *solids.* The most accurate and mechanistic approach to modeling and predicting trace metal  
23 adsorption is *surface complexation modeling*, using a model such as the *Diffuse Layer (DL)*  
24 *model*, which ideally can account for all of these variables (Langmuir, 1997; Stumm and  
25 Morgan, 1996).

26 The degree of mobility of organic contaminants is often expressed by means of a single  
27 *partition coefficient* ( $K_d$ ) value that describes the distribution of a species between sorbed and  
28 dissolved forms (U.S. EPA, 1995a). Mobility is then calculated from the partition coefficient.  
29 Such an approach is applicable to metal adsorption only when the conditions listed above are  
30 practically constant and are the same in the environment as in the laboratory where the  $K_d$  was  
31 determined, which is rarely the case. In fact, when metal adsorption is described using partition  
32 coefficients, the value of such coefficients typically should be varied by two or more orders of  
33 magnitude to reproduce metal adsorption behavior at a specific site. Therefore, single partition  
34 coefficients for metals are of little value except in broad regional surveys when a large  
35 uncertainty in a  $K_d$  value may be acceptable.

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1 **4.1.4.1.2. Surface complexation models: Diffuse Layer model.** It has been observed that in  
2 many soils and sediments low in organic matter (OM), HFO is the most important metal sorbent  
3 and the only sorbent that needs to be considered in predictions of metal sorption behavior  
4 (Benjamin and Leckie, 1981). The DL model (also called the Generalized Two-Layer Model, or  
5 GTLM) in MINTEQA2 has been extensively applied in aquatic environmental studies of metal  
6 transport and attenuation. Loux et al. (1989) used the DL model and MINTEQA2 to predict the  
7 adsorption and precipitation behavior of eight metals in an oxidized, sandy aquifer as a function  
8 of pH. Assuming that HFO was the only sorbent, DL model adsorption adequately described  
9 changes in the concentrations of Ni, Pb, and Zn in the sediment. Cadmium behavior was better  
10 understood, assuming its precipitation in  $\text{CdCO}_3$  (otavite). Changes in Cu, Ba, Be, and Tl were  
11 not simply explained. Copper may have been adsorbed by organic matter, which was not  
12 considered in the modeling.

13 More recently, adsorption of metals by OM and Al oxyhydroxides as well as HFO has  
14 been included in DL modeling with MINTEQA2 (Paulson and Balistieri, 1999). These authors  
15 studied neutralization of acidic ground waters by ambient surface and ground waters using a  
16 mixing model approach. Particulate organic matter (POM) and HFO were the chief metal  
17 sorbents. In pristine systems, Cu is usually the chief metal associated with particulate organic  
18 carbon (POC); in the Paulson and Balistieri study, Zn and Cd were mostly adsorbed by POM,  
19 and Cu was mostly adsorbed by HFO.

20 It may be possible to estimate metal adsorption with acceptable accuracy without having  
21 to measure it, depending on the information available on a specific soil, surface water, or ground  
22 water system. What is needed minimally is the weight and surface area of potentially sorbing  
23 materials (e.g., metal oxides, clays, and organic matter) in a volume of soil or sediment, or in  
24 suspension in a stream. Literature information can then be used to estimate the sorption  
25 properties of these materials for use in a sorption model. For example, as noted above in the  
26 discussion of the DL adsorption model, where HFO is the dominant sorbent and the amount  
27 suspended in a stream is known, estimation of metal adsorption can be accurate to within  
28 10–20%, as shown in Table 4-3 (Smith et al., 1998). As a general observation, other factors  
29 being equal, it has been found that the surface charge density—and thus the metal adsorption  
30 capacity—of most minerals is largely a function of their surface areas exposed to water (Pabalan  
31 et al., 1998). Thus, the adsorption of metals by Al and Fe(III) oxyhydroxides in a system at a  
32 given pH may be assumed to be the same if they have the same surface areas.

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**Table 4-3. Comparison of model predictions<sup>a</sup> and measured values of percent metals associated with the suspended particulate fraction of mine drainage waters from select sites**

Metal	Argo-3 (pH 5.6, HFO = 0.007 g/L)		Rawley-3 (pH 6.2, HFO = 0.11 g/L)		Leadville drain (pH 7.2, HFO = 0.001 g/L)	
	Predicted	Measured	Predicted	Measured	Predicted	Measured
As	–	–	98	<78 <sup>b</sup>	–	–
Pb	82	<71 <sup>b</sup>	80	<93 <sup>b</sup>	86	<71 <sup>b</sup>
Cu	18	27	60	63	–	–
Zn	<1	0 to 8	<1	0 to 9	2	3
Ni	<1	<1	<1	1	–	–
Cd	<1	<1	<1	6	<1	<1

<sup>a</sup>Model predictions made with the Diffuse Layer model and MINTEQA2.

<sup>b</sup>Dissolved concentration was below the detection limit; value was computed using the limit of detection for the dissolved concentration.

Source: Smith et al., 1998.

1 Cederberg et al. (1985) and Yeh and Tripathi (1991) considered surface complexation  
 2 modeling of metal adsorption and metal transport in ground water. Parkhurst (2002) developed a  
 3 computer model called PHAST,<sup>2</sup> which is a three-dimensional reactive transport model that  
 4 combines PHREEQC, which has the DL metal adsorption model, with HST3D, a ground water  
 5 flow and transport model.

6 Several recent studies have measured and modeled trace metal adsorption and metal  
 7 transport in streams using a surface complexation approach to adsorption. U.S. Geological  
 8 Survey researchers of the Toxic Substances Hydrology Program have published a number of  
 9 papers using the OTEQ and OTIS models. OTEQ is a one-dimensional model for studying the  
 10 fate and transport of metals in streams and rivers. The model couples the OTIS transient storage  
 11 model with MINTEQ, which includes DL model adsorption of metals by HFO (Ball et al., 1999;  
 12 Runkel et al., 1999). Runkel et al. (1999) considered in-stream metal transport, metal oxide  
 13 precipitation-dissolution, and pH-dependent sorption of copper and zinc.

14 If greater accuracy or site specificity is required, it may be necessary to measure metal  
 15 adsorption in laboratory experiments. Such measurements can be performed on pure minerals or  
 16 on whole (usually sieved) soils. The sorption results may be used to develop DL model

<sup>2</sup> See [http://wwwbrr.cr.usgs.gov/projects/GWC\\_coupled/](http://wwwbrr.cr.usgs.gov/projects/GWC_coupled/).

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1 parameters for metal adsorption (Stumm, 1992; Dzombak and Morel, 1990). For example,  
2 model results may then be used to compute partition or distribution coefficients as a function of  
3 pH.

4 Several chemical mass transport codes are available that can model metal transport  
5 through porous media using the DL adsorption model. These models include PHREEQC  
6 (Parkhurst and Appelo, 1999) and CHMTRNS (Noorishad et al., 1987) and, for transport by  
7 streams, OTIS/OTEQ (Runkel et al., 1999). Even if model parameters are estimated on the basis  
8 of literature values, such models will generally predict metal adsorption and retardation more  
9 accurately than is possible when using single or linearly varying distribution coefficient values.

10  
11 **4.1.4.1.3. Application of partition coefficients to metal mobility in ground water.** Owing to the  
12 complexity and multiplicity of the processes involved, recourse is often made to the use of a  
13 single partition or distribution coefficient that describes the degree to which the contaminant's  
14 transport is retarded relative to water. This approach starts with defining the retardation factor:

$$R_f = \frac{v_p}{v_c}$$

15 where  $R_f$  is the retardation factor,  
16  $v_p$  is the velocity of water through a control volume, and  
17  $v_c$  is the velocity of contaminant through a control volume.  
18

19 The retardation factor is related to the distribution coefficient through the expression:

$$R_f = 1 + \frac{\rho_b}{n_e} K_d$$

20 where  $\rho_b$  is the porous media bulk density, and  
21  $n_e$  is the effective porosity at saturation given as a volume fraction  
22 (Langmuir, 1997).  
23

24 Laboratory adsorption studies often find that, in simple systems, the value of  $\log K_d$  for  
25 metal adsorption increases linearly with pH. For example, for  $Zn^{2+}$  adsorption by HFO,  
26 Langmuir (1997) noted that adsorption followed the log equation:

$$\log K_d = -5.48 + 1.77 \text{ pH}$$

27  
28  
29  
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1 The properties and applications of partition or distribution coefficients for metals are  
2 comprehensively reviewed in U.S. EPA (1999a). These coefficients may be obtained from the  
3 literature, estimated using mathematical models, or measured. In batch tests, vessels are filled  
4 with a known amount of sorbent solid mixed with a known volume and concentration of a  
5 sorbing metal species (the sorbate). Often the approach is to systematically vary sorbent or  
6 sorbate concentrations and pH in a series of centrifuge tubes, for example. After centrifugation,  
7 the amount of sorbed concentration is determined by difference from the total sorbate added.  
8 The value of  $K_d$ , usually in mL/g, is then computed from the expression:

$$K_d = (\text{wt. adsorbed} / \text{wt. sorbent}) / \text{solute concentration}$$

11  
12 In column test experiments,  $K_d$  may be computed from test results with the expression:

$$K_d = [(C_i - C_f)V] / C_f * M$$

15  
16 where  $C_i$  and  $C_f$  are the initial and final concentrations of sorbate,  
17  $V$  is the volume of soln passed through the column, and  
18  $M$  is the mass of solids in the column.  
19

20 Properly designed column experiments generally give results more representative of the  
21 in situ behavior of soils and sediments than do batch tests (Langmuir, 1997).

22 Retardation coefficients tabulated as a function of pH by  
23 EPA (U.S. EPA, 1998a) are listed in Table 4-4 for several  
24 important metals of potential concern. EPA (U.S. EPA, 1999b)  
25 has also presented non-pH-dependent values for lead (900),  
26 mercuric chloride (58,000), and elemental mercury (1,000),  
27 which for reasons just discussed should be used with considerable  
28 caution.  
29

**Values for Lead**

Values for lead are not tabulated because of their large variability and the difficulty of selecting values that would be representative or meaningful.

30 **4.1.4.1.4. Limitations.** The use of single  $K_d$  values for a given pH contains potentially large  
31 errors. The values in Table 4-4 will apply only to a specific sorbing phase, amount of sorbent  
32 and its reactive surface area, solution-to-solid ratio, and water chemistry. For example, on a  
33 weight basis, clays are far less sorbing for most metals than are the FHO or OM. Single  $K_d$

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1 values should be limited to regional studies and should be adjusted to account for regional  
2 variations in soil properties.

3  
4 **Table 4-4. Partition coefficients as a function of pH for several important elements**  
5 **of potential concern**

6

7 <b>Element</b>	<b>pH 4.9</b>	<b>pH 6.8</b>	<b>pH 8</b>
8 Arsenic(V)	25	29	31
9 Barium	11	41	52
10 Beryllium	23	790	100,000
11 Cadmium	15	73	4,300
12 Chromium(III)	1,200	1,800,000	4,300,000
13 Chromium(VI)	31	19	14
14 Nickel	16	65	1,900
15 Selenium(VI)	18	5	2.2
16 Silver	0.1	8.3	110
17 Thallium(I)	44	71	96
18 Zinc		62	

19 Source: U.S. EPA (1998b).

20  
21 The distribution coefficient approach may apply if fluid flow in the porous media (soil or  
22 sediment) is isotropic and adsorption is fast, reversible, and linear (Freeze and Cherry, 1979).  
23 These assumptions are often not valid for metal adsorption. Some transport models assume a  
24 constant partition coefficient or linearity of the partition coefficient over all concentration  
25 ranges. To the extent that sorption is not constant and follows a nonlinear isotherm (which is the  
26 usual case for metals), these models will be inaccurate. The best that can be hoped for when  
27 single partition coefficients are used to describe metal adsorption is that they represent bounding  
28 values in a given application.

29  
30 **4.1.4.1.5. Use in decision making.** U.S. EPA (1999a) discusses the advantages and  
31 disadvantages of several methods for measuring partition coefficients, including laboratory batch  
32 testing, in situ field batch testing, flow-through testing, and field modeling. In many national  
33 assessments, EPA has used the MINTEQ model and its subsequent versions to generate generic  
34 partition coefficients that may be applied to regional or national mobility evaluations

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1 (<http://www.epa.gov/ceampubl/mmedia/minteq/index.htm> or <http://www.lwr.kth.se/english/OurSoftware/Vminteq/>).

3 For screening assessments:

- 5 • For screening assessments, partition coefficients have been tabulated as a function of  
6 pH by EPA (U.S. EPA, 1998b), and the Agency has also presented non-pH-  
7 dependent values for lead (900), mercuric chloride (58,000), and elemental mercury  
8 (1,000) (U.S. EPA, 1999b).
- 10 • In simple systems, the value of  $\log K_d$  for metal cation adsorption usually increases  
11 linearly with pH, whereas the value of  $\log K_d$  generally decreases with pH for anion  
12 adsorption.

14 For definitive assessments:

- 16 • It may be possible to estimate metal adsorption with some accuracy without having to  
17 measure it, depending on the information available on a specific soil, surface water,  
18 or ground water system. What is needed minimally is the amounts and surface areas  
19 of the potentially sorbing materials (e.g., metal oxides, clays, and OM) in a soil or  
20 sediment or in suspension in a stream, and the detailed chemical composition of the  
21 water, especially its pH and metal concentration. Literature information can then be  
22 used to estimate the sorption properties of these materials for use in the DL sorption  
23 model, for example.
- 25 • If greater accuracy or site specificity is required, it may be necessary to measure  
26 metal adsorption in laboratory experiments designed to parameterize the DL model  
27 for application to a specific study area. The experiments could be batch tests that  
28 attempt to reproduce the composition of waters and sorbing solids in the study area.

#### 30 4.1.5. Sediment Chemistry

31 In addition to the challenges posed by metal chemistry, the sedimentary environment is  
32 complex and often highly heterogeneous. Fortunately, we can generalize about the sedimentary  
33 environment and the main controlling factors to progress toward a method for risk assessment.  
34 This brief review summarizes information on the composition of sediments; processes that act on  
35 sediments and their metal burden; and the chemistry of the sedimentary environment that  
36 influences the fate, bioavailability, and effects of metals. It is important to consider these factors  
37 in light of the aim of estimating potential biological effects of metals in sediments.

38 Sediment solids can hold up to a million times more metal than an equivalent volume of  
39 water. The exact proportions of a chemical held by sediment relative to water is a function of a

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1 metal's chemistry as well as the chemistry of the sediment solid and the surrounding  
2 environment, and this distribution is dynamic (Diamond and Mudroch, 1990). Because of their  
3 large capacity to "hold" metals, sediments have been characterized as "sinks." Although this is  
4 largely true, sediments are now appreciated to be  
5 temporary sinks, wherein some of the metal can enter  
6 ecological and human food webs through several  
7 routes (e.g., Diamond, 1995), primarily through  
8 accumulation in benthic organisms. These organisms  
9 include those that fully or partially live in the  
10 sediments (e.g., tubificids, chironomids, trichopteran  
11 larvae) or those that feed from the sediment bed (e.g.,  
12 suckers, carp). Some organisms obtain their chemical  
13 dose from both pelagic and benthic routes (e.g., lake  
14 whitefish, walleye), but because of high chemical  
15 concentrations in sediments, the benthic route can be  
16 the dominant route of uptake (Morrison et al., 2000).

17 For humans, the route of entry of metals from  
18 the sediments is through water used for drinking,  
19 bathing, and swimming. The availability of these metals is mediated by sediment-water  
20 exchange processes that can result in the release or remobilization of chemicals from the  
21 sediment bed. However, due to the ability of Hg to bioaccumulate in its monomethyl form, fish  
22 consumption is the critical route of exposure to this metal for humans.<sup>3</sup>

23 Many important chemical reactions involving the metals of concern occur in the fine-  
24 grained materials that accumulate in the deep parts of water bodies. The controlling factors or  
25 master variables that influence metal chemistry are redox potential and pH. A depth profile of  
26 the sediments will reveal decreasing sediment porosity and concentrations of dissolved oxygen  
27 because oxygen is consumed as organic matter decomposes. pH is often relatively constant or  
28 may decrease with depth, but alkalinity may increase owing to mineralization of organic matter  
29 (Stumm and Morgan, 1996). As dissolved oxygen is consumed, anaerobic microbes use other  
30 electron acceptors in redox or oxidation-reduction reactions in the order of nitrate, ferric iron,  
31 ammonium, sulfate, and bicarbonate to produce carbon dioxide, ammonia, sulfide, and methane.

#### Sediments

Bed or bottom sediments are found at the bottom of lakes, rivers, and estuaries. Sediments have several sources that influence their composition and chemistry. The type and chemistry of sediments is also determined by their location in the water body as well as the characteristics of the water body. At any given site, metals can be associated with solid-phase minerals, organic matter, colloids, and pore water. The solid phase can vary from sand (>63 μm), to silt (2–63 μm), to clay (<2 μm). Because clays have more active binding sites than do the other grain sizes and because of their high surface area-to-volume ratio, fine-grained particles are of greatest significance in terms of metal binding.

---

<sup>3</sup>Marine biota can also be a significant route of exposure to arsenic in its organic forms, such as methylarsonic acid; however, these arsenic species are significantly less toxic than the inorganic forms (Fowler, 1983).

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1 The sequence of microbially mediated redox reactions that produce these reduced species are  
2 given in Figure 4-2.

3 The redox status of the sediments can be assessed by measuring the concentration of  
4 dissolved oxygen or other redox-sensitive species, by calculating pE (a measure of electron  
5 availability in solution), or by measuring Eh (millivolts) or the electromotive force of the pore  
6 water solution. The zone of transition from oxic to anoxic conditions is the redoxcline, which  
7 can migrate vertically, depending on the mixing of the overlying water column (e.g., Diamond  
8 and Mudroch, 1990). For example, the redoxcline may be 5 to 10 cm below the sediment-water  
9 interface in a well-oxygenated oligotrophic lake or river, but it may be above the sediment-water  
10 interface in a thermally stratified eutrophic lake or river.

#### 11 12 **4.1.5.1. Metal Chemistry in Sediments**

13 In this discussion, two pools of metals should be considered. The first pool consist of  
14 metals that exist as aqueous (or dissolved) species bound to colloids or DOM and those bound to  
15 sediment particles through an exchangeable binding process. This pool is often referred to as the  
16 exchangeable or labile pool. The second pool consists of metals found within the mineral matrix  
17 of the sediment solids. This pool is largely unavailable to biota, and its release will occur over  
18 geologic time scales through diagenetic processes. Because the latter pool is largely unavailable,  
19 we will consider only the exchangeable pool of metals. Note that the exchangeable pool will be  
20 composed of naturally occurring metals that are released into solution due to weathering and  
21 diagenetic processes as well as metals released into the environment due to anthropogenic  
22 activities.

23 The exchangeable pool of metals is subject to speciation in the aqueous phase (e.g.,  
24 within the pore water) and sorption to solid phases, where sorption is a general term that includes  
25 adsorption (the accumulation of matter at the solid-water interface or a two-dimensional process)  
26 and absorption (inclusion in a three-dimensional matrix) (Stumm and Morgan, 1996). Here,  
27 speciation refers to the distribution of metal species in a particular sample or matrix or species  
28 distribution (Templeton et al., 2000). In the aqueous phase, metal will react or bind with  
29 dissolved ligands according to the pH, Eh, ionic strength, and abundance of ligands (see above  
30 discussion on aquatic chemistry).

31 The concentration of metal in the dissolved phase is controlled by sorption to the solid  
32 phase. Although sorption can occur rapidly, desorption or dissolution of metal from the solid  
33 phase may be a two-phase process, where the second phase is rate limiting (e.g., Nyffeler, 1986;  
34 Santschi et al., 1986). If we neglect the kinetic limitation of reactions, the distribution of metals

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1 among aqueous species and between the aqueous phase and the solid phase can be estimated.  
2 Several speciation/complexation models are available to perform this calculation, such as  
3 MINEQL+ (Schecher and McAvoy, 2001), the Windermere Humic Aqueous Model (WHAM)  
4 (Lofts and Tipping, 1998), and MINTEQA2 (Allison et al., 1991). These models work well  
5 under oxic conditions, but estimates of metal binding are less reliable under anoxic conditions,  
6 where metal concentrations are most often controlled by the solubility of metal sulfides.  
7 Furthermore, in some circumstances equilibrium may not be achieved, particularly when the  
8 redoxcline moves more quickly than the rate of metal reaction or when the reaction is governed  
9 by microbial processes, as occurs with the methylation of mercury or arsenic.

10 In oxic sediment pore waters (above the redoxcline), metals will exist as aqueous  
11 species, that is, as freely dissolved ions or metal complexes (e.g., phosphate, sulfate, or  
12 carbonate complexes), and associated with colloids. Solid-phase reactions are controlled by iron  
13 oxyhydroxides and manganese oxides that may exist as colloids, sediment particles or surface  
14 coatings of particles, OM that may also exist as colloids or coat sediment particles, and clay  
15 colloids and particles.

16 As Eh declines, the solid-phase manganese oxides are the first to be reduced and thereby  
17 dissolve, which releases metals that have been sorbed or coprecipitated with these minerals.  
18 Some of the metals released into the pore water may then be adsorbed by iron(III)  
19 oxyhydroxides, which are the next to dissolve as the Eh continues to drop. Under reducing  
20 conditions, particularly as sulfate is consumed and the sulfur is converted to sulfide, metal  
21 concentrations in pore waters again drop as solid-phase metal-sulfides are formed (see discussion  
22 below about the role of AVSs in regulating toxicity).

23 As a result of redox chemistry, metals can undergo seasonal redox-driven cycling  
24 between the water column and sediments or within the sediments, depending on the position of  
25 the redoxcline. The stages in the cycling are, first, the adsorption or coprecipitation of metals  
26 with iron and manganese hydroxides under oxidizing conditions; then with the development of  
27 moderately reducing conditions, the reduction and dissolution of the manganese and iron  
28 oxyhydroxides, and consequent release of the associated metals into the water or pore water;  
29 followed by their diffusion upward toward the zone of low metal concentrations under oxidizing  
30 conditions. It is also possible for dissolved metals to diffuse downward toward the zone of low  
31 metal concentration owing to their precipitation as sulfides. As a result of this vertical cycling,  
32 the depth profile of metals in pore water may not match that of the solid phase (e.g., Carignan  
33 and Tessier, 1985). Moreover, it is possible, but less usual, that the cycling can occur relatively  
34 rapidly and involve a significant portion of the solid-phase metal. Under these conditions, the

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1 solid-phase sediment profile reflects this reworking rather than the historical record of metal  
2 loadings (MacDonald et al., 2000).

3 pH controls metal speciation and binding by affecting the species distribution of  
4 dissolved ligands (e.g., phosphate, sulfate, carbonate, humic substances) and the surface charge  
5 of binding sites on DOM and solid phases such as iron oxyhydroxides. Generally, at low pH,  
6 when surface sites are protonated, the sorption of cationic metals decreases, and, hence, metal  
7 mobility increases. The converse occurs at high pH, which results in low metal solubility and  
8 greater sorption. The patterns of dissolution and sorption are reversed for metalloids, such as  
9 arsenic, that exist as anionic species.

#### 11 **4.1.5.2. *Methods of Estimating Metal Distribution in Sediments***

12 **4.1.5.2.1. *Application.*** A main objective in terms of assessing the hazard or risk posed by  
13 metals is estimating metal in the dissolved phase that is potentially bioavailable. Accordingly,  
14 several methods have been developed to estimate the distribution of metals among dissolved and  
15 solid phases in sediments. These methods have been thoroughly reviewed by Mudroch et al.  
16 (1999, 1997). Although bioavailability is also a function of aqueous phase speciation (see  
17 Section 4.1.3, Aquatic Chemistry), limited research has been conducted to estimate metal  
18 speciation in pore waters. Generally, ecological risk assessments assume that the exposure of  
19 benthic organisms to sediment-associated metal is proportional to the metal concentrations in  
20 interstitial water, although some studies indicate that uptake from overlying water (Hare et al.,  
21 2003; Roy and Hare, 1999) or ingested sediment may be a significant source of body burdens of  
22 metals (see Section 3.2, Metals Risk Assessment Recommendations for Aquatic Environments,  
23 for more discussion of this topic).

24 Distribution of metals in sediment pore waters may be determined by field  
25 measurements, experimental methods, and mathematical modeling, with the latter also requiring  
26 some field measurements. Concentrations of metals in pore waters may be determined in the  
27 field by use of pore water dialysis chambers or peepers and by methods that separate the solid  
28 phase from the pore water, although the latter have been shown to be less reliable (Mudroch et  
29 al., 1997). Several extraction schemes have been developed to determine the distribution of  
30 metal among operationally defined fractions (e.g., Forstner, 1995a; Tessier et al., 1979). It is  
31 well known that sequential extraction methods do not cleanly distinguish the occurrence and  
32 speciation of different forms of metals in sediments and soils (Tye et al., 2003; Verloo, 1999).  
33 Other experimental methods include leaching tests (e.g., Reuther, 1999). The results of any of

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1 these methods are concentrations of metals in pore water, which can be related to toxicity  
2 benchmarks.

3 Because of the need to develop Sediment Quality Guidelines (SQGs) for metals that  
4 explicitly address toxicity and are based on readily measured parameters, several methods have  
5 been developed. For oxic sediments, Tessier and co-workers (Tessier et al., 1993, 1989, 1984;  
6 Tessier, 1992) compiled partition coefficients of metals that were derived from field studies of  
7 freshwater sediments. The partition coefficients are dependent on pH (because Eh is held  
8 constant) and are generally linear over a range of pore water pH values (see above discussion  
9 under ground water chemistry for the theoretical basis for development of partition coefficients).  
10 Speciation/complexation models also may be used to estimate fractions of dissolved and bound  
11 metal species. These models rely on measurements of pH, dissolved oxygen, or Eh to establish  
12 redox conditions. The models assume that solid-phase binding is governed by sorption to iron  
13 and manganese oxides. Model estimates are less reliable when other solid-phase substrates are  
14 dominant (e.g., clay minerals) and are a function of the availability and accuracy of the stability  
15 constants for the metal-ligand reactions that are used in the calculations.

16 For anoxic sediments, the availability of sulfide controls metal distribution and solubility.  
17 Operationally, AVSs—mainly iron monosulfide—have been considered as a measure of reactive  
18 sulfides (Forstner, 1995a). Studies have demonstrated an inverse relationship between sediment  
19 toxicity and AVSs for marine and freshwater sediments (Di Toro et al., 1992, 1990; Ankley et  
20 al., 1991) as well as between pore water concentrations and AVSs (Brumbaugh et al., 1994;  
21 Casas and Crecelius, 1994). As a screening-level tool, the toxicity of anoxic sediments can be  
22 assessed by determining the ratio of AVSs to simultaneously extracted metal (SEM). Low  
23 sediment toxicity is indicated when AVSs are in excess ( $AVS > SEM$ ), which implies sufficient  
24 capacity of the AVS to bind essentially all free metal. This topic is further discussed in Section  
25 4.5.

26 For estimating effects concentrations in the sediment, there are many different SQGs,  
27 which vary in their derivation and the degree to which they incorporate bioavailability  
28 considerations. Many of the published SQGs are based on empirical relationships between  
29 biological effects and the total (dry weight) concentrations of sediment contaminants (e.g.,  
30 McDonald et al., 1996; Long and Morgan, 1991). Although these empirically based guidelines  
31 do show general relationships between the degree of sediment contamination, they do not  
32 explicitly account for site-specific differences in bioavailability of contaminants. Although EPA  
33 has not formally adopted any single SQG approach as an Agency standard, it has been active in  
34 developing the “equilibrium partitioning” (EqP) approach to SQG development. The EqP

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1 approach considers effects of sediment chemistry on bioavailability by comparing the  
2 concentrations of AVSs, SEM, and organic carbon (U.S. EPA, 2000c; Ankley et al., 1996, 1993;  
3 Di Toro et al., 1992, 1990). This approach is reviewed in more detail in Section 4.5,  
4 Characterization of Ecological Effects.

5  
6 **4.1.5.2.2. Limitations.** Model estimates are less reliable when other solid-phase substrates are  
7 dominant (e.g., clay minerals), and they are a function of the availability and accuracy of the  
8 stability constants for the metal-ligand reactions that are used in the calculations.

#### 9 10 **4.1.6. Soil Chemistry**

11 The cationic metals occur naturally in soils as oxides and hydroxides (Fe, Mn, Al); to a  
12 lesser extent as carbonates, phosphates, and sulfates; and in reducing (usually wet or  
13 waterlogged) soils as sulfides, which are highly insoluble. The soil parameters important in  
14 affecting sorption and precipitation reactions and the extent of their influence—and thus  
15 contaminant bioavailability—depend on the intrinsic properties of the contaminants. In the soil  
16 environment, metals can exist as cations, anions, or neutral species. Their form significantly  
17 affects their sorption, solubility, and mobility. For example, most soils are chiefly negatively  
18 charged; thus, metal cations have a higher propensity to be sorbed by soil particles than do metal  
19 anions (U.S. EPA, 2003e).

20 Cationic metals can react with inorganic soil constituents (e.g., carbonates, sulfates,  
21 hydroxides, sulfides), forming aqueous complexes, which may be adsorbed or precipitated in  
22 mineral form. Most complexation and precipitation reactions are pH dependent (U.S. EPA,  
23 2003e).

24 Arsenic, chromium, selenium, and vanadium complex with oxygen and typically exist as  
25 anionic species under most environmentally relevant scenarios (Bohn et al., 1985; Lindsay,  
26 1979). The most common forms of arsenic are arsenate (arsenic(V)) and arsenite (arsenic(III)),  
27 which are present in soil solution in the form of  $\text{AsO}_4^{3-}$  and  $\text{AsO}_3^{3-}$ , respectively. The chemistry  
28 of arsenic resembles that of phosphate (Barber, 1995; Bohn et al., 1985). Chromium can exist as  
29 chromate (chromium(VI) or  $\text{CrO}_4^{2-}$ ), which is usually considered more soluble, mobile, and  
30 bioavailable than the sparingly soluble chromite (chromium(III)), which is normally present in  
31 soil as the precipitate  $\text{Cr}(\text{OH})_3$  (Barnhart, 1997; James et al., 1997). Similarly, selenium can be  
32 present as selenates ( $\text{SeO}_4^{2-}$ ) and selenites ( $\text{SeO}_3^{2-}$ ). For vanadium, vanadate ( $\text{VO}_4^{3-}$ ) is the most  
33 common form (U.S. EPA, 2003e).

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1 Metals can exist in the pore water as aquo-ions or soluble complexes. The bonding of  
2 metal species to soil particles can range from ionic to covalent. For most soils in the United  
3 States, negatively charged sites are more plentiful; less than 5% of the total available charge on  
4 the soil surface is positively charged. Metals existing as cationic species have a greater  
5 propensity to associate with such soils. This makes them less bioavailable, but it also results in  
6 greater loading of metals into the soil ecosystem. Anionic metals generally move into pore  
7 water—and so are more bioavailable—but leach out of the system much more rapidly. In  
8 summary, soil pH and availability of charged sites on soil surfaces are the primary soil factors  
9 controlling release of metals to pore water and, subsequently, bioavailability (U.S. EPA, 2003e).

#### 11 **4.1.6.1. Key Parameters Affecting Metal Bioavailability in Soils**

12 From the preceding overview of how the metals and metal compounds interact with soil  
13 constituents, it is clear that soil plays a very significant role in reducing the potential  
14 bioavailability of metals in the environment. Given the types of contaminant-soil interactions  
15 presented, the primary soil factors controlling the potential bioavailability of metals are soil pH,  
16 the availability and character of sorption sites on soil surfaces, the content of Fe and Al  
17 oxyhydroxides and soil organic matter, and least important, the soil clay mineral content. The  
18 following discussion briefly details the key soil parameters affecting the various contaminants  
19 availability to the pore water.

20 Soil pH is often termed the master soil variable because it controls virtually all aspects of  
21 contaminant and biological processes in soil. These processes include dissolution and  
22 precipitation of metal solid phases, complexation and acid-base reactions of metal species, and  
23 metal sorption as well as microbial activity. Increasing soil pH also results in an increase in the  
24 number of negatively charged soil sites, with a concomitant decrease in the positively charged  
25 sites. Therefore, increasing the soil pH increases the sorption and removal from pore water  
26 (Bohn et al., 1985).

27  
28  
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1 **4.1.6.2. Adsorption Behavior of the Metals of Concern**

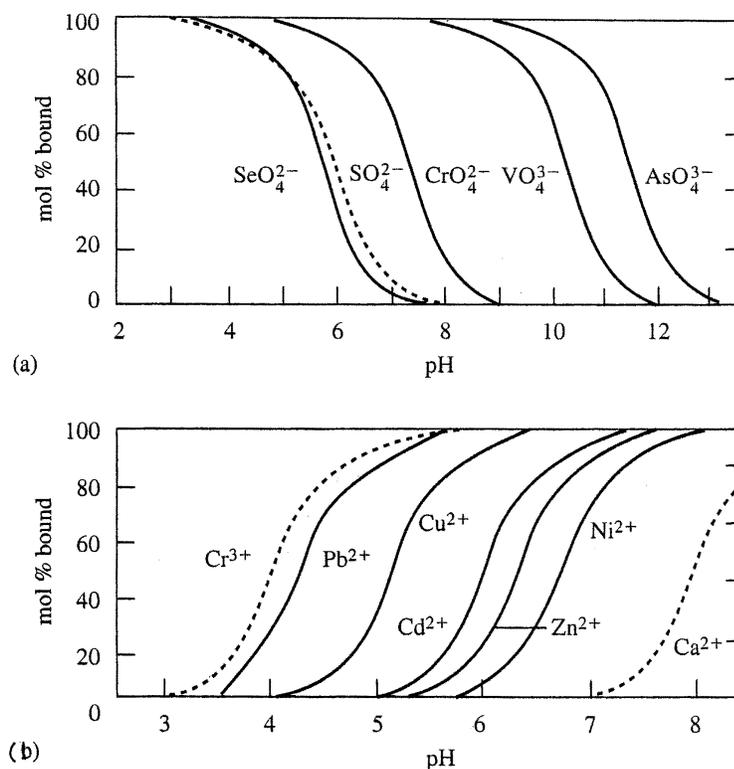
2 **4.1.6.2.1. Surface area and surface density.** In porous media, the most important sorbent solids  
3 for metals are oxyhydroxides of Fe and Mn. Their important surface properties are discussed in  
4 Langmuir et al. (2003). For a given weight of sorbent, metal sorption capacity is proportional to  
5 surface area and surface site density. The greatest surface site densities (positively or negatively  
6 charged sites) are those of organic material and the oxyhydroxides. These phases are the  
7 strongest and most important sorbents of trace metals. Except for kaolinite, the clays (0.02–2  
8 mmol sites/g) have a surface charge that is largely independent of pH, whereas the surface  
9 charge of organic matter and the oxyhydroxides is strongly pH dependent.

10  
11 **4.1.6.2.2. Importance of pH.** The pH at which a solid surface changes sign is referred to as the  
12 zero point of charge (ZPC). The ZPC of organic matter and kaolinite are below the range of  
13 usual pH and while their surface charge decreases with decreasing pH, their surfaces remain  
14 negatively charged above pH 2–3. In contrast, Fe(III) and Al oxyhydroxides, such as goethite  
15 ( $\alpha$ -FeOOH), have ZPC values near pH 7–8 and so have a positive surface charge at low pH and a  
16 negative surface charge at high pH. Thus, the negative surface charge of the oxyhydroxides, and  
17 their sorptive capacity for metals, increases with increasing pH. Conversely, the positive surface  
18 charge of the oxyhydroxides increases as the pH drops, making these phases more effective  
19 sorbents for anions under low pH conditions.

20 These effects are shown in Figure 4-3, which is a plot of percentage sorbed versus pH for  
21 metal adsorption by ferrihydrite, or HFO. The curves are called *sorption edges* for each metal.  
22 The diagram shows that the oxyanions are strongly adsorbed by HFO under acid conditions but  
23 are desorbed (become mobile) with increasing pH. Based on Figure 4-3, combined with  
24 adsorption data assembled by Dzombak and Morel (1990), the order of desorption from HFO  
25 with increasing pH is selenate, antimonate, molybdate, chromate, vanadate, arsenate, and  
26 phosphate. Selenate desorbs between pH 3 and 8, whereas arsenate is strongly held at lower pH  
27 values and desorbed between pH 9 and 11. Also based on Figure 4-3 and Dzombak and Morel  
28 (1990), with increasing pH, HFO preferentially adsorbs metals in the order  
29  $\text{Hg}^{2+} > \text{Be}^{2+} > \text{Ba}^{2+} > \text{Cr}^{3+} > \text{Pb}^{2+} > \text{Cu}^{2+} > \text{Cd}^{2+} \approx \text{Zn}^{2+} > \text{Ni}^{2+}$ .

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**Figure 4-3. Adsorption of various metal cations and oxyanions, each at  $5 \times 10^{-7}$  M, by ferrihydrite ( $\Sigma\text{Fe[III]} = 10^{-3}$  M) as a function of pH at an ionic strength of 0.1 mol/kg. There are  $2 \times 10^{-4}$  M of reactive sites on the oxyhydroxide. The dashed curves are calculated after Stumm (1992).**

1 **4.1.6.2.3. Organic matter (organic carbon) content.** Organic matter includes plant and animal  
 2 remains in various stages of decomposition, cells and tissues of soil organisms, and substances  
 3 exuded from plant roots and soil microbes (Sumner, 2000). Organic matter is primarily  
 4 composed of carbon, oxygen, and minor amounts of nitrogen and phosphorus. On average,  
 5 approximately 58% of organic matter is organic carbon. Soils encompass a range in organic  
 6 matter, from <1% for a sandy soil to almost 100% for a peat soil, with most soils having organic  
 7 matter contents <10% (Bohn et al., 1985). Also, organic matter content is usually higher in

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1 surface soils or in the root zone and decreases with depth in the soil profile. The organic acid  
2 functional groups typically present in organic matter have a high affinity to attract metal cations.

### 3 4 **4.1.6.3. *Aging of Metals in Soil***

5 A distinction should be made between persistence of total metals in soil and persistence  
6 of bioavailable forms of the metal. As metals age in soils, they decrease in bioavailability over  
7 time. It has been well documented that metal chemistry in solutions freshly added or spiked into  
8 soils vary from metal forms in field-contaminated soils. Typically, the metal contaminant pool  
9 requires time to diffuse into micro- or nanopores and to be absorbed onto organic matter and soil  
10 particles. These slow reactions are attributed to micropore diffusion, occlusion in solid phases  
11 by (co)precipitation and (co)floculation, and cavity entrapment. Although the slow reactions  
12 play a key role in metal bioavailability, their rates, mechanisms, and controlling factors have not  
13 been comprehensively elucidated. Evidence of aging processes is provided by studies of metal  
14 extractability and lability (Young et al., 2003; Hamon et al., 1998). It has been frequently  
15 observed that easily extractable pools revert with time (~1 year) to more strongly bound forms.  
16 Isotopic dilution provides a useful way to quantify changes associated with progressive  
17 attenuation of metals in soil. Aging reactions are almost over after about 1 year and are  
18 reversible. At present, information regarding the aging reactions of different metals and  
19 metalloids, and sorbing solids, is very limited, so it is not possible to generalize which metal(s)  
20 age the fastest or with greater/less reversibility.

#### 21 22 **4.1.6.3.1. *Steady-state calculations for metals in soils***

23 **4.1.6.3.1.1. *Application.*** Aging reactions can be determined empirically by calculating  
24 partitioning through measurement of the soil pore water concentration of a metal in well-  
25 equilibrated soils. If  $K_d$ s are calculated from adsorption isotherms, aging should be considered  
26 separately. This is related to the high affinity of metals for soil solid phases. In fact, for most  
27 metals, metal losses by leaching, erosion, or removal by a crop that is harvested are small when  
28 compared with the total metal concentration. However, for some elements, such as Se, the half-  
29 life in soil is significantly shorter. Critical factors that affect the mass balance of metals are the  
30 anthropogenic and natural inputs and outputs via leaching to ground water, the removal through  
31 surface erosion, and crop harvesting. Excluding erosion processes, the elimination half-life of  
32 metals in soil ( $t_{1/2}$ ) can be predicted from a soil mass as follows:

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$$t_{1/2} = \frac{0.69 \times d \times 10000}{y \times TF + \frac{R}{K_d}}$$

1 where d is soil depth in meters,  
 2 y is annual crop yield (t ha<sup>-1</sup> y<sup>-1</sup>),  
 3 TF is the ratio of the metal concentration in plant to that in soil,  
 4 R is the net drainage loss out of the soil depth considered (m<sup>3</sup> ha<sup>-1</sup> y<sup>-1</sup>),  
 5 K<sub>d</sub> is the ratio of the weight of metal adsorbed to the weight of the sorbent,  
 6 divided by the metal concentration in a volume of soil solution (L Kg<sup>-1</sup>).  
 7

8 The time required to achieve 95% of steady state is about four half-lives. This is shown  
 9 in Table 4-5 for select metals and metalloids. Selenium approaches steady state after only 1  
 10 year. As a consequence, Se soil concentrations after 100 years and at steady state are identical.  
 11 In contrast, concentrations of Cu, Cd, Pb, and Cr(III) are still well below steady-state values after  
 12 100 years and, consequently, their concentrations in soil are very similar. The time necessary to  
 13 approach steady state is a function of the loading rate and K<sub>d</sub>.  
 14

15 **Table 4-5. Time to achieve 95% of steady-state metal concentration in soil and total**  
 16 **soil metal concentrations after 100 years and at steady state<sup>a</sup>**  
 17

Metal	Loading rate (g ha <sup>-1</sup> y <sup>-1</sup> )	K <sub>d</sub> (L kg <sup>-1</sup> ) <sup>b</sup>	T (years)	Soil metal concentration (mg added metal kg <sup>-1</sup> )	
				Steady state	After 100 yrs
Se	100	0.3	1.3	0.01	0.01
Cu	100	480	1860	16	2.4
Cd	100	690	2670	23	2.4
Pb	100	19000	73300	633	2.6
Cr III	100	16700	64400	556	2.6

18  
 19  
 20  
 21  
 22  
 23  
 24 <sup>a</sup>Assumes a soil depth of 25 cm and a net drainage loss of 3000 m<sup>3</sup> ha<sup>-1</sup> y<sup>-1</sup>; background was zero at the start of  
 25 loading.

26 <sup>b</sup> Mean K<sub>d</sub> (ratio of total metal concentrations in soils to that in pore water) of 49 Dutch soils (De Groot et al., 1998).  
 27

28 Source: Adapted from Smolders et al. (2004).  
 29

30 **4.1.6.3.1.2. Limitations.** It should be noted that the time needed to approach steady state for all  
 31 the metals considered, except Se, is in the order of thousands of years, and it is difficult to

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1 envision that soil conditions would not change in this time frame. However, the concept is  
2 useful because it provides a time frame within which to consider risk as a function of loading  
3 capacity of the soils and the potential for continued exposure even after cessation of  
4 anthropogenic inputs of metals to soils. Limitations on the application of  $K_d$  are discussed in  
5 Section 4.1.4.

#### 6 7 **4.1.6.3.2. *Laboratory methods to simulate aging in soils***

8 **4.1.6.3.2.1. *Application.*** The aging effect requires laboratory studies on soils to apply a time-  
9 dependent weathering or aging treatment of spiked soils. Critical toxicity values generally are  
10 based on toxicological tests performed during the period of relatively fast metal adsorption that  
11 follows metal addition to soil. Such values would be lower than those derived from a similar  
12 study conducted with soils a year or more after addition of the metal. McLaughlin et al. (2000)  
13 proposed that toxicity thresholds be set using a sequential testing procedure. Tests would be  
14 conducted within 2 to 7 days following incorporation of the test substance to generate an  
15 estimate of acute hazard. Another set of soils would be tested 60 days after mixing, and a third  
16 would be subject to a leaching process and also tested after the 60-day period. It has been  
17 estimated in preliminary studies that toxicity is reduced up to 10-fold owing to aging of metals in  
18 soils. However, further studies are warranted and standardized processes should be agreed upon  
19 before metals aging can be properly accounted for in soil toxicity testing.

20  
21 **4.1.6.3.2.2. *Limitations.*** The leaching and equilibrium times are limited for practical reasons  
22 and are meant to simulate some degree of aging and dissolution as a result of weathering. If  
23 aging occurs at faster rates than does dissolution, then toxicity will decrease with time.  
24 Conversely, if dissolution occurs at a greater rate, then toxicity will increase.

#### 25 26 **4.1.6.4. *Dissolution and Transformation of Metals***

27 **4.1.6.4.1. *Application.*** The dissolution and transformation of a metal compound in soil is  
28 related to a series of chemical and physical properties characteristic of the compound itself and  
29 of the soil. Environmental parameters such as temperature and humidity have a strong influence  
30 on the rate of transformation. When assessing the transformation of a compound in soil, it  
31 should be remembered that aging reactions may take place at the same time as transformation  
32 and dissolution. When metal salts are added to soil, the form of the salt dictates the rate and  
33 amount of soluble metal that will form in the pore water. Insoluble forms of metals (e.g.,  
34 vanadium pentoxide [ $V_2O_5$ ]) will transform to soluble free ion (V) at a slower rate than will

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1 soluble metal salts (e.g.,  $\text{Na}_3\text{VO}_4$ ). However, the rate of formation of the free ion is not  
2 proportional to the dissolution rate of the salt because aging reactions will remove the free ion  
3 from the pore water. The relative rates of dissolution and aging should be considered  
4 simultaneously to accurately predict pore water concentrations.

5  
6 **4.1.6.4.1.1. *Solution speciation (computer-based models).*** Computer-based models employ  
7 either equilibrium constants or Gibbs free energy values to determine metal speciation from  
8 solution chemistry conditions (concentration, pH, Eh, organic complexes, adsorption/desorption  
9 sites, and temperature). Both approaches are subject to mass balance and equilibrium conditions  
10 that should be defined. In recent years, as more accurate thermodynamic data have become  
11 available, the models have undergone extensive development and can provide useful predictive  
12 estimates of metal behavior. A good review of these models and their applications is provided  
13 by Lumsdon and Evans (1995). Examples of computer-based speciation models include  
14 MINTEQL, REDEQL2, ECOSAT, MINTEQA2, HYDRAQL, PHREEQC, and WATEQ4F.

15 Both MINTEQA2 (U.S. EPA, 1991b) and VMINTEQ (Gustafson, 2003) contain  
16 subroutines that allow estimates of the importance of metal-organic complexing if the  
17 concentration of DOC is known. Perhaps more useful in studies of metals in soil solution are  
18 programs such as WHAM (Tipping, 1998, 1994) and the Non-Ideal Competitive Adsorption  
19 (NICA) model (Goody et al., 1995). Application of the chemical speciation model WHAM has  
20 been discussed by Tye et al. (2003), who successfully predicted  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  activities in soil  
21 pore water by assuming the metals were adsorbed by soil humus according to a pH-dependent  
22 Freundlich isotherm model. Competitive adsorption between  $\text{Ca}^{2+}$  and  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  could be  
23 ignored because it did not improve model fits.

24  
25 **4.1.6.4.2. *Limitations.*** In some instances, metal speciation is controlled by simple reactions.  
26 However, in many cases (particularly in contaminated media), the state of equilibrium and the  
27 reversibility of metal reactions are unknown. In addition, mathematical thermodynamic  
28 equilibrium models suffer from other limitations, such as lack of reliable thermodynamic data for  
29 relevant species, inadequacies in models to correct for high ionic strength, poorly known  
30 reaction kinetics, and complex reactions and lack of models for co-precipitation/adsorption. The  
31 first limitation is perhaps the most significant for contaminated media. As an example, none of  
32 the models can predict the behavior of the common, anthropogenic lead phases in paint, solder,  
33 or slag.

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#### 1 **4.1.6.5. Soil Metal Transfer to Plants**

2 The “soil-plant barrier” concept was introduced to communicate how metal addition, soil  
3 chemistry, and plant chemistry affect risk to animals from metals mixed in soil (Chaney,  
4 1980). Reactions and processes that take place at the soil-plant barrier are influenced by the  
5 following factors: (1) soil solid phases (e.g., Fe, Al, and Mn oxyhydroxides and organic matter)  
6 may have adsorptive surfaces that influence soil chemistry; (2) adsorption or precipitation of  
7 metals in soils or in roots limits uptake-translocation of most elements to shoots; and (3) the  
8 phytotoxicity of Zn, Cu, Ni, Mn, As, B, Al, and F, for example, limits residues of these elements  
9 in plant shoots. More recently, inductively coupled plasma-mass spectrometry (ICP-MS) and  
10 other very sensitive analytical methods have been used to examine soil solution and soil-plant  
11 transfer of 60 elements as a function of soil pH (Tyler and Olsson, 2001a, b). These studies  
12 provide evidence that further supports the concept of the soil-plant barrier

13 For strongly adsorbed metal cations, the pattern of plant response to metals contained in  
14 biosolids is strongly curvilinear (i.e., plant metal concentration approaches a plateau with  
15 increasing soil metal concentration), rather than being linear with increasing concentration.  
16 Several areas for potential errors in the research methodology should be avoided when making  
17 these comparisons:

- 18
- 19 • First, comparison of application rates is valid only after the system has equilibrated  
20 utilizing accepted methods.
- 21
- 22 • Second, soil pH levels should be equal across rates studied; co-variance of soil pH  
23 should be used to correct for unequal soil pH (Bell et al., 1988).
- 24
- 25 • Third, the metal concentration in the source applied affects the slope of metal uptake:  
26 higher metal concentration in the source means higher phytoavailability at equal  
27 metal applications (Jing and Logan, 1992).
- 28

29 **4.1.6.5.1. Application/limitations.** Strongly acidic soils increase plant uptake of Zn, Cd, Ni,  
30 Mn, and Co and increase the potential for phytotoxicity from Cu, Zn, and Ni. In alkaline soils,  
31 the high pH increases uptake of Mo and Se. Lead and Cr are not absorbed by plants to any  
32 significant extent at any pH (Chaney and Ryan, 1993). However, each element should be  
33 considered separately because of its unique chemistry. For example, arsenate is more strongly  
34 adsorbed than is arsenite; when a soil is flooded to grow rice, soil microbes can reduce arsenate  
35 to arsenite, and the higher concentration of dissolved arsenite can be phytotoxic to rice in more  
36 highly contaminated soils. Most other elements have little potential for redox change with  
37 change in the redox status of soils. Reduced soils can form sulfide, and sulfide forms low-

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1 solubility compounds with most of the metals of concern in soils, including Pb, Zn, Cd, Cu, and  
2 Ni (see above discussion on sediment chemistry). For essential elements (e.g., Zn, Cu, Ni), low-  
3 solubility species can result in deficiency syndromes. Upon oxidation of the soil, sulfide is  
4 quickly oxidized, and the metals are returned to more normal equilibrium reactions of aerobic  
5 soils.

#### 7 **4.1.7. Atmospheric Behavior/Chemistry**

8 Most metals and metal compounds exist in the solid phase under ambient conditions and  
9 thus occur almost exclusively in the particle phase of the atmosphere, where they are ubiquitous.  
10 Anthropogenic sources include combustion from fossil fuels, and metal industries, as well as  
11 industrial sources employing specific metal compounds in specific processes. Some airborne  
12 metals (e.g., Mn and Ni) may derive largely from crustal sources (U.S. EPA, 1996b).  
13 Richardson (2002) included volcanic eruptions and emissions, entrainment of soil and dust,  
14 entrainment of sea salt spray, and natural forest fires as significant metals emission sources.

15 For purposes of risk assessment, particle size is important. The aerodynamic size and  
16 associated composition of particles determine their behavior in the mammalian respiratory  
17 system. Furthermore, particle size is one of the most important parameters in determining the  
18 atmospheric lifetime of particles, which may be a key consideration in assessing inhalation  
19 exposures, as well as exposures related to exposure pathways involving deposition onto soil or  
20 water (U.S. EPA, 1996b). Metals emitted by combustion processes (e.g., the burning of fossil  
21 fuels or wastes) generally occur in small particles or the fine fraction, which is often  
22 characterized by particles less than 2.5 microns in diameter (PM<sub>2.5</sub>). In contrast, the larger sized,  
23 coarse mode particles result from mechanical disruption, such as crushing, grinding, evaporation  
24 of sprays, or suspensions of dust from construction and agricultural operations. Accordingly,  
25 metals in coarse mode particles (i.e., those larger than approximately 1–3 microns) are primarily  
26 those of crustal origin, such as Si, Al, and Fe (U.S. EPA, 1996b). It is noted that the fine versus  
27 coarse distinction simply differentiates two relatively distinct size distributions of particles, the  
28 separation point of which occurs in the range of 1 to 3 μm. The distinction does not refer  
29 directly to particle sampling methods or size fractionations particular to risk assessment (U.S.  
30 EPA, 1996b).

31 Fine and coarse particles typically exhibit different behavior in the atmosphere; fine  
32 mode particles exhibit longer atmosphere lifetimes (i.e., days to weeks) than coarse particles and  
33 tend to be more uniformly dispersed across a large geographic region (U.S. EPA, 1996b).  
34 Relatively lower dry deposition velocities of fine particles contribute to their persistence and  
35 uniformity throughout an air mass (U.S. EPA, 1997c). The larger coarse particles (i.e., greater

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1 than 10  $\mu\text{m}$ ) tend to rapidly fall out of the air and have atmospheric lifetimes on the order of  
2 minutes to hours, depending on their size and other factors (U.S. EPA, 1996b).

3 In most cases, metals do not undergo transformation while in the particle phase; thus,  
4 their removal from the atmosphere is regulated by the rate at which the particles themselves  
5 participate in wet and dry deposition processes. For example, metals such as As, Be, Cd, Pb are  
6 generally found in airborne compounds with a single predominate oxidation state (As(III),  
7 Be(II), Cd(II), Pb(II)). Some metals (e.g., the transition metals Cr, Mn, and Ni) present the  
8 possibility of changing oxidation state in situ in the particle, although little is known of these  
9 processes (U.S. EPA, 2003d). This is an important consideration for risk assessment as the  
10 different oxidation states also differ in toxicity (such as for Cr).

11 For metals that can change oxidation states, much of the atmospheric chemistry takes  
12 place in the aqueous phase, such as cloud droplets or water films on particles. Metal salts and  
13 oxides that dissolve in water can undergo several reversible reactions, including hydration,  
14 hydrolysis, polymerization, and reaction with other anions. The equilibrium between these  
15 forms depends on the atmospheric conditions, the equilibrium and solubility constants, and the  
16 concentrations of other chemicals. Transformations between oxidation states can occur either to  
17 increase the oxidation state (such as oxidizing Cr(III) to Cr(VI)) or to reduce it. These oxidation  
18 or reduction reactions can occur through reaction with other species, such as dissolved metals,  
19 reduced sulfur species, and organic compounds (Seigneur and Constantinou, 1995). Although  
20 models exist that can be used to estimate metal speciation in aerosols with liquid water, the  
21 reactions are still highly uncertain.

22 Mercury is an exception among the commonly occurring metals; it exists primarily in the  
23 vapor phase under ambient conditions but can also occur in particle and aqueous phases. At  
24 least three species of mercury should be considered: elemental (Hg(0)) mercury, which is largely  
25 present as a gas; divalent (Hg(II)) inorganic mercury compounds, which are more water soluble;  
26 and particulate-phase mercury (Shroeder and Munthe, 1998; U.S. EPA, 1997b). The behavior of  
27 mercury in the atmosphere depends strongly on the oxidation state. Elemental mercury is  
28 capable of being transported long distances, even globally; divalent mercury deposits within a  
29 few hundred kilometers of sources; and particulate mercury is deposited at intermediate  
30 distances, depending on the particle size (Shroeder and Munthe, 1998). Elemental mercury that  
31 is deposited can be reemitted from the surface, as can divalent and particulate mercury after  
32 biological or chemical reduction to the elemental form.

33 In the gas phase, elemental mercury can be oxidized to divalent mercury by  $\text{O}_3$ , OH,  
34  $\text{H}_2\text{O}_2$  and molecular chlorine, although other halogen atoms might also be important (Shroeder  
35 and Munthe, 1998). In the aqueous phase, elemental mercury can be oxidized by OH,  $\text{O}_3$ , and

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1 dissolved chlorine, and divalent mercury can be reduced by processes such as reaction with HO<sub>2</sub>  
2 and S(IV). Both vapor-phase and aqueous atmospheric chemistry may involve multiple phases.

3 EPA has made a substantial effort to evaluate the atmospheric fate of mercury as a result  
4 of the requirements of the Clean Air Act. U.S. EPA (1997b) contains a comprehensive  
5 evaluation of mercury's atmospheric fate, but this is an area of ongoing research and  
6 controversy. EPA continues to be active in investigating mercury behavior in the atmosphere  
7 (e.g., Landis and Stevens, 2003; Jaffe et al., 2003; Bullock and Brehme, 2002; Bullock, 2000a, b;  
8 U.S. EPA, 2001a).

#### 9 10 **4.1.7.1. Application/Limitations**

11 Most sampling and analytical techniques published by EPA for metals in air are oriented  
12 toward evaluation of particular-phase total metals rather than metal species (U.S. EPA, 1999a).  
13 These methods involve collection of a particular size fraction of particles (e.g., PM<sub>2.5</sub>, PM<sub>10</sub>,  
14 TSP), with subsequent analysis by x-ray fluorescence, atomic absorption, inductively coupled  
15 plasma, proton-induced x-ray emission, or neutron activation analysis gamma spectroscopy  
16 techniques. The one notable exception is a method for mercury (Method IO-5) that speciates  
17 vapor and particulate forms. To the extent that metals are sorbed to particulate phases, analysis  
18 of individual metal species can, at least theoretically, be accomplished by the same techniques  
19 used to analyze those species in other solid media.

#### 20 21 **4.1.8. Metals Speciation Techniques**

22 A wide variety of analytical and chemical techniques have been used to characterize  
23 metal speciation in various media (Isaure et al., 2002; Manceau et al., 2000, 1996; Welter et al.,  
24 1999; Szulczewski et al., 1997; Charlatchka et al., 1997; Lumsdon and Evans, 1995; Ma and  
25 Uren, 1995; Hunt et al., 1992; Gupta and Chen, 1975). These techniques provide information on  
26 speciation, particle size, and the source of the metal and also quantitatively determine the metal  
27 level present. Of the techniques tested (physicochemical, extractive, and theoretical), the tools  
28 that have been used most often to evaluate speciation include the following.

#### 29 30 **4.1.8.1. Particle-Bound Metal**

31 For particle-bound metal, tools include x-ray absorption spectroscopy (XAS), x-ray  
32 diffraction (XRD), particle-induced x-ray emission (PIXE and  $\mu$ PIXE), electron probe  
33 microanalysis-scanning electron microscope (EPMA-SEM), secondary ion mass spectrometry

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1 (SIMS), x-ray photoelectron spectroscopy (XPS), sequential extractions, and single-chemical  
2 extractions.

3 Over the past decade, numerous advances in materials science have led to the  
4 development of a wide range of analytical tools for determining metal concentrations, bonds, and  
5 valences of individual particles on a scale that can be considered useful for the speciation of  
6 environmentally important materials (soils, wastes, sediments, and dust). Although most of  
7 these tools are scientifically sound and offer important information on the mechanistic  
8 understanding of metal occurrence and behavior, only a few provide currently useful information  
9 on metal bioavailability for use at a “site” level (see Table 4-9 at the end of this section).  
10 However, other techniques still may be essential for conducting a detailed characterization of a  
11 selected material to describe the chemical or kinetic factors controlling the release, transport,  
12 and/or exposure of a metal.

13 An indirect approach to speciation, in contrast to the direct methods previously described,  
14 includes functional or operational extraction techniques that have been used extensively (Tessier  
15 and Campbell, 1988; Tessier et al., 1979; Gupta and Chen, 1975). These methods use either a  
16 single or a sequential extraction procedure to release species associated with a particular metal  
17 within a medium.

#### 18 19 **4.1.8.2. *Single-Chemical Extractions***

20 These methods generally are used to determine the bioavailable amount of metal in a  
21 functional class (e.g., water soluble, exchangeable, organically bonded, Fe-Mn bound, or  
22 insoluble). In a similar approach, sequential extractions treat a sample with a succession of  
23 reagents that are intended to specifically dissolve different and less available phases. Many of  
24 these techniques are a variation on the classical method of Tessier et al. (1979), in which metals  
25 associated with exchangeable, carbonate-bound, Fe-Mn bound, organically bound, and residual  
26 species are determined. A number of excellent reviews on the use and abuse of extraction  
27 techniques are available (Beckett, 1989; Kheboian and Bauer, 1987; Forstner, 1987). These  
28 techniques can be useful in a study of metal uptake by plants and soil invertebrates, where  
29 transfer takes place predominantly from a water solution phase. However, these methods are not  
30 “selective” for metal species, and these leachable fractions have never actually been correlated to  
31 bioavailability.

#### 32 33 **4.1.8.3. *Plants***

34 When considering the bioavailability of a metal to plants from soil and sediments, it is  
35 generally assumed that both the kinetic rate of supply and the speciation of the metal to either the

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1 root or the shoot are the most important factors. In soils and sediments, there is generally a small  
2 volume of water in contact with the chemical form of the metal. Although the proportion of  
3 soluble metal in pore water is small when compared with the bulk soil/sediment metal  
4 concentrations, it is this phase in pore water that is directly available to plants at the root tips.  
5 Therefore, understanding pore water chemistry is critical; that is, measuring metal concentrations  
6 as simple inorganic species, organic complexes, or colloid complexes is most important. Tools  
7 currently used for metal speciation in plants include the following:

- 8
- 9 • *In situ* measurements using ion selective electrodes (Wehrli et al., 1994; Gundersen et  
10 al., 1992; Archer et al., 1989).
- 11
- 12 • *In situ* collection techniques using diffusive equilibrium thin films and diffusive  
13 gradient thin films followed by laboratory analyses (Zhang et al., 1995; Davison et  
14 al., 1994, 1991; Davidson and Zhang, 1994).
- 15
- 16 • Equilibrium models (WHAM/FREEQC/MINTEQA2).
- 17

#### 18 **4.1.9. Organo-Metals/Metalloids Transformation Processes**

19 Metals/metalloids can exist in the environment in several valence forms and as  
20 organometallic compounds. *Organometallic compounds* (referred to in this section collectively  
21 as “*organometallics*”) are compounds that have a metal/metalloid-carbon bond. The bonds in  
22 organometallic compounds are generally covalent and between soft metals and soft ligands.  
23 Metal/metalloid transformation processes, such as metal methylation, occur through interactions  
24 with other chemicals and biota in the environment. Cycling and distribution of organometallic  
25 compounds between terrestrial, water, and atmospheric phases may be physically, chemically, or  
26 biologically mediated. Examples of some commonly occurring environmentally stable  
27 organometallic compounds are shown in Table 4-6.

28

29

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**Table 4-6. Some stable organometallic compounds**

<b>Metal/metalloid</b>	<b>Stable organometallic compound</b>
Arsenic	Methylarsenic acid, dimethyl arsenic acid, trimethyl arsine, trimethylarsine oxide
Lead	Tetramethyl/ethyl lead, trimethyl/ethyl lead, dimethyl/ethyl lead
Mercury	Methyl mercury, dimethyl mercury
Selenium	Dimethyl selenide, dimethyl diselenide, seleno-amino acids
Tin	Tributyltin, bis(tributyltin) oxide

Environmental methyl-metal concentrations reflect the net methylation rather than simple rates of methyl-metal synthesis. Metal methylation and demethylation rates in ecosystems are influenced by the speciation and biochemical availability of the metal. Metals involved in abiotic or biotic methylation/demethylation processes are presented in Table 4-7. With the exception of arsenic and selenium, the metals listed in the table form stable complexes with either methyl or ethyl groups. In addition to methyl/ethyl compounds, stable organometallic compounds such as lipids and arsenic and amino acids and selenium are incorporated biochemically.

**Table 4-7. Metals/metalloids involved in methylation processes**

<b>Process</b>	<b>Metals affected</b>
Environmentally stable organometallics	Si, Ge, Sn, Pb, Hg, As, Sb, Se
Abiotic chemical methylation	Hg, Pb, Sn
Abiotic demethylation	Sn, Pb
Biotic methylation	As, Cd, Hg, Pb, Se, Sn, (others? Sb, Pt, )
Biotic demethylation	As, Hg, Sn, Pb

Source: Bodek et al. (1988).

Organometallic environmental transformations may affect both the mobility and the toxicity of these metals. *The rates of transformation and the organometallic products are dependent on environmental conditions and the population of microorganisms available.* For example, methylation/demethylation rates are dependent on the speciation of the metal, the microbial community, the environmental variables (e.g., pH, temperature, reduction oxidation potential, organic matter, dissolved oxygen, nutrient availability, salinity, complexing agents) and the distribution of the metal between compartments (sediment, water, gaseous). The inter-

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1 relatedness of these processes has made research into unraveling the factors controlling net  
2 methylation difficult and, to date, incomplete. However, some general trends can be predicted  
3 with some certainty, and these are discussed in this section.

#### 5 **4.1.9.1. Abiotic Transformations**

6 Organometallic compounds that are composed of metals with electronegativities  $>1.7$  are  
7 the most stable under environmental conditions. Carbon-metal bonds with more polar (metal  
8 electronegatives  $<1.7$ ) bonds will undergo hydrolysis (reaction with water). *Abiotic chemical*  
9 *methylation* can occur by three mechanisms: transmethylation reactions between mercury and  
10 tin/lead alkyls, humic/fulvic substances, and photochemical reactions.

#### 12 **4.1.9.2. Biotic Methylation Transformations**

13 Biotic methylation occurs when organisms, primarily microorganisms, transfer alkyl  
14 groups to bioavailable metals. In general, it is thought that anaerobic sulfate-reducing bacteria  
15 are the principal methylators in freshwater and estuarine environments. However, methylation  
16 rates are not always correlated with sulfate-reducing bacteria. Not all sulfate-reducing bacteria  
17 are capable of methylating, and efficiency of methylation is dependent on the activity and  
18 structure of the bacterial community. Other bacteria may be involved in methylation. Biotic  
19 methylation occurs predominantly in the sediment column; however, because the water column  
20 by volume is much larger, water column methylation is important.

21 Maximum methylation rates typically occur at the redox boundary, which varies  
22 seasonally and frequently coincides with the sediment-water interface (Ullrich et al., 2001).  
23 Methylation rates decrease with increasing sediment depth, probably due to a decrease in biotic  
24 habitat. Microorganisms may also demethylate (or dealkylate) organometallic compounds.  
25 Microbial-mediated transformations are frequently the most important environmental  
26 organometallic processes. Generally, as the amount of organic material increases in a system the  
27 microbial populations also increases. Examples of typical bacterial populations in natural waters  
28 and sediments are shown below (Ullrich et al., 2001).

29 High temperatures and anaerobic conditions generally favor metal-methylation  
30 formation, and demethylation processes are generally favored under low temperatures and/or  
31 aerobic conditions. Studies on the effects of pH are not consistent. Interconnected parameters  
32 include pH effects on the microbial communities and effects on the speciation distribution of the  
33 metals/metalloids in the water and the sediment as well as adsorption rates. Organometallic  
34 compounds appear to increase in the water column in low pH environments, but this may be due

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1 to release of methylated metals from the soil and subsequent depletion of organometallic  
2 compounds in the soil. Therefore, pH effects on net methylation in a system are not fully  
3 understood. In freshwater ecosystems, where sulfate concentrations are typically low, increase  
4 in sulfate concentration increases methylation rates. However, in reducing environments,  
5 increasing sulfide concentration decreases methylation rates.

6 The inhibitory effect of sulfide is probably not due to metal sulfide formation but, rather,  
7 to the formation of less bioavailably charged metal-sulfur complexes. High organic matter may  
8 increase abiotic methylation through humic/fulvic metal reactions; however, this mechanism is  
9 poorly understood and confounded because biotic methylation rates may increase in  
10 environments with high organic matter. In ecosystems with high DOC concentrations, DOC  
11 may bind with metals/metalloids, rendering them unbioavailable and thereby reducing biotic  
12 methylation rates.

#### 14 **4.1.9.3. Organometallic Transformation**

15 Organometallic methylation and demethylation rates are influenced by both speciation  
16 and bioavailability of the metal, the microbial community, and a large number of environmental  
17 factors, many of which are interrelated. Sulfide and organic matter are important environmental  
18 variables that significantly affect methylation; however, their effect on  
19 methylation/demethylation is as yet poorly understood. Which variables dominate differs among  
20 locations and between seasons, although it is clear that methylation is predominantly a  
21 biologically mediated process. Methylation/demethylation rates are strongly influenced by the  
22 metal/metalloid speciation and bioavailability. General trends in methylation/demethylation  
23 rates are outlined in Table 4-8.

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**Table 4-8. General trends of environmental factors affecting rates of methylation/demethylation**

Organometallic transformations	Temperature		pH		SO <sub>4</sub> <sup>2-</sup>	Organic matter	Redox		Salinity
	High	Low	High	Low	High		Oxic	Anoxic	High
Net methylation	↑	↓	?	?	?	?	↓	↑	↓
Methylation aq	↑	↓	↓?	↑?	↓	↓↑	↓	↑	↓
Methylation sed	↑	?	↑	↓	↓	?	?	?	?
Demethylation	↓	↑	↓	↑	?	?	↑	↓	?

↑ indicates an increase in rate.

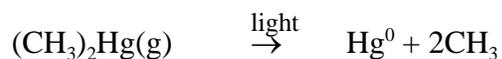
↓ indicates a decrease in rate.

? indicates conflicting data or insufficient data to indicate a likely trend.

#### 4.1.9.4. Atmospheric Transformations

**4.1.9.4.1. Abiotic chemical methylation/demethylation transformations.** Boiling points for organometallics for a given metal/metalloid decrease with increasing alkyl substitution and with shorter alkyl chains. For example, the boiling points of organotin compounds decrease with dimethyltin dichloride > trimethyltin > tetramethyltin. Fully methylated metals such as dimethyl mercury may be transported great distances in the atmosphere owing to the combined low boiling point and low water solubility. Methylation of Se, Hg, Pb, and As volatilizes these compounds, contributing to their air concentrations.

Demethylation of organometallic compounds in the atmosphere occurs by primarily by photolysis, such as:



Demethylation may also occur by reaction with ozone, hydroxyl radicals, nitrate radicals, and sorption to particulate matter. Organometallic compounds are also removed from the atmosphere by wet/dry deposition.

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1 **4.1.9.5. Aquatic Transformations**

2 **4.1.9.5.1. Abiotic chemical methylation/demethylation transformations.** In stratified aquatic  
3 systems, methyl-metal/metalloid formation occurs predominantly at the oxic/anoxic interface.  
4 Organosiloxanes and other silicone-related substances have been considered as possible abiotic  
5 methylating agents. *Overall, abiotic methylation is probably of minor importance except in*  
6 *ecosystems with high organic matter.*

7  
8 **4.1.9.5.2. Biotic Methylation/Demethylation Transformations.** Microbial methylation  
9 processes play the major role in methylation of Hg, Sn, and As, with methylcobalamin the most  
10 likely environmental methyl donor. Metal speciation is a prime factor regulating the methylation  
11 potential in a system. Until recently, Hg<sup>2+</sup> was considered the main mercury species methylated  
12 by bacteria; however, current research indicates that uncharged Hg complexes are more likely  
13 the principal species methylated (Ullrich et al., 2001). Arsenate can be reductively methylated  
14 (via arsenite) under anoxic conditions to dimethylarsine. Selenium and selenite can be  
15 methylated via microorganisms; demethylation of organoselenium via other biotic processes is  
16 also known.

17  
18 **4.1.9.5.3. Environmental Factors affecting methylation.** Seasonal variation of methylated  
19 mercury is thought to be related to temperature, redox potential, and productivity. Seasonal  
20 variation for organic arsenic (dimethylarsenic) has also been reported, with organoarsenic  
21 species decreasing in late fall and winter.

22  
23 **4.1.9.6. Terrestrial Transformations**

24 **4.1.9.6.1. Abiotic chemical methylation/demethylation transformations.** Methylation and  
25 demethylation of organic mercury compounds in soils appear to be mediated by the same types  
26 of abiotic and microbial processes that occur in aquatic systems. The frequency and magnitude  
27 of soil moisture play an important role in availability and transformation processes. Because  
28 soils are primarily oxygenated systems, particularly in the root zone, conditions favorable to  
29 sulfide formation and bacterial methylation occur infrequently. With the exception of peat bogs  
30 and similar anoxic, highly saturated soils, methylation generally occurs only at very low rates in  
31 soils.

32  
33 **4.1.9.6.2. Biotic methylation/demethylation transformations.** Plants have the capacity to  
34 transform metals and metalloids that are taken up from the soil. The most notable example is

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1 selenium. Soluble inorganic oxyanions of selenium are readily taken up by plants and converted  
2 to organoselenium compounds, such as selenomethionine, selenocysteine, dimethyl selenide, and  
3 dimethyl diselenide. Demethylation/dealkylation of organoselenium (e.g., trimethylselenonium),  
4 organomercury, organoarsenic, and organotin can occur in soil.

5  
6 **4.1.9.6.3. Environmental factors affecting methylation.** Formation and degradation of  
7 organometallic compounds in soils appears to be mediated by many of the same types of  
8 microbial and environmental processes as in aquatic systems. Speciation of metals/metalloids  
9 dominates the methylation and/or uptake. Methylation/demethylation rates are affected by soil  
10 moisture: low moisture decreases biotic processes in soils. Soils high in iron and aluminum  
11 oxide, silts, and clay minerals interfere with methylation of metals/metalloids owing to the  
12 reduced bioavailability of metals/metalloids. Plants methylate selenium, predominantly to  
13 selenomethionine and some selenocysteine. Plant uptake and methylation of selenium or arsenic is  
14 specific to plant species. Many soil organisms are capable of converting arsenate/arsenite to  
15 volatile methylated arsines. Losses of 15–30% per year due to volatilization of arsenic in soil  
16 have been reported (ATSDR, 2003). Organolead complexes, on the other hand, are thought to be  
17 relatively stable in soils.

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**Table 4-9. Metal speciation techniques for solids and their associated pore waters**

Tools	Species lability	Species particle size	Species valance state	Species bonding	Species composition	Species abundance	Element specificity	Isotopic character	Element sensitivity	Resolution	Availability	Cost
XRD	No	No	No	No	No <sup>a</sup>	No	No	No	3–4 vol%	Bulk	1	\$
EMPA/SEM	Yes	Yes	Yes <sup>b</sup>	No	Yes	Yes <sup>c</sup>	B-U	No <sup>d</sup>	100 ppm	0.5–1 μ	2	\$\$
SIMS	No	Yes	No	No	Yes <sup>e</sup>	Yes <sup>f</sup>	Li-U	Yes	1 ppb	10 μ	4	\$\$\$
XPS	No	No	Yes	Yes	Yes <sup>e</sup>	Yes <sup>f</sup>	H-U	No	wt.%	100 μ	2	\$\$
XAS	No	No	Yes	Yes	Yes <sup>e</sup>	Yes <sup>f</sup>	He-U	No	ppb	2 μ	5	\$\$\$\$
PIXIE	No	No	No	No	Yes	Yes <sup>f</sup>	B-U	No	10 ppm	4 μ	4	\$\$\$\$

<sup>a</sup>Identifies crystalline compounds and stoichiometric compositions only.

<sup>b</sup>Valence determined by charge balance of complete analyses.

<sup>c</sup>Technique has limitations based on particle-counting statistics.

<sup>d</sup>Limited when combined with ICP/MS/LA.

<sup>e</sup>Technique requires that each element be tuned and standardized, requiring unreasonable time limits.

<sup>f</sup>Techniques designed and tested only on simple systems. Multiple species require lengthy analytical and reduction.

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1 **4.2. HUMAN HEALTH EXPOSURE PATHWAY ANALYSIS**

2 **4.2.1. Introduction**

3 This section and Section 4.3 discuss issues of importance when performing human health  
4 risk assessments for metals and metal compounds. The information provided will complement  
5 other general Agency guidance on the risk assessment process (e.g., Carcinogen Risk  
6 Assessment (U.S. EPA, 2003b), Exposure Assessment (U.S. EPA, 1992b), Developmental  
7 Toxicity (U.S. EPA, 1991b) and focuses on the unique and specific characteristics of metals and  
8 metal compounds that might be applied in metals risk assessments for human health. This  
9 section provides some of the scientific basis that underlies metal-specific characteristics of  
10 human health effects assessment, but it is not intended to be comprehensive. Appropriate and  
11 sufficient reference material provided in this framework document will guide readers to  
12 additional details on any of the topics addressed.

13  
14 **4.2.2. Human Exposure**

15 Assessment of human exposures to any chemical agent includes (1) identifying how  
16 people come into contact with metals in the environment; (2) determining the concentrations of  
17 specific forms (speciation) of the metal in specific media (e.g., soil, water, air, and biota); (3)  
18 identifying the pertinent exposure metric (via consideration of dose-response assessment); (4)  
19 estimating the exposure metric (e.g., oral intake, inhalation exposure concentration, blood  
20 concentration), which may involve quantifying relationships between exposure concentrations  
21 and intakes; and (5) identifying sources of uncertainty and natural variability and, where  
22 possible, quantifying these in estimates of exposure. Although these components are common to  
23 exposures to human and nonhuman receptors, and to metal as well as nonmetal toxicants, some  
24 specific aspects of human metal exposure assessment are discussed below.

25  
26 **4.2.2.1. Environmental Background Concentrations**

27 For assessments performed to assess impacts associated with particular human activities  
28 (e.g., hazardous waste sites, environmental releases), the term “ambient background” generally  
29 refers to all other sources of the metals of interest. The contribution of the background to human  
30 metal exposure may be significant. For example, metals are natural components of the  
31 environment and are repeatedly cycled throughout the biosphere (this component of the ambient  
32 background is referred to as “natural background”). Metals are also present as background from  
33 persistent anthropogenic activities. During the early 1970s, for instance, industrial sources

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1 accounted for more than 90% of the airborne lead, the deposition of which supplied a large  
2 fraction of the lead in many ecosystems (Nriagu and Pacyna, 1988). Various strategies are  
3 available to estimate background metal concentrations in environmental media (soil, water, and  
4 sediment) (U.S. EPA, 1995a, b; 1989b), and a number of documents dealing with the  
5 background concentrations of metals in soils have been published by the U.S. EPA (2002b, c, d;  
6 2001b, c).

7 Dietary pathways account for the major impact of background sources of metals. A  
8 number of sources provide information on dietary exposure background for metals (Capar and  
9 Cunningham 2000; Schoof et al., 1999a, b; Thomas et al., 1999; Bolger et al., 1996; Dabeka and  
10 McKenzie, 1995; Gunderson 1995; Tsuda et al., 1995; Dabeka et al., 1993). The prominence of  
11 diet in exposure assessment is discussed by Thomas et al. (1999). The United Kingdom also has  
12 extensive archives of metal content of beverages and infant foods  
13 (<http://archive.food.gov.uk/maff/archive/food>). Additional human dietary exposures may occur  
14 secondary to commercial processing of foods (e.g., use of preservatives, emulsifiers, taste  
15 enhancers, and packaging products that contain metals). Products such as liquid diets for weight  
16 loss, infant formula, and supplements for geriatric patients contain metals.

17 Exposure to metals in the diet can be high enough to approach or even exceed  
18 occupational or other well-known exposures to the metal (e.g., arsenic in drinking water). Thus,  
19 biomarkers (e.g., urinary arsenic, blood mercury) should take diet into account when estimating  
20 exposure. For example, in the 24 hours following ingestion of a seafood meal, the urinary  
21 arsenic concentration can often rise to 1 mg/L (WHO/IPCS, 1981). In contrast, persons living in  
22 Taiwan in an area with endemic arsenic contamination of the water supply of 50–300 µg/L had  
23 urinary arsenic concentrations of 140 µg/L (WHO/IPCS, 2001). Workers at a copper smelter  
24 where there was considerable arsenic exposure were found to have urinary arsenic  
25 concentrations in the range of 200–600 µg/L (WHO/IPCS, 2001). In a study of 380 American  
26 dentists, Brady et al. (1980) reported a mean concentration of 8.5 µg per litre, 7.4% of the  
27 participants having blood mercury levels greater than 15 µg/L (WHO/IPCS, 1991). In contrast,  
28 median cord blood mercury concentrations in a cohort with high fish consumption in the Faroe  
29 Islands was 24 µg/L (NRC, 2000). Mercury concentrations in the blood of people with long-  
30 term exposure to methyl mercury from fish were reported to be as high as several hundred µg/L  
31 (WHO/IPCS, 1990a).

32 Lifestyles expose people to metals in many different contexts and contribute to ambient  
33 background and total exposures. These exposures occur in the workplace, in nutritional

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1 supplements, in pharmaceuticals taken by the oral and injection routes, in medical diagnostic  
2 procedures, in cosmetics, in recreational drugs (e.g., smoking), in folk medicines, and in paints  
3 and pigments. For example, cadmium from tobacco smoking can significantly affect a person's  
4 total exposure.

5  
6 **4.2.2.1.1. Application.** Depending on the scope, purpose, and approach employed for the  
7 exposure assessment, background may be considered in different ways. For example, in the case  
8 of predicting exposure estimates from information on environmental releases of interest,  
9 background estimates may be included as well, and depending on the methodology, the risk  
10 estimates may be developed for the aggregate exposure as well as for the different sources of  
11 interest, including background. In this approach, differences in bioavailability among the  
12 various metal sources should be considered and incorporated into the calculation of total  
13 exposure. Alternatively, biomarkers may be used for exposure assessment, which already  
14 account for bioavailable fractions. Therefore, while it may not be feasible to partition out  
15 contributions specific to the pollutant source or exposure pathway of interest, biomarkers will  
16 provide an estimate of aggregate exposure.

17  
18 **4.2.2.1.2. Limitations.** The approach taken for background may depend on the dose-response  
19 relationship for the health endpoint of interest. In the case of cancer assessments, where the  
20 Agency has derived a probabilistic relationship and the risk management decision framework is  
21 particular to the incremental risk of the pollutant source or human activity of interest, the basic  
22 background evaluation can simply focus on providing the foundation for that incremental  
23 context. Yet for many other health endpoints, the Agency's traditional approach for toxics  
24 involves the Reference Dose (RfD) or Reference Concentration (RfC) approach, which may be  
25 conceptually described as an uncertainty bound on a No-Observed-Adverse-Effect Level  
26 (NOAEL). The NOAEL by itself provides no information about the slope of the dose-response  
27 relationship or, therefore, the incremental change in risk expected for an incremental increase in  
28 exposure above the background level. Thus, in those cases, it will be more important for the  
29 exposure assessment to consider estimates of total, rather than incremental, exposures.

#### 30 31 **4.2.2.2. Air Pathways of Exposure**

32 Air pathway inhalation is the major pathway for human intake of metals in which air  
33 serves as the primary medium of contact. Indirect pathways in which air serves as an antecedent

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1 medium also include the following: deposition of metals to surface dusts and subsequent intake  
2 from ingestion or inhalation; deposition to surface water and sediment and intake from ingestion;  
3 and uptake of deposited metals into or onto aquatic and/or terrestrial biota, entrance into the  
4 human food chain, and intake from ingestion. Both systemic bioavailability and local actions on  
5 nasal mucosa, airways, and lung tissue should be taken into account when considering human  
6 health effects incurred via the air pathway.

7  
8 **4.2.2.2.1. Applications.** Most airborne metals, with a few important exceptions (e.g., mercury  
9 and arsine) occur in particulate form, which necessitates certain considerations for inhalation  
10 exposure assessment. For example, particle size affecting respirability (i.e., how much of the  
11 pollutant enters the respiratory system). Additionally, inhalation dosimetry for particles involves  
12 some distinctly different processes than for gases (i.e., deposition, clearance, dissolution, etc.),  
13 which are also influenced by particle size (U.S. EPA, 1997c, as revised in 2004b). Particle size  
14 is thus an important factor in assessing metals exposure, with the focus generally being on  
15 particles less than or equal to 10 microns ( $\mu$ ) in diameter ( $PM_{10}$ ). Larger particles do not  
16 generally penetrate far into the respiratory tract and can be cleared to the ingestion route. They  
17 may play a larger role in irritant and other effects on eyes and nasal passages, and if deposited in  
18 the uppermost reaches of the respiratory tract may be transferred to the digestion tract. Thus, for  
19 exposure assessments involving measurements (e.g., using area or personal samples), the size of  
20 particles sampled is an important consideration.

21 For metals for which the Agency has developed RfCs and IURs, the exposure estimate  
22 should be for the form of metal used in the dose-response assessment that established the  
23 reference values (e.g., exposure concentrations, usually with focus on the respirable fraction)  
24 (U.S. EPA, 2004c). For more information on the consideration of particle size in the dose-  
25 response assessment for RfCs and IURs, refer to U.S. EPA, 1990. For metals for which the  
26 Agency has developed alternative dose-response metrics (e.g., blood lead concentration),  
27 respirability, deposition, and clearance as well as absorption into the circulatory system may  
28 need to be addressed as part of the exposure assessment.

29  
30 **4.2.2.2.2. Limitations.** In developing inhalation exposure estimates, attention should be given to  
31 the form of the metal pertinent to the dose-response assessment (e.g., RfC, IUR). Simply  
32 measuring the total amount of a metal without regard to speciation may introduce uncertainties  
33 into inhalation exposure estimates, as it can with all exposure routes. Metal speciation affects a

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1 range of processes that change how the metal is deposited in the respiratory tract and  
2 subsequently distributed throughout the body and, consequently, its potential toxicity (Bailey  
3 and Roy, 1994; Oberdoerster, 1992). For example, in assessing the risk of inhaled chromium,  
4 the assessor should consider speciation (e.g., Cr+3 vs. Cr+6), as the dose-response assessment  
5 includes that specification. The bioavailability of metals via inhalation can be much higher than  
6 that of other routes of intake. This may result in relatively high internal doses, even when  
7 intakes are similar to those from other routes. An example is the large contribution made by  
8 cigarette smoking to the body burden of cadmium (e.g., Friis et al., 1998; Ellis et al., 1979).  
9 Variations in airway structure and respiratory conditions (e.g., as with age) may alter the  
10 deposition pattern of inhaled particles and contribute to variations in bioavailability (James,  
11 1994; Xu and Yu, 1986; Phalen et al., 1985).

#### 12 13 **4.2.2.3. Dust and Soil Pathways of Exposure**

14 Surface dusts and soil are particularly important media of human contact with metals.  
15 Both serve as long-lasting repositories for airborne metal particles, and soils also contain metals  
16 due to direct contamination from runoff or mixing with solid wastes. Humans are exposed to  
17 metals in surface dust and soil primarily through incidental ingestion or inhalation of suspended  
18 dust particles. Dermal contact with metals in soil represents another potential route of exposure,  
19 but the relatively low lipid solubility of most metals generally limits absorption through the skin  
20 (Paustenbach, 2000; Hostynek et al., 1998). Few studies have actually attempted to quantify the  
21 extent or kinetics of dermal penetration of metals deposited on the skin, and the applicability of  
22 these studies to metal species and complexes that occur in surface dust or soil is highly  
23 uncertain. Therefore, this exposure route is of lesser importance for most metal assessments than  
24 the ingestion and inhalation exposure routes.

25 Infants and children are particularly vulnerable to exposures to metals through the surface  
26 dust pathway because (1) their crawling and play activity put them in close proximity to surface  
27 dust, and (2) they often mouth their hands (e.g., finger sucking) and objects in their environment.  
28 This causes intakes of surface dust that are generally greater than those normally found in adults  
29 (e.g., Barnes, 1990). The amount of soil ingested by children can be expected to vary with  
30 numerous factors, including age, activity patterns, and accessibility to soil and dust. Data are  
31 limited with regard to distinguishing between the quantity of dust ingested and the quantity of  
32 soil ingested. This parameter is important in connecting measured soil metal concentrations with  
33 surface dust ingestion that occurs in the indoor and outdoor environments (U.S. EPA, 1994b).

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1 Exposure assessment methods for direct soil ingestion is described in the Risk Assessment  
2 Guidance for Superfund (RAGS) (U.S. EPA, 1989b;  
3 <http://www.epa.gov/superfund/programs/risk/ragsa/index.htm>). Additional guidance with  
4 respect to children (e.g., amount of soil a child may ingest) may be found in the *Child-Specific*  
5 *Exposure Factors Handbook* (U.S. EPA, 2002e).

6 Few studies of soil ingestion in adults have been conducted; however, the estimates  
7 support the general assumption that average daily soil ingestion rates of adults who do not  
8 participate in activities in which intensive exposure to surface dust and soil occur (e.g.,  
9 occupational gardening, construction work) are lower than those of children (Calabrese et al.,  
10 1990; Hawley, 1985). Because concentrations of the metal contaminants in soil can be expected  
11 to vary with depth, exposure assessments should consider soil metal concentrations at the depth  
12 appropriate to the metal(s) of concern and human behaviors and activities. The size of metal-  
13 bearing particles also varies with depth. For example, higher concentrations of lead and smaller  
14 particles are found near the soil surface (Duggan and Inskip, 1985; Duggan et al., 1985;  
15 Fergusson and Ryan, 1984). For review, see Chaney et al. (1988). Because bioavailability of  
16 lead and other metals increases with decreasing particle size (U.S. EPA, 2004a; Barltrop and  
17 Meek, 1979), both particle size and depth of contamination become very important  
18 considerations in metal exposure assessments.

#### 19 20 **4.2.2.4. Dietary Pathway**

21 Food can be a major contributor to human metal exposures from contaminant point  
22 sources as well as containing trace amounts of naturally occur metals. Failure to accurately  
23 account for the dietary contribution can result in significant errors in exposure and risk estimates  
24 for metals (Choudhury et al., 2001). Human dietary exposures to metals may occur through  
25 various pathways. Livestock grazing in metal-contaminated areas can take up metals from soils  
26 or surface water. Metals also can migrate into surface water and sediments and can be taken up  
27 by aquatic organisms that are consumed by humans. Human food crops also can take up metals  
28 from soils and surface water or become contaminated with metals through deposition of airborne  
29 particles. Metals can enter food during harvesting or processing of produce and livestock, during  
30 food storage from metals in food containers, and during preparation of foods for meals.

31  
32 **4.2.2.4.1. Application.** Estimation of intakes of metals in food requires information or estimates  
33 on the levels of the metal in food and the amount of food consumed. Although large-scale

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1 surveys of the metal contents of foods and food consumption patterns have been conducted (e.g.,  
2 Egan et al., 2002; Ryan et al., 2001; U.S. FDA, 2001; O'Rourke et al., 1999; Thomas et al.,  
3 1999; U.S. DHHS, 1996), these surveys have several limitations for applications to risk  
4 assessment at a contaminated site. Analysis is often conducted with "market basket" samples of  
5 packaged processed foods. With few exceptions, such applications have not been empirically  
6 evaluated against biomarkers of exposure (Clayton et al., 2002, 1999; Choudhury et al., 2001).  
7 For some risk assessments, multimedia fate and transport models may be used to predict  
8 concentrations in locally grown or raised foods or wildlife consumed by the population of  
9 interest.

10  
11 **4.2.2.4.2. Limitations.** Because some metals (e.g., cadmium and lead) could have long  
12 residence time in the body, reconstructing historic dietary exposures can be a challenge. Food  
13 consumption surveys generally are limited to short-term consumption (e.g., 1 to 3 days) and do  
14 not capture intra-individual variability that would affect long-term averages. Furthermore, food  
15 consumption patterns can be expected to change over time; thus, patterns discerned at any given  
16 time may not accurately represent historical exposures. An additional challenge is the  
17 integration of data from separate metal residue surveys and food consumption surveys (e.g.,  
18 Tomerlin et al., 1997). This leads to considerable uncertainty in estimates of metal exposure  
19 through the dietary route.

20 Furthermore, estimates of dietary intakes of metals in food based on national or regional  
21 data cannot be expected to accurately reflect intakes of metals in locally harvested foods owing  
22 to differences in natural background levels of metals. This can be a particularly important  
23 limitation when the receptors of concern are subsistence fishermen or hunters. These surveys  
24 also may not accurately reflect the amount of metals that enter the food pathway during local  
25 food preparation or storage (e.g., in the home).

#### 26 27 **4.2.2.5. Water and Sediment Pathway**

28 Part of the human population obtains drinking water from untreated sources, such as  
29 wells. However, people also consume water specifically treated for human consumption.  
30 Treatment of ambient water for human consumption removes DOC and suspended organic  
31 sediments that can form complexes with metals (AWWA, 1999). Thus, the exposure context of  
32 metals in human drinking water will be very different from that of ambient water. That is,  
33 inorganic forms of metals in treated drinking water will consist of the more bioavailable, water-

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1 soluble species. Treatment also removes bacteria that can participate in organification reactions  
2 of toxicological significance to humans (e.g., methylation of inorganic mercuric mercury).

3  
4 **4.2.2.5.1. Applications.** Estimation of the intake of metals in drinking water requires  
5 information about or estimates of concentrations of metals in the water and the amount of water  
6 consumed. Data on the metal content of tapwater can be obtained from EPA's Office of  
7 Drinking Water (U.S. EPA, 2000d). EPA's *Exposure Factors Handbook* contains exposure  
8 information on daily drinking water ingestion and incidental ingestion of water during swimming  
9 and showering (U.S. EPA, 1997h). People also can be exposed to metals dissolved in ambient  
10 surface water or in association with suspended sediments; the latter can serve as a long-term  
11 repository for waterborne metal particles. Such exposures occur during swimming or other  
12 recreational activities as a result of incidental water ingestion or during occupational activities in  
13 which the sediments are disturbed or resuspended in the water column. Metal bioavailability in  
14 ambient surface water can be expected to be much more diverse than in treated drinking water or  
15 in ground water because of the presence of organic carbon, inorganics, and suspended organic  
16 material that can serve as ligands or reactants for metals. Speciation and concentration will also  
17 vary with pH of the surface water. Therefore, estimation of intake of metals from surface water  
18 will require appropriate adjustment for relative bioavailability.

19  
20 **4.2.2.5.2. Limitations.** Metals can enter treated drinking water at various stages of water  
21 treatment, distribution, or delivery. Generally, water metal concentrations are measured at the  
22 distribution point for municipal water delivery systems. Distribution systems within homes  
23 (pipes, storage containers, etc.) and, in the case of lead, glassware, can contribute metals to the  
24 water (Graziano et al., 1996). The contribution of metals from pipes (either from the distribution  
25 system to the home or within the home) is rarely assessed. This can be highly variable, both  
26 within the system and temporally; water that remains in pipes overnight frequently has a higher  
27 metal load than water used during the day. Furthermore, the concentration of organic material  
28 and other ligands in the water may vary across drinking water sources and can affect the  
29 bioavailability of the metal. There is also wide variability in the form of metals in ambient  
30 surface water. These factors can be incorporated into site-specific assessments, but local data  
31 should be collected on a case-by-case basis. Inhalation of inorganic metal contaminants in water  
32 can result from aerosolization, or in special cases, from volatilization (e.g., Hg<sup>0</sup>). The  
33 aerosolization pathway can be a major source of intake of inorganics under certain conditions.

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1 As an example of aerosol exposure, sea spray can be a significant contributor to iodide intakes in  
2 populations that live near the seashore (Whitehead, 1984). A pathway more typical of human  
3 exposure to contaminated water is showering, in which aerosolization can occur at the water tap.  
4 Although models have been developed to predict human inhalation exposures to volatile  
5 organics from showering (e.g., Moya et al., 1999; McKone, 1987) (see Guo, 2002, for review),  
6 comparable models do not exist for aerosolized metals (Wilkes, 1998), and the magnitude of  
7 exposure from showering is unknown.

### 8 9 **4.2.3. Routes of Entry**

10 The most frequently encountered routes of entry of metals into humans are ingestion and  
11 inhalation. The dermal route is of less concern for most metals but can be important for skin  
12 toxicity of some metals (e.g., nickel, chromium). Other routes can be important in specific  
13 circumstances, for example, explosions of metal-bearing materials can result in intra- or trans-  
14 dermal exposures (Robinson et al., 1983). It is also important to remember that metals can  
15 produce toxicity at the point of entry. While metal absorption from skin may be minimal, dermal  
16 irritation and sensitization can occur without absorption. Similarly, human health effects such as  
17 lung disease and lung cancer can occur from inhalation exposures that do not result in substantial  
18 systemic uptake. Routes of entry do not necessarily correspond to the expected exposure  
19 pathway. For example, uptake of airborne metals associated with larger particles can actually be  
20 attributed to ingestion of surface dust rather than absorption from the respiratory tract. The  
21 significance of this process relative to other pathways will depend on the exposure scenario  
22 generally being less significant than diet or soil ingestion pathways (see sections 4.2.1 and  
23 4.2.2.2). Thus, uptake in the lung can be the source of exposure by two routes.

24 Although dermal contact with metals occurs through soil, air, and water pathways, the  
25 relatively low lipid solubility of most metals limits absorption through the skin (Paustenbach,  
26 2000; Hostynek et al., 1998). An exception is Hg<sup>0</sup>, which is dermally bioavailable (Hursh et al.,  
27 1989). In general, empirical information on dermal absorption of metals should be consulted  
28 when available (Stauber et al., 1994; Hostynek et al., 1993; Wester et al., 1992; Hursh et al.,  
29 1989; Ilyin et al., 1975).

### 30 31 **4.2.4. Integrated Exposure Approaches**

32 Approaches to integrating exposure across pathways and physiological routes of uptake  
33 include modeling, relative bioavailability estimates, and biomarker assessment.

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#### 1 **4.2.4.1. Modeling**

2 Few specific exposure models have been developed for metals with the exception of lead.  
3 The Integrated Exposure Uptake Biokinetic (IEUBK) model for lead in children (White et al.,  
4 1998; U.S. EPA, 1996c, 1994b) was specifically developed for translating exposure  
5 measurements into risk estimates at sites contaminated with lead. The IEUBK model and  
6 background documentation are available on line at  
7 <http://www.epa.gov/superfund/programs/lead/ieubk.htm>. The IEUBK model assumes that 10  
8  $\mu\text{g}/\text{dL}$  is a no effect level, while current science indicates there may be no safe level of lead  
9 exposure. The model is not readily generalized to other metals because blood lead levels are  
10 used as the dose metric, and dose-response assessment for most other metals use estimates of  
11 oral or inhalation intake rates.

12 A stochastic human exposure model for lead that is linked to a lead pharmacokinetics  
13 model (O'Flaherty et al., 1995) has also been developed (Beck et al., 2001). Less complex  
14 models linking adult exposures and blood lead concentrations are available as well (Carlisle,  
15 2000; Stern, 1996, 1994; U.S. EPA, 1996c; Bowers et al., 1994; Carlisle and Wade, 1992).  
16 These models have not been reviewed by the Agency authors. An exposure model for arsenic  
17 has also been reported (Cohen et al., 1998).

18 Other general exposure models used in risk assessment can be potentially applied to  
19 metals, including population-based models. The EPA's Stochastic Human Exposure and Dose  
20 Simulation (SHEDS) model is a probabilistic, physiologically based model that simulates  
21 aggregate human exposures and doses (i.e., via inhalation, dietary, dermal, and nondietary  
22 routes) for population cohorts and multimedia, multipathway chemicals of interest (Zartarian et  
23 al., 2000). EPA recently has applied the SHEDS model to estimate arsenic exposure of children  
24 from chromated copper arsenate (CCA)-treated wood (Dang et al., 2003). EPA has also  
25 developed a Dietary Exposure Potential Model (DEPM) that links national food consumption  
26 and chemical residue data to allow estimates of average dietary intakes of metals and other food  
27 contaminants (Tomerlin et al., 1997). EPA's Total Risk Assessment Methodology (TRIM) is  
28 also being developed for multipathway risk assessment for air pollutants, including metals (See  
29 [http://www.epa.gov/ttn/fera/trim\\_gen.html](http://www.epa.gov/ttn/fera/trim_gen.html)). A generic exposure model, RESRAD, developed  
30 by the U.S. Department of Energy for risk assessment of radionuclides (U.S. DOE, 2001;  
31 LePoire et al., 2000), includes an extensive human exposure module applicable to other metal  
32 contaminants.

33  
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1 **4.2.4.2. Relative Bioavailability**

2 **4.2.4.2.1. Discussion.** Evaluating bioavailability is  
3 important, because a given dose of a metal in an  
4 environmental medium may be absorbed to a  
5 different extent than the same dose administered in  
6 the study used to derive a toxicity value (e.g., oral  
7 RfD or cancer slope factor (CSF)). In addition, it is usually assumed that the bioavailability of  
8 all metal species is the same, regardless of exposure media. Studies clearly show that the  
9 bioavailability of metals does vary by environmental medium and the species present.  
10 Therefore, bioavailability information can significantly reduce uncertainty in risk assessments  
11 and more accurately characterize potential risks to exposed populations.

Evaluating metals bioavailability can significantly reduce uncertainty in human health risk assessments and more accurately characterize potential risks to exposed populations.

12 The EPA Office of Solid Waste and Emergency Response (OSWER) has used  
13 bioavailability information in making quantitative adjustments. In 1989, OSWER published a  
14 risk assessment guidance for use in human health risk assessments at Superfund sites and has  
15 updated this guidance periodically (U.S. EPA, 1989b). This guidance recognizes that the  
16 toxicity of an ingested chemical depends on the degree to which it is absorbed from the  
17 gastrointestinal (GI) tract into the body. Thus, adjustments to bioavailability assumptions were  
18 developed to account for differences in absorption efficiencies between the medium of exposure  
19 and the medium from which the toxicity value was derived. Further, because RfDs and CSFs are  
20 generally expressed in terms of administered dose rather than absorbed dose, it also discusses the  
21 need to adjust for differences in the expression of dose between the exposure and toxicity value  
22 (e.g., absorbed vs. administered dose). The Agency guidance recommends that the relative  
23 bioavailability adjustment (RBA) of a chemical should be assumed to be equal in food, water,  
24 and soil in the absence of data to the contrary.

25 Estimating bioavailability of metals is particularly difficult because it is dependent on  
26 many variables, including the physical and chemical form of the metal, the physical and  
27 chemical characteristics of the association between the metal and soil particles, particle size of  
28 metal species, and the metal source. In addition, metal species continuously undergo reactions in  
29 soil, referred to as “aging or weathering,” that affect bioavailability.  
30

31 **4.2.4.2.2. Current Practice.** The Agency currently addresses bioavailability through the use of  
32 default values and, in some cases, through the development of site-specific and medium-specific  
33 values. To date, the most common treatment of bioavailability for human health assessments is

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1 to assume that the bioavailability of the metal exposure from the site is the same as the  
2 bioavailability of the source used to derive the toxicity value (RfD or CSF). The RfD and CSF  
3 are typically developed from laboratory toxicity tests using highly bioavailable forms and are  
4 usually based on administered rather than absorbed doses. This is true for all chemicals, but it is  
5 of special importance for ingested metals because metals can exist in a variety of chemical and  
6 physical forms, and not all forms of a given metal are equally well absorbed. For example, a  
7 metal in contaminated soil may be absorbed to a lesser extent than when ingested in drinking  
8 water or food.

9 It is important to recognize that a default RBA value of 1 (100%) is not necessarily  
10 conservative (i.e., more protective of human health). The bioavailability of the metal in the  
11 exposure medium of concern at the site may actually be greater than in the exposure medium  
12 used in the critical toxicity study that formed the basis of the RfD or CSF. If this is the case,  
13 assuming RBA of 1 for the medium of concern would result in an underestimate of risk at the  
14 site. The Agency recognizes that some cases may exist where sufficient data are available to  
15 support development of medium-specific default absorption factors for a particular chemical.  
16 The purpose of these medium-specific and chemical-specific default values is to increase the  
17 accuracy of exposure and risk calculations even when site-specific studies are not available.

18 Lead is an example of a chemical for which the Agency has established medium-specific  
19 default absorption factors for both children and adult populations. The IEUBK model for lead in  
20 children predicts PbB concentrations for a hypothetical child or population of children (birth to  
21 84 months of age) resulting from exposure to environmental sources of lead, including soil, dust,  
22 air, drinking water, and diet (U.S. EPA, 1994b; White et al., 1998). An assumption in the model  
23 is that the ABA of lead in soil and dust, at low intake rates, is 0.3 (30%) and the ABA of soluble  
24 lead in water and food is 0.5 (50%). This corresponds to an ABA of 0.6 (60%) for lead in soil  
25 (or dust) compared to lead in water or food. The model also allows for the input of site-specific  
26 values.

27 The Agency has developed the Adult Lead Methodology (ALM) for assessing lead risks  
28 in adult populations (U.S. EPA, 1996c). An assumption in the ALM is that the ABA of lead in  
29 soil is 0.12 (12%). This value is based on assumptions that the ABA of soluble lead in water is  
30 0.2 (20%) and that the RBA of lead in soil, compared to soluble lead, is 0.6 (60%).

31 The Agency has also derived RfDs that are specific for an exposure medium, based on  
32 consideration of bioavailability or other factors that might suggest unique dose-response  
33 relationships in that medium. For example, separate RfDs for cadmium in food and drinking

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1 water have been derived through the rationale that the bioavailability of cadmium in water is  
2 greater than that of cadmium in food by a factor of 2 (i.e., 5% vs. 2.5%, respectively [U.S. EPA,  
3 2003f]). Similarly, the Agency recommends that a modifying factor of 3 be applied to the  
4 chronic oral RfD for manganese when the RfD is used to assess risks from drinking water or soil  
5 to account, in part, for potential differences in bioavailability of manganese in water and soil  
6 compared to that in food (U.S. EPA, 2003g). Therefore, use of default values should not  
7 substitute for site-specific assessments of bioavailability, where such assessments are deemed  
8 feasible and valuable for improving the characterization of risk at the site.

9 As described in the NRC report (NAS, 2002), a  
10 variety of tools are available to attempt to measure  
11 bioavailability. The approaches include biomarkers of  
12 exposure (e.g., ALA activity from lead exposure), cell  
13 culture studies, isolated GI tract tissue, whole-animal  
14 approaches, and clinical studies. Of these options, the  
15 use of whole animals is most feasible (Weis and Lavelle, 1991); clinical studies offer desirable  
16 advantages but present many obstacles (Maddaloni et al., 1998). The following discussion  
17 focuses on the oral route of exposure.

While a variety of tools are available to attempt measurement of bioavailability, the use of *whole-animal approaches* are the most feasible.

18  
19 **4.2.4.2.3. Animal Models.** Historically, a variety of experimental animal models (in vivo) have  
20 been used to evaluate bioavailability, including rats, rabbits, monkeys, guinea pigs, and swine.  
21 Within an animal model, evaluation of bioavailability has included measuring the amount of  
22 metal in blood, body tissues, or excreta (e.g., feces and urine). The appropriate study design is  
23 dependent on the pharmacokinetics of the metal in the animal model and differences between the  
24 selected animal species and humans. In other words, it is important to consider how soluble  
25 forms of the metal are absorbed, how the metal is excreted, and whether there are any tissues  
26 where the metal might accumulate. The most common methods for measuring bioavailability in  
27 vivo are blood, urine, fecal and tissue measures.

28 The principal advantage of whole-animal oral chemical absorption studies is that they  
29 measure bioavailability in its most clinically relevant form, that is, from the GI tract and into the  
30 systemic circulation. This integrates all of the relevant biological components related to  
31 systemic absorption, including pre-systemic elimination, if present. By using the animals as  
32 surrogates for humans, these studies avoid the experimental and ethical problems associated with  
33 the use of human subjects. Currently, certain in vivo bioavailability studies conducted with an

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1 appropriate species are considered the gold standard for developing bioavailability information  
2 suitable for use in quantitative human health risk assessments, and they are often used to validate  
3 other bioavailability tools. For example, the young swine model for lead bioavailability has been  
4 used to validate in vitro extraction tests.

5  
6 **4.2.4.2.3.1. *Animal Model Applications.*** Animal  
7 models have served as the basis for making site-specific  
8 bioavailability adjustments at several Superfund sites.  
9 Scientists from EPA Region 8 sponsored the  
10 development of whole-body *in vivo* bioavailability  
11 studies in juvenile swine as a model of young children  
12 who were exposed to lead in soil contaminated with  
13 various forms of mine wastes (Lavelle et al., 1991; Weis  
14 et al., 1992, 1993a). The results of these efforts were  
15 subjected to outside peer review and found to be valid  
16 and acceptable for use in adjusting the RBA for lead in  
17 human health risk assessments. As a result, the juvenile swine model is presently being used as  
18 the preferred animal model for lead (U.S. EPA, 2004a). Site-specific bioavailability adjustments  
19 based on results from the juvenile swine model have been accomplished at several sites across  
20 the country, including the Murray Smelter in Colorado; Palmerton, PA; Jasper County, MO;  
21 Smuggler Mountain, CO; and the Kennecott site in Salt Lake City, UT.

The *juvenile swine model* is presently being used as the preferred animal model by EPA for making site-specific bioavailability adjustments for humans exposed to lead in soil. Site-specific bioavailability adjustments based on successful application of the model have been accomplished at several sites across the US.

22 Interim draft guidance has also been developed by EPA Region 10 for making  
23 bioavailability adjustments with arsenic-contaminated soil (U.S. EPA, 2000a).  
24 Recommendations are based on literature data on arsenic bioavailability and the results of a  
25 Region 10 animal study in which immature swine were dosed with arsenic-contaminated soil  
26 derived from the Ruston/North Tacoma Superfund site, which was a former smelter site (U.S.  
27 EPA, 1996d). This interim guidance recommends default values of RBA for arsenic in soil  
28 ranging from 60 to 100%, depending on the source of contamination (e.g., mineral processing,  
29 fossil fuel combustion, pesticides/wood treatment processes). As with lead, the juvenile swine is  
30 the recommended animal model for supporting departures from the default RBA assumptions.

31  
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1 **4.2.4.2.3.2. *Animal Model Limitations.*** Currently,  
2 bioavailability studies conducted with an appropriate  
3 animal species are considered the most reliable for  
4 making quantitative bioavailability adjustments in  
5 human health risk assessments. They are also used as  
6 a means to validate other tools, such as

Animal models are generally complex, time consuming, and more resource intensive than other tools, which may limit their feasibility to large sites where it is difficult to adequately characterize variability in bioavailability across the site.

7 physiologically based extraction tests. However,  
8 animal models are generally complex, time consuming-, and much more expensive than other  
9 tools. As a result, use of animal models is usually limited to large sites where it is difficult to  
10 adequately characterize the variability in bioavailability across the site. An investigator must  
11 ensure that the study design and animal model selected are appropriate for the metal being tested.  
12 The best measure of bioavailability (blood vs. tissue vs. feces vs. urine) is dependent on the  
13 pharmacokinetics of a particular compound. As discussed above, each measure has its own  
14 inherent limitations. Further discussion of the limitations of animal models are discussed  
15 elsewhere (NAS/NRC, 2002; NFESC, 2000a, b).

16  
17 **4.2.4.2.4. *In vitro Methods.*** Recently, significant effort has been expended on developing *in*  
18 *vitro* methods for assessing the RBA of metals, due to their ease of use and potential cost savings  
19 when compared against more traditional *in vivo* methods using laboratory animals. Several  
20 researchers have investigated *in vitro* models that attempt to simulate the conditions in the GI  
21 tract (Drexler et al., 2004; Medlin, 1997; Rodriquez et al., 1999; Ruby et al., 1993, 1996, 1999)  
22 and which are often referred to physiologically based extraction tests. These methods are based  
23 on the concept that the rate and/or extent of metals solubilization in GI fluid is likely to be an  
24 important determinant of metals bioavailability *in vivo*. These assays provide a measure of  
25 bioaccessibility or the amount solubilized in the GI fluid and available for potential absorption  
26 (Ruby et al., 1993).

27 Model development has focused on the simulation of complex physiological and  
28 biological functions within the GI tract, including considerations of pH, solids:fluid ratios,  
29 motility/transit, and solution chemistry. The most common approach has been the two-solution  
30 method, which addresses pH changes in the GI tract by providing an exposure to both the low  
31 pH of the stomach (1.3 to 3) and the higher pH of the small intestine (5.5 to 7). Solution pH is  
32 usually maintained either by titrations with drop-wise addition of acid or base while solutions are  
33 continuously monitored or by using buffers.

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1 The solids:fluid ratio used for *in vitro* models has ranged from 1:10 to approximately  
2 1:150 (g/mL). None of these ratios reflects the 2:1 ratio observed in adults (3000 g daily food  
3 intake vs. the 1500 mL stomach volume) (Washington et al., 2001). It is recommended that this  
4 ratio be dictated by practical considerations. Therefore, a sample mass that can be accurately  
5 weighed and is representative should be provided, along with a volume that can help maintain  
6 good particle-to-solution contact and minimize any unusual kinetics.

7 Motility and transit time within the GI tract are difficult to model with standardization.  
8 Both processes vary greatly and can be affected by diet and daily cycles. Historically,  
9 investigators have used either diffusers, stirrers, or rotation devices to mimic these factors. All  
10 methods are adequate; however, the diffuser system is difficult to control and clean. Also,  
11 rotation mechanisms are not favorable to techniques that require constant pH monitoring.

12 Variations observed in extraction fluid chemistry are by far the greatest source of method  
13 deviations. Although most methods have the gastric solution dominated by HCl, other acids,  
14 proteins, and peptides have been added, with extraction times of about 1 hour. Intestinal  
15 solutions have their pH adjusted by addition of sodium bicarbonate and/or other biological salts  
16 and are extracted for 3 to 5 hours. Most systems were maintained at a temperature of 37 °C, and  
17 some methods have used argon to maintain anaerobic conditions, even though the GI tract is  
18 aerobic in humans.

19  
20 **4.2.4.2.4.1. *Applications.*** Physiologically based extraction tests have been conducted for a  
21 variety of metals, including lead, arsenic, cadmium, nickel, and mercury. Model results are  
22 currently being used as a screening tool only until adequate model validation has occurred. To  
23 date, EPA has not endorsed the use of *in vitro* techniques for making site-specific bioavailability  
24 adjustments.

25  
26 **4.2.4.2.4.2. *Limitations.*** *In vitro* methods are clearly significantly cheaper and less time  
27 consuming than standard *in vivo* studies. However, these extraction techniques are based on the  
28 premise that solubility or bioaccessibility is the primary factor controlling bioavailability, which  
29 is not necessarily the case. In addition, these tests cannot reflect the complex physiological or  
30 pharmacokinetic aspects of human absorption.

31  
32 One of the key limitations of this approach is adeptly defined in NAS/NRC (2002):  
33

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1 Regulatory acceptance of the tools used to generate bioavailability information in  
2 risk assessment is expected to be influenced by several factors, including the  
3 relevance of the tools to the site conditions and the extent of tool validation.  
4 Validation variously refers to the performance of a tool or approach in terms of  
5 reproducibility, reliability, and multi-lab calibration. An appropriate body of  
6 experimental work to validate a tool would (1) clarify where and when a tool  
7 yields a definitive response; (2) clarify that the tool can be linked to a biological  
8 response of a similar magnitude, and that the linkage stands up across a range of  
9 conditions in the type of environment that is being managed; (3) test the  
10 prediction of bioavailability using different types of experiments and field studies;  
11 (4) clarify which types of biological responses are best predicted by the approach;  
12 and (5) include critiques of the best applications and the limits of the tool,  
13 especially compared to alternatives. A tool that is well accepted and validated  
14 should be given greater weight than one that is new or experimental.

15  
16 OSWER is currently developing a bioavailability document on metals to advise risk  
17 assessors and managers on whether to collect site-specific information on the bioavailability of  
18 metals in soil and how to evaluate bioavailability data for use in human health risk assessments.  
19 The document will outline a decision framework that explains how to use bioavailability data  
20 consistently as part of a human health risk assessment. The decision framework will consist of a  
21 two-tiered approach. The first tier presents general guidelines for determining whether  
22 bioavailability is worth considering at a particular site. The second tier involves an ordered  
23 process for the actual collection and analysis of bioavailability data.

#### 24 25 **4.2.4.3. Biomarkers**

26 Integration of exposures across media, route, and time of exposure can be reflected in  
27 biomarkers of exposure. The World Health Organization (WHO/IPCS, 1993) defines a  
28 biomarker of exposure as “an exogenous substance or its metabolite or the product of an  
29 interaction between a xenobiotic agent and some target molecule or cell that is measured in a  
30 compartment within an organism.” Ideally, there should also be a well-established relationship  
31 between biomarker of exposure and outcome, in that the biomarker not only provides  
32 information about exposure levels but also can be predictive of an effect. For example, urinary

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1 cadmium is directly correlated to the concentration of cadmium in the renal cortex, which is one  
2 site for toxicant action of this metal.

3  
4 **4.2.4.3.1. *Application.*** In the case of metals, urinary cadmium and blood lead are examples of  
5 exogenous substances used as biomarkers of exposure. The measurement of metals in biological  
6 fluids has been used as the primary means of quantifying biomarkers of exposure for metals by  
7 occupational health organizations such as the American Congress of Governmental Industrial  
8 Hygienists. An interaction between a metal and a target molecule, such as the adduction of  
9 chromium(VI) with DNA and protein, is used to a more limited extent. Some biomarkers of  
10 exposure such as the DNA adducts of chromium(VI) fall into the area of transition in the  
11 continuum from exposure to effect.

12 For many of the metals of interest, biomarkers of exposure and effect are used as basic  
13 tools for population or molecular epidemiology studies of effects of exposure to humans of  
14 various metals. The Centers for Disease Control and Prevention (CDC) conducts an extensive  
15 biomonitoring program of human blood and urine that includes lead, mercury, cobalt, uranium,  
16 antimony, barium, beryllium, cesium, molybdenum, platinum, thallium, and tungsten (CDC,  
17 2003). The data are summarized in age, gender, and ethnicity categories.

18  
19 **4.2.4.3.2. *Limitations.*** A biomarker of exposure is a measure of cumulative exposure to a metal  
20 and also of metal actually existent in the body, as occurs with chronic exposure for metals.  
21 However, such an approach may not be appropriate for metals that are not extensively  
22 accumulated in tissues, and it does not differentiate between metal present in a tissue in a  
23 sequestered or inactive form and metal engaged in toxic or pathological processes.

24 There are environmental (water, air, soil, dust), occupational, medicinal, and dietary  
25 sources of metal exposure. For this reason, use of biomarkers increases the need for  
26 comprehensive, multi-pathway assessments of exposure. Reference or background levels of  
27 biomarkers of exposure are essential for any assessment. Failure to consider background dietary  
28 sources of metals may result in a misinterpretation of the exposure. For example, arsenobetaine  
29 is a nontoxic organic form of arsenic found naturally in shrimp and other seafood. The analysis  
30 of total unspiciated urinary arsenic of individuals who consume seafood, without recognition of  
31 their diet history, will lead to an overestimation of exposure to potentially toxic (inorganic)  
32 arsenic species—some assessments of arsenic exposure have assumed that 10% of total

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1 elemental arsenic in seafood and 100% of arsenic in all other foods is in a toxic, inorganic form  
2 (NAS/NRC, 1999).

3 The correct frequency and timing of sampling of biological fluids and tissues, as well as  
4 the correct interpretation of the results, depend on knowing the elimination half-life of the metal.  
5 The half-life of lead in plasma, blood, soft tissues, and bone ranges from hours to months to  
6 years (Sakai, 2000). A detection of lead in plasma above background levels would be indicative  
7 of an acute exposure, whereas a detection in bone would be indicative of chronic exposure.  
8 Thus, sampling plasma every other day or week, or analyzing bone, would not be the best way to  
9 determine whether an acute exposure to lead occurred.

10 The validity of a biomarker is supported by three kinds of relevance: analytical,  
11 toxicokinetic, and biological (Schulte and Talaska, 1995; Grandjean et al., 1994; WHO/IPCS,  
12 1993). Key analytical issues include specificity, sensitivity, standardization of methodologies (to  
13 reduce intra- and interlaboratory variability), speciation, quality assurance, and the availability of  
14 reference samples. Analytical methods for the detection of metals include ICP-MS, hydride  
15 generation atomic absorption, and fluorescence spectrometry. When coupled with  
16 high-performance liquid chromatography, these methods are enhanced because of the ability to  
17 detect speciated parent metal and metabolites. Although these methods can be very reliable for  
18 the analysis of metals in biological fluids, using them for tissue analysis is more difficult.  
19 Digestion and extraction make it difficult to fully speciate the metal and produce interfering  
20 matrix factors. Reference standards for tissues are seldom available. X-ray fluorescence  
21 spectrometry, used to detect lead in bone (Ambrose et al., 2000), and neutron activation analysis,  
22 used for manganese in liver (Arnold et al., 1999), are highly powerful noninvasive in vivo  
23 techniques; however, the sensitivity is extremely limited. A disadvantage of any in vivo method  
24 is that the metal species in the environmental exposure cannot be estimated correctly.

#### 26 **4.2.5. Toxicokinetics and Toxicodynamics**

27 Several specific properties of metals that affect their absorption, distribution, metabolism,  
28 and elimination can be identified. These properties of metals can influence their physiological  
29 handling as compared to organics (Table 4-10) and should be taken into account in  
30 pharmacokinetic analyses related to human health risk assessment. Properties that affect  
31 toxicodynamics should also be taken into account in these analyses.

32  
33  
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**Table 4-10. Metal (versus organic) compound properties affecting absorption, distribution, metabolism, and elimination**

Organic compounds	Metals
Metabolism is generally extensive and often species-specific.	Metabolism is usually limited to oxidation state transitions and alkylation/dealkylation reactions.
Persistence in body fat is common because of lipid solubility (not capacity-limited).	Often sequestered, bound to specific plasma or tissue proteins (intrinsically capacity-limited) or bone.
Predominantly eliminated by excretion in urine and exhaled air after biotransformation from lipophilic forms to hydrophilic forms.	Predominantly eliminated in urine and bile. Metal compounds are hydrophilic.
Tissue uptake is most commonly a blood flow-limited process, with linear partitioning into tissues.	Metals and their complexes are often ionized, with tissue uptake (membrane transport) having greater potential to be diffusion-limited or to use specialized transport processes.
Interactions with other structurally similar compounds may occur, especially during metabolism.	Interactions among metals and between metals and organics are numerous and occur commonly during the processes of absorption, excretion, and sequestration.

The evaluation of toxicodynamics addresses the sequence of biochemical events at the cellular and molecular levels that begin when toxicologically active form of the metal interacts with the target (e.g., from molecule to a protein, enzyme, or other cellular molecule, that leads to a toxic physiological response. Toxicodynamics involves the biological processes that underlie the severity of an effect as well as its reversibility, recovery, and adaptive response. These evaluations are applied for single metals as well as for metal mixtures, recognizing that one metal can induce multiple effects (from critical to secondary and more, as the exposure levels increase) and common effects can be exerted across many metals. To assure that effects are appropriately combined, it is important to reflect the underlying mechanism or mode of action; the potential influence of toxicokinetic processes should be considered (e.g., changes in gastrointestinal absorption and liver and kidney retention can change liver and red blood cell functions).

Two properties of metals, their hydrophilic nature and their characteristic protein binding, are discussed in more detail below, followed by a discussion of available physiologically based pharmacokinetic (PBPK) models.

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#### 1 **4.2.5.1. *Hydrophilic Properties of Metals***

2 Solubility in aqueous media is one of the major factors influencing absorption of metals  
3 and metal compounds. The water solubility of a metal compound depends on its chemical  
4 species, on the pH of its medium (H<sup>+</sup> ions), and on the presence of other chemical species in the  
5 medium (see Section 4.1). Nitrates, acetates, and all chlorides of most metals except silver,  
6 mercury, and lead are soluble. Sulfates of most metals are also soluble, except for barium and  
7 lead. On the other hand, most hydroxides, carbonates, oxalates, phosphates, and sulfides are  
8 poorly soluble. Another factor influencing absorption of poorly soluble compounds is particle  
9 size: fine particles are usually more soluble. Metallic lead in body tissues (as may occur  
10 following gunshot wounds) is probably absorbed after being oxidized to soluble salt. Metallic  
11 mercury is corrosive when embedded in body tissues, but metallic mercury swallowed into the  
12 gastrointestinal tract is not soluble (Goyer and Clarkson, 2001).

13 The relative hydrophilic nature of metals versus organic compounds influences  
14 absorption at different sites. Absorption of metals in the gastrointestinal tract is hindered by the  
15 lipid nature of intestinal cell membranes but is favored by solubility in the hydrophilic contents  
16 of the gastrointestinal tract (preabsorption). In the lungs, the absorption of aerosols of  
17 particulate forms of metals and metal compounds and of lipophilic organic compounds may not  
18 be as dependent on the lipophilic or hydrophilic nature of the substance, depending more on  
19 particle size and on whether the substance is presented as a vapor or a gas (e.g., elemental  
20 mercury). Human skin is not very permeable and provides a good barrier against absorption of  
21 metals and metal compounds as well as highly lipophilic organic compounds. Elemental  
22 mercury and dimethyl mercury (Siegler et al., 1999) are notable exceptions. When dermal  
23 absorption does occur, the mechanism may differ between organic substances and metals. Polar  
24 substances, like metal compounds, appear to diffuse through the outer surface of protein  
25 filaments of the stratum corneum, which is hydrated, whereas lipophilic nonpolar organic  
26 molecules diffuse through the lipid matrix between the protein filaments (Rozman and Klaassen,  
27 2001).

28 Hydrophilic metal ions do not readily diffuse into richly lipophilic tissues, such as the  
29 brain, liver, and neutral fat stores, where they would be difficult excrete. Although  
30 biotransformation of metals occurs, the products typically maintain a hydrophilic profile.  
31 Entrance of metals or inorganic metal compounds into lipid-rich tissues like the brain depends on  
32 hydrophilic pathways. Retention in tissues of metals or metal compounds is generally related to  
33 formation of inorganic complexes or metal protein complexes (e.g., lead in bone and cadmium in  
34 tissues bound to the low-molecular-weight protein metallothionein).

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1           Although the low lipid solubility of metal ions could limit their accessibility to tissues  
2 and cells, recent rapid progress in identifying metal transporters (Foulkes, 2000) suggests that  
3 generalizations are not appropriate, and each metal should be assessed in terms of its ability to  
4 access transporters and of the presence of transporters in potential target organs. Furthermore,  
5 complex lipids can offer high-affinity binding sites for metal ions that promote their distribution  
6 to lipid compartments, and some metals, such as thallium, have a demonstrated affinity for  
7 adipose compartments.

8           Generally, the primary factor that influences the uptake and distribution of organic  
9 substances within the body is the substances' lipophilicity. Passage through cellular membranes  
10 and partitioning to organs occurs primarily through passive diffusion through the lipid portions  
11 of cellular membranes. Metabolism (or lack thereof) can, of course, influence the distribution  
12 and excretion of organic chemicals primarily because of the accompanying changes in chemical  
13 structure and, therewith, lipophilicity. The uptake, distribution, metabolism and excretion of an  
14 organic chemical can often be predicted by consideration of the substance's chemical structure  
15 and lipophilicity (i.e., octanol/water partition coefficient).

16           In contrast to organic substances, the uptake and distribution of metals in inorganic forms  
17 is generally influenced by atomic size and charge, and the availability of active cellular  
18 processes that naturally exist for the uptake and distribution of nutritional metals (e.g., calcium,  
19 sodium, potassium, magnesium) that can also transport other metals into or across cell  
20 membranes. The assumption of linear partitioning in tissues commonly applied to organics, and  
21 which is largely lipophilicity-based, is therefore not appropriate for the capacity limited  
22 processes that generally control the uptake and disposition of metals. Rather, the uptake,  
23 distribution, metabolism, sequestration, and mechanisms of action of metals is generally  
24 considered in the context of their kinetic behavior.

25           Metals can have residence times in the body as long as months or years, particularly  
26 when they are bone-seeking elements or are associated with tissue storage proteins. Metals, such  
27 as lead, strontium, and uranium, may be incorporated into bone, stored there, and many years  
28 later may be released from the bone tissue into the systemic circulation to sites of toxic action,  
29 by mechanisms closely linked to bone metabolism. These mechanisms include incorporation  
30 and loss with bone formation and resorption and, depending on the ionic radius and charge of the  
31 metal, can include migration within the bone complex as well as out of the bone (O'Flaherty,  
32 1998). While long-term deposition of a metal (e.g., lead) in a tissue (e.g., bone) may not  
33 necessarily result in toxicity to that tissue, subsequent release of that metal from that tissue  
34 enables transportation of the metal to its site of toxic action elsewhere in the body. It is

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1 important for the risk assessor to consider the potential toxicological consequences of storage or  
2 deposition of metals in body compartments as part of the overall human health assessment.

3 Modeling the kinetics of metals with extended tissue residence times presents challenges  
4 in that anatomical and physiologic functions, along with changes in exposure, need to be  
5 characterized as a function of age and clinical status. For example, bone turnover is rapid in  
6 children and relatively quiescent in adults, but is increased in adults during conditions such as  
7 menopause, pregnancy, and lactation, to name a few. Ninety to ninety five percent of the adult  
8 human body burden of lead is contained in the bone. However, lead is not permanently fixed in  
9 bone tissue, but returns to the blood as bone is resorbed and by return from the bone surface  
10 (O'Flaherty, 1998). Gulson et al (1995) observed that between 45 and 75 percent of blood lead  
11 in a group of adult women had originated from the bone. Physiologically based kinetic models  
12 have been developed or are in the process of development for arsenic, lead, chromium, and  
13 mercury (O'Flaherty, 1998).

#### 14 15 **4.2.5.2. Metal-Binding Proteins**

16 Metals react with many different proteins in the body that may modify kinetics. Many  
17 metals bind with albumin for purposes of transport in the circulatory system and across cell  
18 membranes and within cells. However, research is identifying a growing number of proteins that  
19 play specific roles in transport, cellular uptake, and intracellular storage of metals (Goyer and  
20 Clarkson, 2001), including the following:

- 21  
22 • **Transferrin.** Transferrin is a glycoprotein that binds most of the ferric ion in plasma  
23 and has a role in transporting iron across cell membranes. This protein also  
24 transports aluminum and manganese.
- 25  
26 • **Ceruloplasmin.** Ceruloplasmin is a copper-containing glycoprotein oxidase in  
27 plasma that converts ferrous to ferric iron, which then binds to transferrin.
- 28  
29 • **Membrane carrier proteins.** A number of recently discovered carrier proteins  
30 transport metals across cell membranes. Many of these carrier proteins are  
31 multispecific (e.g., divalent metal transporter 1 and 2, metal transporter protein 1),  
32 Some metals are transported as complexes with endogenous ligands; no transport  
33 systems are intended for the ligand itself, accepting substrates that vary considerably  
34 but are recognized by the attached metal ion (Dawson and Ballatori, 1995).
- 35  
36 • **Metallothioneins.** The metallothioneins are a group of low-molecular weight (MW)  
37 proteins (MW about 6,000 daltons), rich in sulfhydryl groups that serve as ligands for

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1 several essential and nonessential metals. In vitro studies have found that the highest  
2 affinity is for silver, then in descending order mercury, copper, bismuth, cadmium,  
3 lead, and zinc (Kagi and Kogima, 1987). However, studies of in vivo  
4 metallothioneins from various sources included zinc, copper, and cadmium.  
5 Metallothioneins have multiple binding sites that have different affinities for metals.  
6 Also, the types of metals bound to metallothioneins differ depending on the species,  
7 the organ, and previous exposures to metals, but most of them contain at least two  
8 different types of metals. For example, metallothioneins isolated from adult or fetal  
9 human livers contain mainly zinc and copper, and those from human kidneys contain  
10 cadmium, copper, and zinc (Cherian and Goyer, 1995). In most cases, the  
11 metallothioneins are inducible and perform a number of functions, including serving  
12 as a storage protein for zinc and copper in the liver, kidney, brain, and possibly skin  
13 and having an important protective role in cadmium toxicity (Goyer and Clarkson,  
14 2001). Although metallothioneins have an affinity for lead in vitro, in vivo binding to  
15 lead has not been demonstrated. Also, mercury may induce synthesis of  
16 metallothionein in vivo, but binding is only temporary regardless of the demonstrated  
17 in vitro affinity.

- 18
- 19 • **Ferritin.** Ferritin is primarily a storage protein for iron in reticuloendothelial cells of  
20 the liver, spleen, and bone. It plays an important role in turnover of iron. It has also  
21 been suggested that ferritin may serve as a general metal agonist because it binds a  
22 number of metals, including cadmium, zinc, beryllium, and aluminum.
- 23
- 24 • **Lead-binding protein(s).** Lead binds with a number of lead-binding proteins, but  
25 their identity or function is not as well defined as that of other metal-specific proteins.  
26 The most studied lead-binding protein is the denatured lead-protein complex  
27 identified as the intracellular inclusion body occurring in cells, particularly in the  
28 liver and kidney in persons with high-level lead exposure. It has been suggested that  
29 lead-binding proteins may have a protective effect for lead (Goyer and Clarkson,  
30 2001).
- 31

#### 32 **4.2.6. Pharmacokinetic/Pharmacodynamic Modeling of Metals**

33 Physiologically based pharmacokinetic/physiologically based pharmacodynamic  
34 (PBPK/PBPD) modeling of metals entails the mathematical description and modeling of a  
35 substance's behavior in the body (e.g., absorption, distribution, metabolism, excretion, and toxic  
36 effects). PBPK and PBPD models are valuable risk assessment tools for interspecies, high-  
37 dose/low-dose, route-to-route, and exposure scenario extrapolation (Krishnan and Andersen,  
38 1994). PBPK models consist of multiple compartments representing tissues or tissue groups that  
39 are linked by blood flow. PBPD models describe the relationship between target tissue dose and  
40 health endpoints or target tissue effects. PBPK models that include a fetal compartment are

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1 particularly valuable for human metal risk assessment. In utero exposure can be important for  
2 establishing the body burden of certain metals before birth that result from transplacental  
3 transfer.

4 PBPK models have historically been developed and used for risk assessment of volatile  
5 organic compounds (VOCs) (e.g., methylene chloride) (Andersen et al., 1987) but have also been  
6 applied to many metals (White et al., 1998; Clarke, 1995). Metals differ in their kinetic behavior  
7 from VOCs in a number of ways, as discussed by O'Flaherty (1998). Guidelines exist for model  
8 evaluation, and these should be used as a framework to determine whether a particular model is  
9 appropriate for use in risk assessment (Clark et al., 2003; Andersen et al., 1995).

#### 11 **4.2.6.1. Application**

12 Combined use of PBPK and PBPD models provides understanding of the complex  
13 relationships between exposure and target organ effects. PBPK models are often capable of  
14 predicting aggregate exposures for comparison to exposure models.

15 For metals that have long retention times in tissues, the maternal tissues can serve as a  
16 reservoir for exposures during fetal development. This can be a particularly important exposure  
17 pathway for metals that accumulate in the inorganic matrix of bone (e.g., lead, strontium,  
18 uranium) because mobilization of bone minerals to develop the fetal skeleton can result in a  
19 transfer of maternal bone stores of metals to the fetus (e.g., Gulson et al., 1999a, b; Tolstykh et  
20 al., 1998). Transplacental exposures cannot be directly measured from environmental  
21 measurements but require the use of PBPK models. A few models of transplacental transfer of  
22 lead in humans have been developed; models for other metals are not available for use in risk  
23 assessment. The lead models reported to date are limited in that they rely on assumptions of a  
24 steady state between maternal and fetal blood lead concentrations (U.S. EPA, 1996c, 1994b;  
25 Leggett, 1993; O'Flaherty, 1993). This assumption will be violated if the mother is no longer  
26 exposed to lead and if the fetal exposure is due to remobilization of lead from the mother's  
27 bones. Furthermore, if the mother experiences lead exposures for the first time during  
28 pregnancy, the lead will be partitioned among the various body compartments (bone, hair, blood,  
29 fetus) in a dynamic manner.

30 When using PBPK models or other dosimetric adjustments in the risk assessment process  
31 for metals, one should explicitly consider the absorption/distribution and kinetic factors  
32 discussed above. There are special considerations for cellular uptake, interaction with  
33 nutritionally essential and nonessential metal, protein-binding behavior and function,  
34 incorporation into bone, metabolism, and excretion as outlined in Table 4-11.

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**Table 4-11. Kinetic factors to consider when evaluating the use of PBPK models or other dosimetric adjustments in the risk assessment process for metals in humans**

Cellular uptake	<ul style="list-style-type: none"> <li>▶ Carrier-mediated uptake (e.g., phosphate or sulfate transporters)</li> <li>▶ Facilitated transport in the form of organic complexes</li> </ul>
Interaction with nutritionally essential and nonessential metal	<ul style="list-style-type: none"> <li>▶ Competition for binding sites on membrane transport proteins</li> <li>▶ Interactions at enzyme active sites?</li> <li>▶ Systemic level interactions altering absorption</li> </ul>
Protein-binding behavior and function	<ul style="list-style-type: none"> <li>▶ Capacity limited to binding to specific proteins</li> <li>▶ Inducibility of binding proteins</li> <li>▶ (Zn, Cu, Cd, As, Ni, Hg to metallothionein)</li> <li>▶ Protein binding as sequestration mechanism</li> <li>▶ Pb-binding protein in inclusion bodies</li> </ul>
Incorporation into bone	<ul style="list-style-type: none"> <li>▶ Lead sequestered in bone</li> </ul>
Metabolism	<ul style="list-style-type: none"> <li>▶ Relative contribution to overall elimination compared to excretory mechanisms</li> </ul>
Excretion	<ul style="list-style-type: none"> <li>▶ Relative contribution of urinary and biliary excretion</li> <li>▶ Capacity limitation (saturation kinetics)</li> </ul>

1            Pharmacokinetic models for use in human metal risk assessment include three models  
2 for lead. The O’Flaherty Model is a PBPK model for children and adults. It includes the  
3 movement of lead from exposure media (i.e., intake via ingestion or inhalation) to the lungs and  
4 gastrointestinal tract; subsequent exchanges between blood plasma, liver, kidney, and richly and  
5 poorly perfused tissues; and excretion from liver and/or kidney (O’Flaherty, 1995). The IEUBK  
6 model was developed by EPA for predicting lead levels in children (U.S. EPA, 1994b). The  
7 Leggett model allows simulation of lifetime exposures and can be used to predict blood lead  
8 concentrations in both children and adults (Leggett, 1993). EPA has a research program for the  
9 development of an all ages lead (biokinetic) model and a cadmium biokinetic model based, at  
10 least initially, on the Kjellstrom and Nordberg model (Kjellstrom and Nordberg, 1978).

11  
12 **4.2.6.2. Limitations**

13            Many of the processes controlling the disposition of metals are intrinsically capacity-  
14 limited and highly metal-specific. This makes it necessary to understand physiology well  
15 enough to model these processes and methods to estimate binding constants. Another  
16 overarching theme is that metal-metal interactions of multiple types (e.g., competition,

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1 antagonism, and synergism as well as essential-nonessential metal interactions) commonly occur  
2 at multiple points during the processes of absorption, distribution, metabolism, and excretion.  
3 Another distinctive characteristic of metals is that common sequestration mechanisms, such as  
4 incorporation into bone and binding to storage proteins, can result in extended residence times.

5 In addition to kinetic factors, constructive use of PBPK and PBPD models in the risk  
6 assessment process also requires some consensus concerning mode(s) of action and the form of  
7 the chemical responsible for the effect of greatest toxicological concern to select an appropriate  
8 dose metric. The issue of which endpoints are matched with what form or species of the metal  
9 will influence the functional form of the model and hence the dose metric selection. A critical  
10 consideration will be to match the toxic endpoint with the active form of the metal in cases  
11 where sufficient data exist to suggest that there are one or more active forms of the metal or  
12 metalloid (e.g., arsenic).

13

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1 **4.3. HUMAN HEALTH EFFECTS**

2 **4.3.1. Introduction**

3 Issues of importance when conducting the Effects Assessment (dose-response  
4 determination) for human health risk assessments for metals and metal compounds are discussed  
5 in this section. The discussion provides some of the scientific basis that underlies metal-specific  
6 characteristics of human health effects assessment, but it is not intended to be a comprehensive  
7 review. Further detailed discussions on the points raised here can be found in Goyer et al.  
8 (2004). Suggested approaches for including metal-specific information in risk assessments are  
9 intended to complement other general Agency guidance (e.g., Carcinogen Risk Assessment,  
10 Exposure Assessment (U.S. EPA, 2003), Developmental Toxicity (U.S. EPA, 1991).

11  
12 **4.3.2. Essentiality Versus Toxicity**

13 Seven elements are designated as nutritionally essential for humans by the National  
14 Academy of Sciences (NAS) (Table 4-12, first column). The categorization as an “essential  
15 nutrient” includes the identification of Recommended Dietary Allowances (RDAs) or Adequate  
16 Intakes (AIs) by the Food and Nutrition Board (FNB) of the NAS. The FNB has also examined  
17 the possible beneficial effects of other elements (Table 4-12, second column), which have not yet  
18 been categorized as essential (or not) (NAS/IOM, 2002). The extent to which these elements can  
19 currently be considered beneficial to humans and animals varies and is discussed in Goyer et al.  
20 (2004) and in NAS/IOM (2001, 2002).

21  
**Table 4-12. Metals classified by their known essentiality**

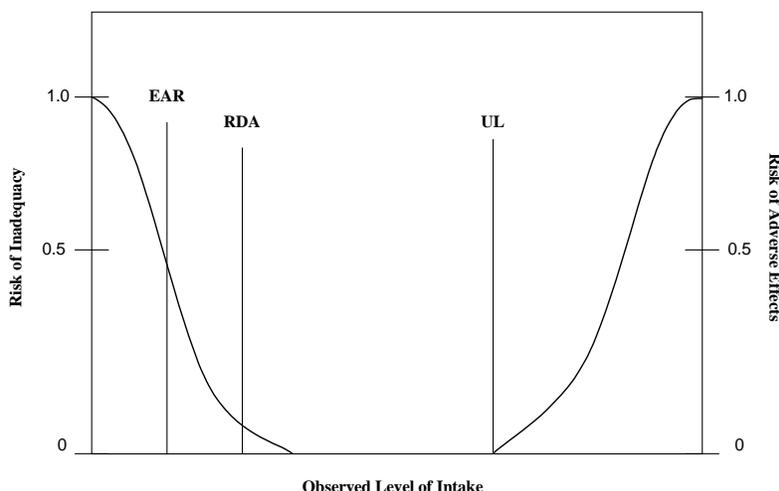
Nutritionally essential metals	Metals with possible beneficial effects <sup>a</sup>	Metals with no known beneficial effects
Cobalt Chromium(III) Copper Iron Manganese (animals but not humans) Molybdenum Selenium Zinc	Arsenic Boron Nickel Silicon Vanadium	Aluminum Antimony Barium Beryllium Cadmium Lead Mercury Silver Strontium Thallium Tin

<sup>a</sup> Possible beneficial effects in some physiological processes and some species as reviewed by the Food and Nutrition Board (FNB) (NAS/IOM, 2000). Beneficial effects have been posited for other metals but have not been officially reviewed by the FNB.

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Table 4-12 (third column) also lists some metals of interest to the EPA; the FNB has not yet reviewed their status as essential/beneficial nutrients. This category includes the metals lead, mercury, and cadmium. Dose-response curves for essential elements are shown in Figure 4-4.



**Figure 4-4. Dose-response curves for essential elements.** The response of organisms to metals that have both toxic and nutritional properties is conceptualized as having three phases: an area where there is a risk of inadequacy, a neutral area, and an area where there is a risk of adverse effects. Source: FNB (2000)

Note: This figure simply illustrates basic concepts and does not reflect an exact scale. (That is, although it might appear that the RDA is set at a level associated with a risk of inadequacy above 0.02-0.03, that is not the intent.)

### Dietary Reference Intakes

**Recommended Dietary Allowance (RDA)**—The average daily dietary nutrient intake level sufficient to meet the nutrient requirement of nearly all (97 to 98 percent) healthy individuals in a particular life stage and gender group.

**Adequate Intake (AI)**—The recommended average daily intake level based on observed or experimentally determined approximations or estimates of nutrient intake by a group (or groups) of apparently healthy people that are assumed to be adequate; used when an RDA cannot be determined.

**Tolerable Upper Intake Level (UL)**—The highest average daily nutrient intake level that is likely to pose no risk of adverse health effects to almost all individuals in the general population. As intake increases above the UL, the potential risk of adverse effects may increase.

**Estimated Average Requirement (EAR)**—The average daily nutrient intake level estimated to meet the requirement of half the healthy individuals in a particular life stage and gender group.

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1           Considerable concern for human health has been focused on the toxicological aspects of  
2 essential metals. Because of this concern, the FNB of NAS has provided Tolerable Upper Level  
3 Intake Values and RDAs for these metals where data permit and has provided guidance for  
4 assessing risk from dietary exposures to these elements (NAS/IOM, 2001). The World Health  
5 Organization (WHO/IPCS, 2002) also has provided guidance on methods for assessing risks  
6 from excessive exposure to nutritionally essential metals.

#### 7 8 **4.3.3. RDAs and RfDs/RfCs**

9           The RDA and Reference Dose (RfD)/Reference Concentration (RfC) differ significantly.  
10 The RDA is indexed to a clinical effect of deficiency; and it is used to guide people to take in  
11 enough of that material to maintain their health. Conversely, RfDs/RfCs are used to assess the  
12 potential for toxic effects from elevated exposures (note that RfDs/RfCs are based on many  
13 endpoints, which have differing levels of relevance to whole organism effects). In any case, the  
14 RDA is designed to minimize adverse effects associated with nutritional deficiency, so is  
15 applicable only to dietary exposures (notably food and beverages). Note that these values are not  
16 necessarily synonymous with optimal health (for example, it has been suggested that zinc intakes  
17 above the RDA might have certain beneficial/protective effects, including for the prostate in  
18 men, for limiting menopausal calcium loss in women, and assisting copper metabolism). RDAs  
19 have been identified for different age groups and genders, with RDAs listed for 16 different  
20 age-sex and 6 age-pregnancy combinations (NAS/IOM 2003). These RDAs are based on  
21 distributions from empirical data or general assumptions that cover 97 to 98% of the population  
22 in the given age/gender category, and for women, additional pregnancy/lactational categories.  
23 The RDA is given as mass (milligram, or mg) per day.

24           In contrast, the RfC is used to assess the potential for a noncancer effect. It represents  
25 the amount of a given metal or other material that humans can take in every day without  
26 appreciable risk of any harmful effect during a lifetime, including for sensitive subpopulations.  
27 The RfD is chemical- and route-specific (oral). The RfC represents a similar measure, but for  
28 the air concentration to which someone can be exposed (via inhalation). Sensitive subgroups are  
29 commonly addressed through the use of uncertainty factors (which also account for variability)  
30 incorporated into these values. Dietary exposures to commercial foods (not locally grown) are  
31 typically categorized as “background” and are not often included in environmental risk  
32 assessments. The RfD is given as a mass (mg) per kilogram (kg) body weight that can be  
33 safely ingested orally every day, and the RfC is given per m<sup>3</sup> of air (i.e., amount safely inhaled).

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1 Both RDAs and RfDs/RfCs have been developed for several metals; for example, values  
2 exist for chromium, copper, manganese, molybdenum, selenium, and zinc), and ULs are  
3 available for boron and vanadium, and RfDs (but not RDAs) are available for certain compounds  
4 of those metals. This highlights the basic concept underlying all of toxicology that *the dose*  
5 *makes the poison* (from Renaissance physician Paracelsus). That is, dose is the key factor in  
6 determining whether the effect of a given metal will be beneficial or adverse, considering other  
7 exposure conditions.

#### 8 9 **4.3.4. Biology Relevant to Toxic and Essential Properties of Metals**

10 The bioaccessibility and bioavailability of metals in locally grown food will depend on  
11 the species of the metal in the environment and after ingestion; this exposure may be affected by  
12 other chemicals such as metal-chelating agents that restrict uptake (e.g., phytates in plant foods)  
13 or facilitate uptake (e.g., ascorbic acid). Metals in food can be found in organic forms or  
14 complexes not found in soil, sediment, water, or air. For these reasons, the bioavailability of a  
15 metal within food may differ considerably from its bioaccessibility in an environmental media;  
16 to illustrate, different oral RfDs have been developed for cadmium in food and cadmium in  
17 water.

18 The term “molecular or ionic mimicry” has been applied to situations in which a metal  
19 forms a complex with an endogenous ligand, and the resulting compound mimics the behavior of  
20 a normal substrate, disrupting normal function. Such interactions play an important role in the  
21 health assessment for specific metals. One well-studied example is the replacement of zinc by  
22 lead in heme-synthesis, which inhibits the function of heme-synthesizing enzymes (Goyer and  
23 Clarkson, 2001).

24 Single nutrient deficiencies, such as iron deficiency anemia or iodine deficiency goiter,  
25 are well characterized. Generally, intake of essential elements is sufficient in affluent  
26 populations to avoid such diseases. However, mild multiple nutrient deficiencies may occur that  
27 may be indistinguishable from the pathophysiological effects of primary deficiency of a single  
28 metal. In conducting environmental exposure analyses, it is important to consider whether any  
29 nutritional deficiencies exist in the population being assessed, or if intake of all trace elements  
30 needed to maintain health is met, so results do not confuse adverse health effects due to  
31 nutritional deficiency with those resulting from toxic responses to excessive levels.

32 The uptake of essential metals from dietary food and water and their subsequent  
33 distribution within the body are regulated by homeostatic mechanisms that allow enhanced  
34 absorption or excretion at low dietary intakes and diminished absorption/excretion with high

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1 dietary intakes. Furthermore, dietary factors can reduce the uptake of essential metals. For  
2 example, high amounts of phytates in the diet complex with endogenous and exogenous zinc,  
3 thus preventing reabsorption and increasing zinc elimination. Specialized carrier proteins that  
4 may originally have developed as a transfer mechanism for the essential elements also are used  
5 to sequester or eliminate nonessential metals (see above, Section 4.3.3.3).

6 However, homeostatic control mechanisms can be overwhelmed (e.g., by ~~the~~ a high dose  
7 level) or otherwise circumvented to produce a toxic effect . (Note that actions of metals on skin  
8 or pulmonary membranes are not driven by homeostatic controls.) For example, at high doses  
9 chromium salts can produce severe effects in the skin or respiratory tract mucosa, and inhalation  
10 of zinc oxide fumes can give rise to inflammatory response (per cytokine excretion) that can  
11 produce a local tissue effect (unrelated to systemic zinc exposure levels)..

12 Toxic effects of metals also result from interactions that block the availability or activity  
13 of essential metals. For example, lead can block the utilization of iron in heme synthesis by  
14 inhibiting the enzyme ferrochelatase, and there is evidence that cadmium can block the entry of  
15 zinc into the fetus, thereby causing a variety of developmental defects in the newborn.  
16 Cadmium, lead, and mercury, in combinations or by themselves, may reduce the availability of  
17 zinc, copper, and selenium when these essential elements are present in marginal amounts in the  
18 diet. In addition, a competitive interaction between one or more essential metals could lead to  
19 toxic effects, e.g., copper toxicity may be enhanced by reduced levels of molybdenum or vice  
20 versa.

#### 21 22 **4.3.4.1. Limitations**

23 The RDAs have been developed for essential metals (as well as other materials, including  
24 vitamins) and are designed to provide adequate nutritional intake for 97 to 98% of the  
25 population. Should there be a narrow window between the required and toxic amounts of an  
26 element (e.g., as for selenium), then it is possible that sufficient amounts for those who need the  
27 most may be above toxic levels for the majority of the population. This potentiality should be  
28 evaluated on a case-by-case basis using what is known about potential sensitivities to that  
29 element. As a note, RDAs have been established for single chemicals, not mixtures, while metal  
30 RfDs to date are also for single chemicals. Under this situation, the component-based approach  
31 to assess mixtures is commonly applied (see section 4.3.6).

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### 1 4.3.5. Toxicity

2 Metals are associated with numerous health effects that are reviewed in detail in reports  
3 from EPA (IRIS reports), the Agency for Toxic Substances and Disease Registry (ATSDR)  
4 Toxicological Profiles, reports from the World Health Organization's International Programme  
5 for Chemical Safety, and toxicology textbooks. At least five transition metals—arsenic,  
6 cadmium, chromium(VI), beryllium, and nickel—are accepted as human carcinogens in one  
7 form or another or in particular routes of exposure (NTP, 2002), and inorganic lead compounds  
8 are considered probable human carcinogens by EPA's IRIS program, while IARC (2004) has  
9 concluded there is limited evidence of carcinogenicity to humans (See:  
10 <http://www.epa.gov/iris/subst/0277.htm#carc> and  
11 <http://monographs.iarc.fr/htdocs/announcements/vol87.htm>). Other effects of metals are also  
12 well documented, including effects on the neurological, cardiovascular, hematological,  
13 gastrointestinal, musculoskeletal, immunological, and epidermal systems. For example,  
14 following oral exposure, beryllium can cause intestinal lesions and copper can cause intestinal  
15 irritation, while nickel can decrease kidney weight and cadmium can cause proteinuria, and both  
16 trivalent chromium and nickel can decrease liver and spleen weights. Many metals, including  
17 those that are toxic (including carcinogenic), follow the metabolic pathways of similar essential  
18 metals, the result of similar binding preferences among various metals (Clarkson, 1986).  
19 Carcinogenic metals typically do not require bioactivation, at least not in the sense that an  
20 organic molecule undergoes enzymatic modification that produces a reactive chemical species  
21 (Waalkes, 1995). Enzymatic modification is generally not a mechanism available for detoxify  
22 metals. The body may use other detoxification mechanisms, such as long-term storage (e.g.,  
23 cadmium) and biliary and/or urinary excretion.

#### 24 25 4.3.5.1. Application

26 Many factors related to pharmacokinetics and susceptibility act as determinants of  
27 toxicity following exposure to a metal. Short-term exposures may produce target organ effects  
28 very different from those produced by a similar dose over a longer period of time. Short-term,  
29 high-level exposure by ingestion may give rise to well-recognized acute toxicity syndromes,  
30 usually involving the gastrointestinal tract initially and possibly secondarily involving the renal,  
31 cardiovascular, nervous, or hematopoietic systems. Survivors of acute high-dose arsenic  
32 ingestion usually experience multiple organ effects, sometimes with long-term sequelae. Long-  
33 term, low-dose exposures from ingestion of metals in food and water generally cause an  
34 accumulation in target organs over time. Such exposures can involve any organ system but do

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1 not usually produce overt gastrointestinal symptoms. For example, low-level, long-term  
2 exposure to cadmium in food—sometimes combined with inhalation exposure from cigarette  
3 smoking—will cause cadmium to accumulate in target organs (e.g., kidney) but will not produce  
4 any obvious clinical effects until “excess” capacity is diminished to a point where the normal  
5 function is lost (e.g., onset of renal disease and/or osteoporosis later in life).

6 Nickel and nickel compounds and chromium and chromium compounds are well-  
7 established contact allergens. Other metals that have been cited as contact allergens include  
8 copper (WHO/IPCS, 1998), cobalt salts (AIHA, 2003), organomercurials (AIHA, 2003),  
9 beryllium (WHO/IPCS, 1990b), palladium (Kimber and Basketter, 1996), and gold (Kimber and  
10 Basketter, 1996). Toxic interactions with the immune system that result in exaggerated  
11 responses are known as hypersensitivity or allergic reactions. Allergic contact dermatitis (or  
12 delayed hypersensitivity) is one such example and occurs as a result of allergy to a substance  
13 (antigen) through cell-mediated immunity. In sensitized persons, such reactions can be provoked  
14 by minute amounts of the allergen. There are two main phases in cell-mediated immunity, the  
15 sensitization phase (in which the person becomes allergic to the antigen) and the elicitation  
16 phase. Sensitization usually takes at least 10 days. When sensitization has been achieved and  
17 the individual is then re-exposed, a reaction is obvious after a characteristic delay of 12–48 hours  
18 (hence the term “delayed”) (AIHA, 2003). Although there is some connection between skin and  
19 respiratory sensitization, it does not follow exact rules, and the dermal mode is a much more  
20 common reaction to metals.

#### 21 22 **4.3.6. Metal Mixtures**

23 Metal mixtures present interesting challenges for the risk assessor. As described below,  
24 certain metals can mimic other metals, and depending on the dose and composition of other the  
25 metal mixture can result in either a toxic or protective effect. Because metals are naturally  
26 occurring substances, they more frequently occur as mixtures than as single, toxic-level  
27 exposures. Thus, it is important to account for multiple, simultaneous metal exposures to  
28 realistically assess health risks.

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#### 1 **4.3.6.1. *Molecular or Ionic Mimicry***

2 A large body of literature provides examples of *molecular* or *ionic mimicry* of metals.  
3 This phenomenon is central to aspects of uptake and biokinetics for metals within the body. The  
4 term *molecular* or *ionic mimicry* has been applied to situations in which a metal forms a complex  
5 with an endogenous ligand and the resulting compound mimics the behavior of a normal  
6 substrate, disrupting normal function. A number of reviews discuss this phenomenon, giving  
7 examples of the mechanism of toxicity for specific metals (Ballatori, 2002; Clarkson, 1993).  
8 Most of these examples involve replacement of an essential metal with a nonessential metal, and  
9 *molecular* or *ionic mimicry* may be viewed as a form of metal-metal interaction. For example,  
10 the protective effects of zinc against copper toxicity are most likely due to diminished  
11 gastrointestinal uptake of copper. Lead replaces zinc in heme synthesis by inhibiting the  
12 function of heme-synthesizing enzymes (Goyer and Clarkson, 2001). The substitution of  
13 calcium by lead results in toxicity of several vital enzyme systems in the central nervous system.  
14 This toxicity impaired the development and function of enzymes involved in the production and  
15 transport of neurotransmitters (NAS/NRC, 1993). The uptake of lead from the gastrointestinal  
16 tract likely occurs via both passive diffusion processes and via active transport mechanisms used  
17 in the uptake of essential minerals such as calcium. Calcium deficiency increases the uptake of  
18 lead into the body, presumably as a result of lead uptake via calcium active transport processes.  
19 Calcium supplementation then diminish lead uptake via both competitive binding to uptake  
20 proteins and down-modulation of active transport activity. Lead is actively taken up into the  
21 body and sequestered into bone because of ionic mimicry for calcium. Similarly, cadmium  
22 uptake may in large part be related to ionic mimicry of zinc. Divalent inorganic mercury forms  
23 linear bonds that form a complex that structurally mimics oxidized glutathione. Arsenate  
24 complexes with phosphate in the sodium-dependent transport system in renal cells, and the  
25 arsenate replace the phosphate in mitochondria, impairing synthesis of ATP and energy  
26 metabolism.

27 As a converse of the enhanced toxicity that can result from nutritional deficiency, the  
28 effects of moderate doses of naturally occurring metals that are not required for nutrition can be  
29 reduced (or antagonized) by essential metals found in foods. Diet, therefore, can be a major  
30 factor in determining whether potential adverse health effects of additional metal exposures are  
31 moderated or enhanced. For example, humans can be exposed to mercury by consuming fish  
32 that have absorbed mercury from contaminated water, whereas selenium present in the same  
33 water can act as a natural antagonist for mercury toxicity, and vice versa (although the protection  
34 is inferred through formation of non-reactive mercury-selenide complexes, rather than as result

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1 of ionic mimicry); cadmium in soil can be ingested via fruits and vegetables grown on that soil,  
2 while zinc in nuts also eaten as part of that diet can limit (or antagonize) cadmium toxicity.  
3 Relative intakes of zinc, sulfur, or iron play a significant role in modulating copper deficiency or  
4 toxicity. Suttle and Mills (1966) showed that dietary levels of copper at 425 mg/kg caused  
5 severe toxicosis in pigs. However, all signs of toxicity were prevented by simultaneously  
6 supplementing the diet with 150 mg/kg zinc and 150 mg/kg iron.

#### 7 8 **4.3.6.2. *Studies of Metal Mixtures***

9 Few controlled studies exist on the toxicologic interactions of metals relevant to levels  
10 found in the environment. ATSDR has compiled and evaluated interaction studies for various  
11 combination of chemicals, including two sets consisting only of metals: (1) arsenic, cadmium,  
12 chromium, and lead; and (2) copper, lead, manganese, and zinc. Additional interaction profiles  
13 that include two or more metals are: (3) cesium, cobalt, strontium, trichloroethylene (TCE), and  
14 polychlorinated biphenyls (PCBs), and (4) arsenic, strontium-90, TCE, hydrazine, and jet fuels  
15 (available online at <http://www.atsdr.cdc.gov/iphome.html>). Few studies quantified the  
16 magnitude of the interaction, whether using the authors' definitions of toxicologic interaction or  
17 EPA's definitions based on dose and response addition. (This same limitation also applies to  
18 non-metal mixtures.) The summaries below indicate some of the qualitative conclusions  
19 available regarding potential toxic interactions.

20 A study of a mixture of cadmium, lead, and zinc study in rats found slightly more marked  
21 adverse hematological effects with the ternary mixture exposure than with the cadmium-lead,  
22 cadmium-zinc, or lead-zinc mixtures (Thawley et al., 1977); inconsistencies in dietary levels of  
23 calcium and vitamin D in this study, however, may have made comparisons problematic. A  
24 well-controlled rat study has reported significant synergism between cadmium and lead  
25 regarding testicular atrophy (Saxena et al., 1989). That study also demonstrated protective  
26 effects of high dietary levels of zinc, which effectively reduced the testicular effects of the  
27 cadmium-lead mixture to control levels. No studies have been located that would allow  
28 extrapolation of those high exposure results to more common, lower environmental levels.  
29 Fowler and Mahaffey (1978) investigated a relatively wide range of endpoints in studies that  
30 covered each metal singly and all possible binary and ternary mixtures. Body weight gain was  
31 depressed equally by the ternary mixture and the cadmium-lead mixture, and to a lesser extent by  
32 the arsenic-lead and cadmium-lead mixtures, whereas food utilization was depressed more by the  
33 ternary and arsenic-cadmium mixtures than by the other binary mixtures. In general, the

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1 biological parameters studied in this report indicated changes of smaller magnitude and  
2 inconsistency in direction for binary mixtures compared with ternary mixtures.

3 For some endpoints, the data are not sufficiently robust to show even the direction of  
4 interaction (i.e., whether the joint action will be dose additive or greater or less than additive).  
5 Many animal studies use commercial diets or semi-purified diets that may have higher or lower  
6 levels of essential metals than human diets. Much higher doses of the metals appear to be  
7 required to elicit effects when commercial diets are used than when semi-purified diets are used.  
8 At the other extreme, effects are seen at very low doses when deficient diets are used.  
9 Comparisons among studies are therefore problematic, particularly when the diets are not  
10 specified. Experimental efforts to identify and quantify interaction mechanisms among metals  
11 are still needed.

### 12 13 **4.3.6.3. Human Health Assessment**

14 Approaches for assessing joint toxicity following exposure to mixtures have been  
15 developed by EPA (U.S. EPA, 2000b, which updates U.S. EPA, 1986a). The selection of risk  
16 assessment approach begins with the assessment of data quality. Available data are classified  
17 into three categories: data on the mixture of concern, data on a mixture of a toxicologically  
18 similar nature, and data on individual effects of each component chemical in the mixture. The  
19 first approach—assessment of the data based on the mixture of concern—is the preferred  
20 approach when the data allow and is essential when considering a complex mixture (e.g.,  
21 containing hundreds of component chemicals). Such data include epidemiologic studies on the  
22 complex mixture or in vitro data on the complex mixture. The typical example of a complex  
23 mixture is coke oven emissions; few metal mixtures relevant to environmental exposures would  
24 be expected to fall within this category. The second approach is an assessment of data on a  
25 mixture of a toxicologically similar nature to that of the mixture of concern, and a similar  
26 limitation applies to the usefulness of this approach. The third approach is based on the toxic or  
27 carcinogenic properties of the components in the mixture. In this approach, information on  
28 toxicologic interactions of components is incorporated into the assessment. This last approach  
29 will likely be most useful for assessing metals in the environment, because this situation is most  
30 common (i.e., something is known about the components of the mixture but nothing is known  
31 about the mixture itself, or a similar mixture).

32 When no information on interactions exists (i.e., to indicate higher-than-additive or  
33 lower-than-additive results), a dose-additive or risk-additive model is applied as a default. The  
34 decision on which of these to use depends primarily on how toxicologically similar the metals in

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1 the mixture are. For metal mixtures, a dose-additive approach would be suggested if each metal  
2 could be thought of as a concentration or dilution of every other metal in the mixture. Three  
3 approaches have been used when the components are considered to be dose additive: the Hazard  
4 Index, the Relative Potency Factors, and the Toxicity Equivalence Factors (a special case of the  
5 Relative Potency Factors). The choice of approach will depend on the quality of the data. If the  
6 metals were assumed to behave independently of each other, then response addition would be the  
7 approach applied. This approach is often used for cancer assessments at Superfund sites and  
8 other environmental contamination situations; note that response addition is not valid for high  
9 exposure concentrations. For mixtures containing metals known to interact, then an interaction-  
10 based Hazard Index can be estimated (U.S. EPA, 2000b; Hertzberg et al., 1999).

11 Dietary information is an important part of evaluating the potential for mixture toxicity  
12 and interactions. As described above, some quantitative data exist for some metal combinations,  
13 and where sufficient data exist, they can be used to predict the pattern of interactions for various  
14 proportions of the mixture components or to quantitatively modify the risk assessment.

15 Although at this time few such data are available, some studies do exist, and in some cases the  
16 results can offer at least qualitative insights. For example, *in-vitro* studies showed that  
17 chromosome mutagenicity resulting from co-exposure to arsenic and antimony was less-than-  
18 additive. That is, less cell damage was observed than would have been expected from an  
19 additive effect from the two metals (Gerbel, 1998). However, it may be difficult to validate  
20 laboratory data in the absence of comparable field (epidemiological) data (McCarty et al., 2004).  
21 Exposures to other types of stressors also must be considered. A recent progress report on a  
22 mixtures study of metals and polycyclic aromatic hydrocarbons (PAHs) indicated that  
23 environmental metals decreased levels of the enzyme needed to catalyze PAH bioactivation (a  
24 precondition of carcinogenesis), affecting induction at both transcriptional and post-translational  
25 levels (Kaminsky et al., 2003). (Note that the carcinogenicity of the metals themselves has not  
26 yet been assessed.)

27 A number of *in vivo* studies of multiple metals and metal compounds were published  
28 before 1980. The draft interaction profiles developed by the Agency for Toxic Substances and  
29 Disease Registry and EPA include evaluations of these metals and more recent publications for a  
30 selected set of metals.

31 The EPA provisional interaction profile for arsenic, cadmium, lead, and zinc includes  
32 information relevant to chronic exposures as well as insights from acute toxicity studies. Some  
33 acute studies considered a variety of endpoints – mortality, testicular necrosis, and  
34 hepatotoxicity (Hochadel and Waalkes, 1997) – while others focused only on mortality (Yanez et

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1 al., 1991; Diaz-Barriga et al., 1990; Yoshikawa and Ohta, 1982). The studies captured in these  
2 draft profiles often include measurements of changes in chemical distributions in target tissues,  
3 which can be difficult to interpret in terms of potential health risks. Nevertheless, some specific  
4 joint toxicity insights can be gained. An example of how this information can be summarized for  
5 at least qualitative consideration in a health risk assessments is given in Table 4-13.

6 Current studies of toxicological interactions are limited, often relying on in vitro assays  
7 of bacteria and cell lines. These studies have tended to focus on mechanisms at the cellular and  
8 molecular levels, although their implications for health risk assessment on the tissue or whole  
9 organism level are not yet clear. For example, Pounds et al. (2004) identified responses in a  
10 recent study of interaction toxicity among simple chemical mixtures of cadmium, mercury,  
11 methylmercury, and trimethyltin, using cultured murine renal cortical cells that are targets for  
12 metal toxicity. Meanwhile, a model for synergistic metal activation leading to oxidative damage  
13 of DNA (genetic material, deoxyribonucleic acid) has been developed by Sugden et al. (2004).  
14 This model used in vitro assays to assess the oxidative activation of chromate by arsenite.  
15 Although toxicokinetics are a common focus of recent work, interesting new studies are also  
16 breaking ground in the area of toxicodynamics, including those under way at TNO in the  
17 Netherlands (by J. Groten and collaborators). Future studies are expected to produce key  
18 insights that will improve the ability to assess human health implications of exposures to  
19 multiple metals.

#### 20 **4.3.7. Variations in Susceptibility**

21 Many factors can contribute to human variation in susceptibility to metals. In addition to  
22 diet/nutritional status discussed above, these include other lifestyle factors such as smoking or  
23 alcohol consumption. Smoking can damage the lung, which can affect the ability to withstand  
24 the insult caused by simultaneously or subsequently inhaling metals, particularly those that act  
25 directly on the lung such as beryllium, cadmium, chromium, and nickel. Similarly, alcohol  
26 damage to the liver can be exacerbated by metals that also cause liver pathology. Other factors  
27 influencing susceptibility include age, gender, concurrent damage or disease, and genetic  
28 polymorphisms. The discussion below focuses on susceptibility issues considered key for  
29 metals; such issues should be at least qualitatively considered in identifying and evaluating  
30 potentially susceptible subpopulations for metals risk assessments. Examples for each of these  
31 additional factors are offered below.

##### 32 33 **4.3.7.1. Age**

34 Differences in the pharmacokinetic behavior of metals exist at different life stages,

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1 particularly for the nutritionally essential metals (WHO, 1996a). During the immediate postnatal  
2 period, absorption of essential metals is poorly regulated (e.g., chromium, iron, zinc) until  
3 homeostatic regulatory mechanisms become established with increasing gut maturity. Much of  
4 what is known about gastrointestinal absorption during infancy is derived from animal studies;  
5 few studies have been conducted on humans. On the other hand, numerous studies have been  
6 conducted on the effects of lead and the developing nervous system in humans (IPCS, 1995;  
7 NAS/NRC, 1993). It is suspected that the human placenta is resistant to transport of cadmium  
8 (Goyer, 1995). It has also been shown that neonate experimental animals have a higher  
9 absorption of both lead and cadmium (Kostial et al., 1978). The efficiency of intestinal uptake of  
10 some trace metals, particularly zinc, declines in the elderly, but differences between mature  
11 adults for other metals of interest to EPA were not identified in a study conducted several years  
12 ago (WHO, 1996c). For comparison, the RDA for copper for children up to 8 years old is just  
13 under half that identified for teenagers to adults (NAS, 2001).

14 The sensitivity to skin irritants is considered to generally decrease with age, so children  
15 could be more sensitive to metal irritants. Meanwhile, the general loss of renal function with age  
16 means older adults will be less able to withstand the harmful effects of metals that affect the  
17 kidney, (e.g., cadmium; U.S. EPA, 1999). Note that the RDAs specifically tabulated for older  
18 adults (FIU 2004) remains similar to those identified by NAS for this population group within  
19 the overall tables published in 2001 (FIU 2004). Thus, when assessing metal risks across  
20 different population groups, it will be important to review the current status of metal-specific  
21 information with age implications for the metal(s) in that study.

#### 22 23 **4.3.7.2. Gender**

24 Pregnancy and lactation increase demand for some essential metals, particularly copper,  
25 zinc, and iron (NAS/IOM, 2003; Picciano, 1996). Because of physiological changes that include  
26 higher iron (and calcium) requirements, hormonal changes, and susceptibility to respiratory  
27 disease, it has been suggested that pregnant women could be predisposed to the toxic effects of  
28 beryllium, lead, and manganese (U.S. EPA, 1999f). As a note, the RDAs for copper, iron,  
29 molybdenum, and selenium are higher for pregnant and lactating women than for other women  
30 or men; for selenium, the RDA for other women is the same as that for men (NAS/NRC, 2000);  
31 in contrast, the RDA for zinc is the same for pregnant women 19 to 50 years old as it is for men  
32 aged 19 to 70 and older. For additional comparison, the RDA for chromium for pregnant women  
33 is roughly the same as that for men, which is higher than for non-pregnant women and lower  
34 than for lactating women (NAS/NRC, 2001).

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1           References to women as being highly susceptible to metal toxicity usually refer to effects  
2 on the fetus during pregnancy (e.g., of lead and mercury), but basic gender differences  
3 independent of pregnancy also may account for differences in toxicokinetics between women  
4 and men. Women have only about two-thirds the fat-free body mass of men—so their protein  
5 and energy requirements are lower—while having a larger percentage of body fat. The  
6 male/female ratio for urinary creatinine excretion (an index of body muscle mass) is 1.5. Men  
7 are generally larger than women. Skeletal size as well as body calcium are a function of height,  
8 and calcium can also be lost as women age. These differences have an impact on body content  
9 of minerals (WHO/IPCS, 2002). Women also have significant loss of iron during menstruation,  
10 and it has been shown that absorption and toxicity of cadmium are greater in women, related to  
11 decrease in iron stores (Berglund et al., 1994).

#### 13 **4.3.7.3 Concurrent Damage or Disease**

14           People with higher-than-average biological sensitivity to environmental stressors include  
15 allergics and those with pre-existing medical conditions (e.g., with compromised immune  
16 systems as a result of a disease or treatment for it, such as chemotherapy). For example,  
17 chemotherapy can damage the kidney over time, and at certain levels other medicines (such as  
18 acetaminophen) can damage the liver. This may increase metal sensitivity for those metals that  
19 cause liver dysfunction.

20           Skin abrasions or other irritations also can alter exposures to and subsequent effects of  
21 metals (although dermal absorption is not a primary route of metals exposure for intact skin).  
22 For example, both nickel and chromium can cause allergic contact dermatitis, so their combined  
23 presence could result in joint toxicity with a potential for interaction; broken skin could  
24 potentially increase the absorption of other metals (and other toxic agents) across the exchange  
25 boundary. Beyond these well-established contact allergens, other metals have also been cited as  
26 possible contact allergens, including copper (WHO/IPCS, 1998), cobalt salts (AIHA, 2003),  
27 beryllium (WHO/IPCS, 1990b), palladium (Kimber and Basketter, 1996), and gold (Kimber and  
28 Basketter, 1996).

29           With regard to effects associated with high-dose metals exposures, elevated arsenic  
30 intakes – from drinking water with levels 17 times above the U.S. drinking water standard – have  
31 been linked to blackfoot disease, a vascular complication that represents a severe form of  
32 arteriosclerosis (EPA 2004). like many such diseases, the incidence of blackfoot increases with  
33 age and thus is also linked to the age factor. Some studies suggest that higher doses of copper  
34 and manganese might be linked with Parkinsonism (which could be reversible) (Feldman, 1992;

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1 Gorell et al., 1999). Thus, these studies offer insights for specific metals risk assessments where  
2 exposure conditions, including levels and durations, were sufficiently similar.

#### 3 4 **4.3.7.4. Genetic Polymorphisms**

5 Individuals vary considerably in the nature and severity of their response, and potential  
6 repair and recovery, from exposure to metals and metal compounds. Such differences may be  
7 due to genetic polymorphisms that can alter the transport and metabolism of a metal. The most  
8 apparent of these genetic polymorphisms affecting metabolism and toxicity of metals are  
9 disorders in homeostatic mechanisms for nutritionally essential metals.

10 Two disorders affect copper metabolism: Wilson disease and Menkes disease. Wilson  
11 disease is an autosomal recessive abnormality (prevalence of 1 in 30,000) that is believed to be  
12 due to impaired biliary excretion of copper, resulting in accumulation in and damage of various  
13 tissues, particularly the liver, brain, kidney, and cornea; hemolytic anemia can also result.  
14 Menkes disease is an X-linked recessive disorder of copper metabolism (prevalence of 1 in  
15 200,000) that resembles copper deficiency regardless of level of copper intake (WHO/IPCS,  
16 2002).

17 Hemochromatosis is a common inherited disorder related to iron homeostasis. This  
18 disorder is characterized by excessive iron absorption, elevated plasma iron concentration, and  
19 altered distribution of iron stores (altered iron kinetics). One long-term effect is liver cirrhosis,  
20 with increased risk of liver cancer (NAS/IOM, 2003).

21 ATSDR (2003) reported that a relationship between human leukocyte antigens (HLA)<sup>1</sup>  
22 and nickel sensitivity was observed in patients who had a contact allergy and positive results in a  
23 patch test for nickel. The nickel-sensitive group had a significant elevation in HLA-DRw6  
24 antigen compared with normal controls. The relative risk for patients with DRw6 to develop a  
25 sensitivity to nickel was approximately 11-fold.

26 A genetic polymorphism for a heme-metabolizing enzyme affecting lead metabolism was  
27 identified in 1973 (Granick et al., 1973), but the molecular characteristics and potential clinical  
28 implications did not receive attention until about ten years ago (Smith et al., 1995). Fleming et  
29 al. (1998) found that the relationship of bone lead to the cumulative blood index for workers

---

<sup>1</sup> The major histocompatibility complex is a group of genes on chromosome 6 that code for the antigens that determine tissue and blood compatibility. In humans, histocompatibility antigens are called human leukocyte antigens because they were originally discovered in large numbers on lymphocytes. There are thousands of combinations of HLA antigens.

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1 with occupational exposure was greater in those workers with the ALAD1 allele, suggesting that  
2 the ALAD2 genotype decreased transfer of lead from blood to bone. This effect was only  
3 demonstrated in workers with higher blood lead levels than the general population.

4 It is suspected that genetic polymorphisms also exist for arsenic metabolism (NAS/NRC,  
5 2001), but these have not yet been defined. Other genetic polymorphisms that may affect the  
6 metabolism of chemicals continue to be described. The human MT gene locus is very complex;  
7 four individual isoforms and more than ten subforms of the MT1 gene exist, including functional  
8 and nonfunctional genes. Because of the protective role that MT plays in metal metabolism, an  
9 understanding of the biological role of significant variations in this MT gene locus (both within  
10 and among individuals) can offer insights into differential sensitivity or susceptibility and will  
11 help characterize variability across a population.

12 Although all MT proteins are associated in some way with a protective role, each of the  
13 multiple isoforms appears to have a unique function, as reflected by their unique tissue  
14 distributions, the specific conditions under which each is expressed, and unique properties of the  
15 metal binding clusters (Bogumail et al., 1998). The multiplicity of the human MT gene loci  
16 makes them potentially important for understanding genetic variability. Emerging technologies  
17 are enabling new research in the areas of genomics, proteomics, and metabolomics that will  
18 eventually improve human health risk assessments for metals and other stressors, including the  
19 role that MT isoforms play in variability in response.

20 Other creative approaches have coupled human tissue studies with animal toxicology  
21 studies. For example, studying lung slices from humans (from transplant or tumor resection)  
22 and from rodents exposed to the same metals offers promise for improving our understanding of  
23 potential effects from inhalation exposures. Comparison of study results between rats and mice  
24 under similar exposure regimes and linking them to similar human exposures will provide  
25 insights into whether human responses are better predicted by rats or by mice.  
26

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1 **4.4. ECOLOGICAL EXPOSURE PATHWAY ANALYSIS**

2 Risk is a function of both hazard (i.e., the toxicity of a substance) and level of exposure.  
3 The route of exposure, as well as other details such as timing and duration, is also important  
4 because exposures vary in their effectiveness in delivering a dose across a biological boundary  
5 (Ryan, 1998; Ott, 1985). No risk exists unless an effective exposure to a receptor occurs.

6 *Exposure routes* include inhalation, ingestion, and  
7 dermal or, for plants, root uptake and leaf exposures.  
8 *Pathways* describe the specifics of any exposure and  
9 include transport of the contaminant in the environment as  
10 well as exposure route for organisms of concern (e.g.,  
11 dietary ingestion of a soil contaminant that has been taken  
12 up by plants). *Phase association* and *chemical speciation*  
13 influence metal movement through pathways and the  
14 availability of a metal for delivery of the dose to receptors of concern.

<p><b>Metal Exposure Assessment</b></p> <p><i>Metal exposure assessment</i> includes consideration of the exposure routes and pathways specific to metals, the phase associations and chemical forms of the metals, and the expression of exposure and target doses in a manner consistent with defining hazard thresholds for particular organisms.</p>
--

15 Major metal sources to waters and lands include diverse manufacturing, mining,  
16 combustion, and pesticide activities. Major atmospheric sources are oil and coal combustion,  
17 mining and smelting, steel and iron manufacturing, waste incineration, phosphate fertilizers,  
18 cement production, and wood combustion (Haygarth and Jones, 1992). Metals from these  
19 atmospheric sources can find their way into soils, sediments, and water. Other major sources to  
20 aquatic and terrestrial systems include chlor-alkali, acid, pigment, electronics, and copper sulfate  
21 production.

22 Some exposure assessments do not involve anthropogenic releases of metals to the  
23 environment. Rather, they focus on changes in exposure to ambient metals that result from other  
24 aspects of human activities. For example, acidification of freshwater changes aluminum phase  
25 association and speciation, resulting in an increased dose of naturally occurring metals to aquatic  
26 biota (e.g., Campbell et al., 1992). Intensive irrigation mobilizes selenium that is naturally  
27 present in relatively high concentrations in western soils, and consequent evaporative  
28 concentration in wetlands, impoundments, and other low-lying areas in arid regions of the  
29 United States can lead to toxic exposures (e.g., Wu et al., 1995).

30 Methods for assessing transport of metals through various media (water, soil, air) are  
31 described in this section. Exposure to aquatic organisms through water and dietary routes  
32 (looking again at models to account for site-specific differences in bioavailability) is also

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1 discussed. The section concludes with discussions about quantifying exposure to metals for  
2 terrestrial organisms.

#### 3 4 **4.4.1. Aquatic and Terrestrial Transport Pathways for Metals**

5 When an exposure assessment is completed for a site, the available data are usually  
6 limited in their coverage with respect to the characterization of exposure levels over both time  
7 and space. Depending on the situation, it may be advantageous to use a transport and fate model  
8 (i.e., a computational model) to fill in the data gaps, such that an improved characterization of  
9 exposure levels is available for use by the risk assessor. Models are also useful in situations  
10 where it is desirable to have an estimate of future exposure levels that are expected to result from  
11 the implementation of remediation measures. These results can be used to quantitatively  
12 evaluate the effectiveness of alternative remediation scenarios that are being considered. The  
13 models can also be used to refine the design of the viable alternatives so that an optimal  
14 remediation strategy can be developed.

15 Although numerous models are available for use, most are based on the same  
16 fundamental principles. That is, metals are ubiquitous in the environment, as they are found in  
17 the aquatic, terrestrial, and atmospheric compartments. Within each compartment, they are  
18 present in association with water (freely dissolved metal or as organic and inorganic metal  
19 complexes), particles (sorbed, precipitated, or incorporated within a mineral phase), and air. The  
20 evaluation of metal transport therefore requires evaluation of the distribution of the metal among  
21 these phases, within each compartment, as well as the movement (i.e., the transport) of each of  
22 these within and among the various compartments. It is important to simulate the movement of  
23 water and particles explicitly because this provides a way to evaluate differences in the degree to  
24 which various chemicals/metals may be transported in association with particles (i.e., via settling  
25 and resuspension) or in association with the dissolved phase (diffusive flux of dissolved metal).  
26 The analyst represents the environmental setting of interest as a series of discrete, interconnected  
27 volumes. Mass balance equations for air, water, solids, and metal are then formulated for each  
28 volume to obtain a system of mass balance equations that may then be solved for the  
29 concentrations of interest over both time and space. Note that it is not necessary to include all of  
30 the compartments in every model. For example, models for a site that is impacted by a smelter  
31 might call for use of a model of an atmospheric compartment (e.g., to simulate transport of a  
32 release from a stack) and a terrestrial compartment (to simulate fate of atmospheric inputs to the  
33 soil). Alternatively, for an aquatic setting dominated by previously contaminated sediments, it

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1 may be sufficient to consider water and sediment transport alone and to neglect the atmospheric  
2 and terrestrial compartments.

3 Although the underlying principles of most models are similar, the features included vary  
4 widely from one model to the next. The output from a relatively simple model may be adequate  
5 for decision-making purposes in some instances. Some relatively simple models are limited in  
6 their applicability to steady-state analyses and spatially uniform conditions, but if this will  
7 provide a reasonable and/or conservative representation of conditions at a site, it may provide the  
8 analyst with a useful and cost-effective modeling alternative. In other cases, the analysis may  
9 require the completion of time-variable simulations to properly represent conditions that vary  
10 over time, such as daily or seasonal variations in flow and upstream boundary concentrations,  
11 point source loads, and pulse exposures. The analyst should select is an appropriate model  
12 because not all models will be applicable to every situation. Although in principle the more  
13 sophisticated models provide the risk assessor with the capability to complete a more detailed  
14 and mechanistically based analysis than will a simple model, successful application of these  
15 models will require greater resources (data, time, and funding) than will the use of a simpler  
16 model, and that the analyst also must possess a relatively high level of modeling expertise. It is  
17 for this reason that the more sophisticated models are usually reserved for use in higher level,  
18 definitive assessments.

19 Many of the models available for use in evaluating the transport and fate of metals were  
20 originally developed for application to neutral organic chemicals. As a result, these models  
21 frequently include a variety of reactions that are not necessarily germane to an analysis of metal  
22 transport and fate (e.g., biodegradation, photo-oxidation, and volatilization). Although these  
23 models still may be of use in an exposure assessment for metals (the nonapplicable processes  
24 often may be bypassed), a more significant problem is that they often fail to represent some  
25 important metal-specific processes. For example, the evaluation of metal speciation and metal  
26 partitioning between dissolved and particulate phases will be represented only in a very simple  
27 manner in such models. This limitation may be overcome, at least in part, by performing the  
28 requisite metal-specific analyses with a stand-alone chemical equilibrium model, but this  
29 approach will place an added burden on the analyst to integrate the results of the two models in a  
30 technically defensible manner.

31 An overview of some of the aquatic fate and transport models available for use is  
32 presented in the following subsection. Although models that include some metal-specific  
33 capabilities will be noted, no single model that is currently available for use includes all of the

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1 metal-specific features that would be desirable for use. More detailed discussions of these and  
2 other transport and fate models, as well as a number of chemical equilibrium models, may be  
3 found in Paquin et al. (2003) for aquatic systems and Allen (2002) for terrestrial systems.  
4 Additionally, work is in progress to develop updated models that will offer improved metal-  
5 specific capabilities (e.g., see the discussion of the Unit World model in Section 5). As a result,  
6 metal fate and transport models should be viewed as an evolving technology, with new models  
7 expected to become available in the not too distant future.

#### 8 9 **4.4.1.1. Aquatic Transport Models**

10 Modeling of metal transport and fate within aquatic systems involves the representation  
11 of hydrodynamic transport to simulate movement of water, particulate transport to simulate the  
12 movement of particles, and chemical transfers and kinetics to simulate exchange of metal  
13 between dissolved and particulate phases and between the water column and benthic sediment  
14 (Figure 4-5). The analyst has the option of using independent hydrodynamic transport, sediment  
15 transport and chemical fate models, or an integrated model that incorporates all of these  
16 processes. Although the models described below are mainly in the latter category, their use does  
17 not preclude the use of a stand-alone model (e.g., a hydrodynamic or sediment transport model)  
18 as an aid in the development of inputs to the integrated model.

19 Modeling the movement of metals through an aquatic system begins with a  
20 characterization of the movement of water through the system. The time scale for the  
21 hydrodynamic analysis should be represented in a way that will satisfy the needs of the sediment  
22 transport and chemical fate analyses that are also being performed. For example, low-flow  
23 conditions associated with minimum dilution may be judged to be the most critical conditions in  
24 a setting involving a point source discharge, while peak flow conditions may need to be  
25 simulated in a setting where resuspension of contaminated sediments is the primary concern. A  
26 steady-state model might be appropriate for use in the former case, and a time-variable model  
27 would likely be needed in the latter case. Thus, the details of the specific problem setting will  
28 necessarily have an influence on both model selection and how the model will be used.

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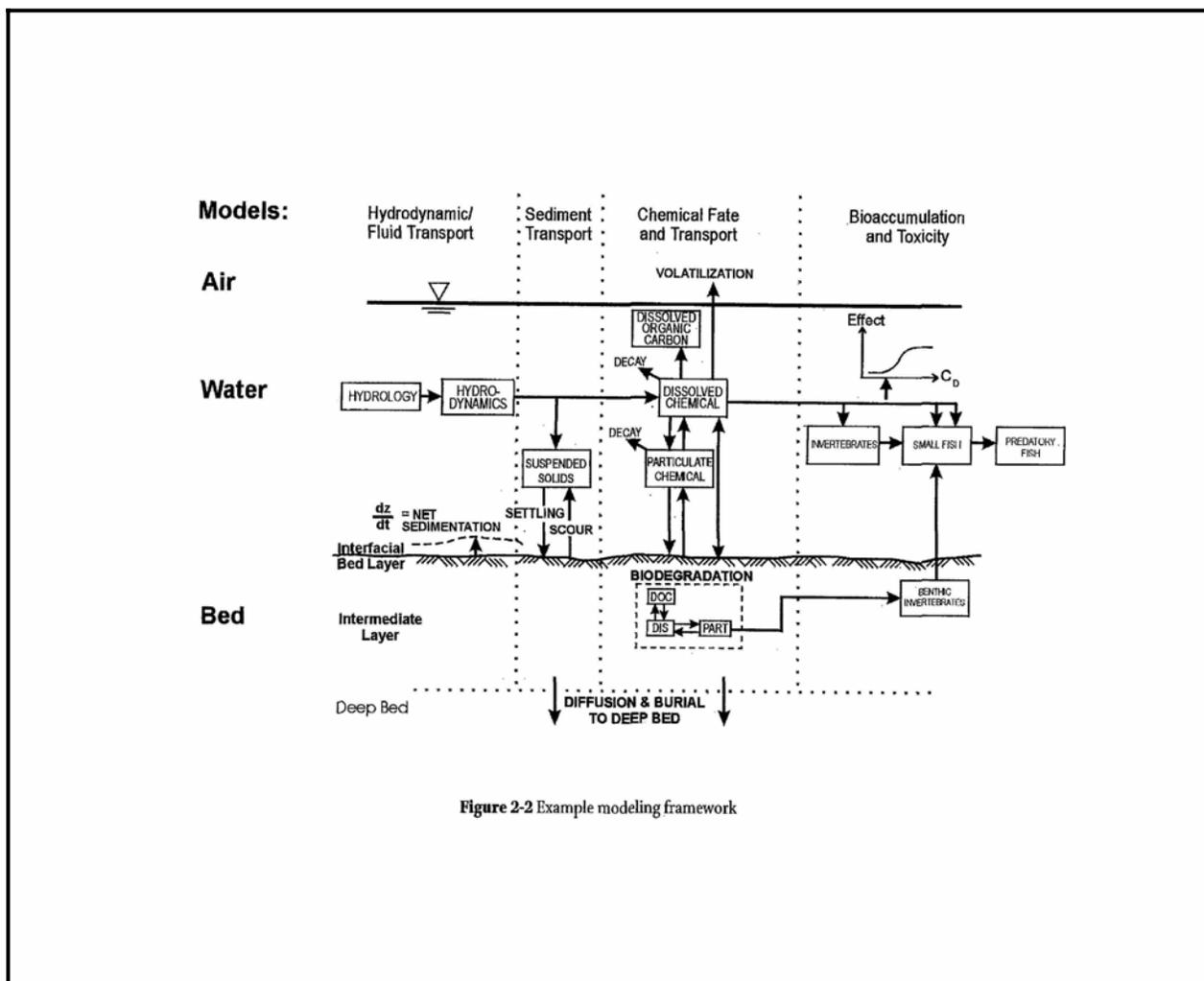


Figure 2-2 Example modeling framework

Figure 4-5. A generalized model framework for chemical fate and transport in an aquatic system.

Source: Paquin et al., 2003.

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1 The reader also should be aware of several reviews of fate and transport models that have  
2 been completed since the early 1980s. In earlier reviews, Delos et al. (1984) reviewed models  
3 for wastewater load allocations, and Mills et al. (1985) described screening-level analyses for  
4 water quality assessments of conventional pollutants. The Agency subsequently prepared an  
5 updated summary of the features included in the water quality models that were available for use  
6 in exposure assessments (U.S. EPA, 1987). Schnoor et al. (1987) simultaneously published their  
7 review and included detailed descriptions of fate and transport models and their required input  
8 parameters. Later, EPA published a review describing the use of modeling tools for the  
9 development of total Maximum Daily Loads (TMDLs) in watersheds (U.S. EPA, 1997a). More  
10 recently, Paquin et al. (2003) completed a review of exposure, bioaccumulation, and toxicity  
11 models for aquatic systems, with a focus on their applicability to metals (exclusive of  
12 organometallics). Because of the advances that have been made since the mid-1990s with regard  
13 to the development of fate and transport models, including some recent efforts to couple these  
14 models with metal speciation models and more sophisticated stand-alone hydrodynamic and  
15 sediment transport models, the latter reviews by U.S. EPA (1997a) and Paquin et al. (2003) tend  
16 to include the most up-to-date information with regard to the availability of models that are  
17 appropriate for use. These reviews also include example applications of many of the models  
18 discussed.

19  
20 **4.4.1.1.1. Applications.** As noted above, fate and transport  
21 analyses may be performed by using an integrated  
22 hydrodynamic, sediment, and chemical transport model or  
23 by employing what tend to be relatively sophisticated stand-  
24 alone versions of these three submodels. The advantage of  
25 the former approach is that integration of the hydrodynamic,  
26 sediment, and chemical transport results takes place in a seamless manner with limited need for  
27 an analyst's intervention. This is in contrast to the use of stand-alone models, where the output  
28 of one model should be formatted in a way that ensures it is amenable to use with the other  
29 models that are to be applied. A distinct advantage of the latter approach is that it has the  
30 potential to reduce the time needed to complete a model run, an important consideration for a  
31 multiyear simulation of a large and complex problem setting. For example, it may not be  
32 necessary to repeat the simulation of hydrodynamic and sediment transport if the model input  
33 being modified affects only chemical transport (e.g., partition coefficient). A similar line of

**Integrated Model**

The fate and transport of metals in aquatic systems is most reliably predicted using *integrated models*, rather than stand-alone hydrodynamic or sediment transport models.

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1 reasoning would apply to the use of an integrated transport model that includes metal speciation  
2 versus one that uses a stand-alone chemical equilibrium model to satisfy this need.

3 The fate and transport models that may be considered  
4 for use in the analysis of an aquatic setting are listed in Table  
5 4-13 and discussed below. The models range in complexity  
6 from simple mass-balance calculations that can be performed  
7 on a hand calculator, to one-dimensional steady-state  
8 models, to multidimensional time-variable models. The  
9 models are listed in groups that correspond to the  
10 mathematical solution technique used to solve the governing mass-balance equations (analytical  
11 solution, steady-state numerical solution, and time-variable numerical solution), a categorization  
12 that is approximately in accordance with their ease of use as well. Paquin et al. (2003) describe  
13 example applications of many of these models to illustrate how they have been previously  
14 applied. Although publications of successful modeling applications serve as useful illustrations  
15 of how models may be used, it is important to bear in mind that “past performance should not be  
16 viewed as a promise of future returns.” That is, the successful application of any model will  
17 depend on many factors other than the model framework itself, including the complexity of the  
18 problem setting, the data that available for use, and the experience of the analyst. Another  
19 important point to keep in mind is that a complicated model is not necessarily a “better” model  
20 to use than a simple one. Models should be selected on a case-by-case basis.

**Analytical Solution Models**

*Analytical solution models* of aquatic system fate and transport are the simplest models with the lowest computational requirements. They are solved analytically to provide concentrations over metal of time or space.

21 The simplest modeling analysis to consider involves simple screening-level calculations.  
22 While clearly applicable to a screening level of analysis, such calculations can frequently  
23 provide a quantitative perspective on the severity of a problem, one that will be useful in  
24 defining the nature of subsequent modeling analyses that may be required.

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**Table 4-13. List of fate and transport models**

<b>Abbreviation</b>	<b>Name</b>	<b>References</b>
<b>Analytical Solution Models:</b>		
WQAM	Water Quality Assessment Methodology	Mills et al. (1985, 1982a, b)
DJOC-III	Donald J. O'Connor (1988), a triad of papers	O'Connor (1988)
RIVRISK	RIVer RISK	Grieb (1995); EPRI (1996)
SLSA	Simplified Lake and Stream Analysis	Di Toro et al. (1981) HydroQual (1982)
USES	Uniform System for Evaluation of Substances	RIVM et al. (1994), as described in Johnson and Luttik (1995)
QWASI	Quantitative Water Air Sediment Interaction	Mackay (1991); Mackay et al. (1983)
TRANSPEC	Transport and Speciation	Bhavsar et al. (2004)
<b>Steady-State Numerical Solution Models:</b>		
CTAP	Chemical Transport and Analysis Program	HydroQual (1982, 1981)
PAWTOXIC	PAWtuxent TOXICs	Wright (1987)
SMPTOX3 (Version 2)	Simplified Method Program - Variable - Complexity Stream Toxics Model	LTI (1992) Dilks et al. (1995, 1994)
MEXAMS	Metals EXposure Analysis Modeling System Includes EXAMS and MINTE	Felmy et al. (1984) Burns et al. (1982)
<b>Time-Variable Numerical Solution Models:</b>		
EXAMSII	EXposure Analysis Modeling System - II	Burns and Cline (1985); Burns et al. (1982)
RIVEQLII	RIVEr QuaLity II	Chapman (1982)
WASTOX	Water Quality Analysis Simulation of TOXics	Connolly and Winfield (1984)
RCATOX (AESOP)	Row-Column AESOP for TOXics (Advanced Ecological Systems Operating Program)	HydroQual (2003)
WASP5	Water Quality Analysis Simulation Program, Version 5 Distributed w/ DYNHYD5, Dynamic Hydrodynamics 5	Ambrose et al. (1993) Ambrose et al. (1993)
META4	Metal Exposure and Transformation Assessment Model	Medine (1995) Martin and Medine (1998)
DELFT3D	Delft 3D Model	Delft Hydraulics (1998)
MIKE21 HSPF	MIKE21 Hydrologic Simulation Program - FORTRAN	Danish Hydraulic Institute Bicknell et al. (1993)
RECOVERY	RECOVERY	Boyer et al. (1994) Ruiz et al. (2000)
EFDC	Environmental Fluid Dynamics Code	Hamrick (2002)

Source: Paquin et al. (2003).

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1 Water Quality Assessment Methodology (WQAM) is an excellent resource for use in  
2 these types of analyses. WQAM describes a variety of simple screening-level procedures that  
3 may be used in water quality assessments. It provides a useful overview of the types of  
4 simplified analyses that may be performed and is a generic framework that does not generally  
5 attempt to address metal-specific modeling needs. An analyst having limited water quality  
6 modeling experience would probably find this to be a useful reference.

7 A series of three papers by O'Connor (1988) also provide a helpful introduction to the  
8 basic concepts and use of fate and transport models. These publications include analytical  
9 solutions and example applications for a variety of conditions, including spatially varying  
10 suspended solids concentrations. Although these solutions are not currently available in the form  
11 of a computer program, they could be readily adapted for this purpose.

12 Analytical solution models tend to be inherently simple models, a requirement of the  
13 solution technique used. As a result, they tend to have modest development and computational  
14 requirements (an advantage), but, at the same time, they limit the analyst with regard to the level  
15 of detail that can be used to represent the problem setting with the model. For example, these  
16 models normally allow the user to represent the system as a one-dimensional, uniform cross-  
17 section stream or a completely mixed water body, with constant inputs over time, and particulate  
18 and dissolved exchange of chemicals between the water column and sediment. Their simple  
19 structure makes these models well suited for use in screening level analyses. Models in this  
20 category include the River Risk (RIVRISK) model, Simplified Lake and Stream analysis (SLSA)  
21 model, and Uniform System for the Evaluation of Substances (USES) model. Each of these  
22 models includes a water column and a single-layer sediment compartment. RIVRISK is a  
23 steady-state model that may be applied to a one- or two-dimensional stream setting. It is one of  
24 the few models in Table 4-13 that includes a bioaccumulation subroutine. However, it is a  
25 proprietary model of the Electric Power Research Institute and is available only to registered  
26 users. The SLSA program (HydroQual, 1982; Di Toro et al., 1981) may be used to represent a  
27 one-dimensional stream or a completely mixed lake. Its utility in time-variable mode was  
28 demonstrated by its success in simulating the long-term (about 5 years) recovery of a quarry that  
29 had been dosed with instantaneous releases of the organic chemicals DDE and lindane (Di Toro  
30 and Paquin, 1984). Finally, the USES model was developed in Europe to model dissolution and  
31 movement of antifoulant paints on boats (Johnson and Luttk, 1995). It provides the analyst with  
32 a simplified way to estimate concentrations of organic chemicals and metals in a completely  
33 mixed water body, with successful model applications having been reported for metals (Johnson

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1 and Luttkik, 1995). USES has recently been replaced by a considerably more complex version,  
2 the European Union System for the Evaluation of Substances (EUSES) model (Berding et al.,  
3 1999).

4 Another analytical solution model that has been refined to include metal-specific features  
5 is the Quantitative Water Air Sediment Interaction (QWASI) model (Mackay, 1991; Mackay et  
6 al., 1983). QWASI, developed as a fugacity model for application to lakes and rivers, is used to  
7 simulate the exchanges of chemicals with measured vapor pressure between air, water, and  
8 sediments (the Unit World Model for organic substances). A numerical solution version was  
9 subsequently developed, one that considers multiple species, as occurs with metals (Diamond et  
10 al., 1992). More recently, it has been coupled with MINEQL to allow for interconversion of  
11 species and consideration of metals in dissolved, colloidal, and particle phases in water. It is  
12 known as the TRANSport and SPECiation (TRANSPEC) model (Bhavsar et al., 2004) and is not  
13 yet applicable to sediments.

14 The remainder of the models listed in Table 4-13 and described below are numerical  
15 solution models. The first four are applicable to steady-state conditions and the remainder to  
16 time-variable conditions. The steady-state models are appropriate for use in a screening level or  
17 definitive level of analysis. Their main advantage in comparison to analytical solution models is  
18 their capability to represent more complex system geometry and transport regimes (at the  
19 expense of an increased level of effort required to set up the model). The Chemical Transport  
20 and Analysis Program (CTAP) is essentially a numerical solution version of SLSA, with the  
21 added capability of being able to represent two- and three-dimensional systems and multiple-  
22 particle size classes. Pawtuxent Toxics (PAWTOXIC) is a relatively simple one-dimensional  
23 model for estimating particle deposition or resuspension in river and streams, assuming EqP  
24 conditions. PAWTOXIC does not include exchange of dissolved metals between water and  
25 sediments, and it can be used only to simulate net settling or resuspension.

26 The Simplified Method-Program Variable-Complexity Stream Toxics (SMPTOX) is of  
27 particular interest for metals because it represents one of the first models to include the  
28 simulation of AVS and SEM (see Section 3.4) for copper, cadmium, nickel, lead, and zinc. It  
29 does not consider the potential for oxidation of metal sulfides. SMPTOX, another model that has  
30 evolved from the SLSA framework, is applicable for use in simulating a one-dimensional river  
31 or stream and it also includes both particle settling and resuspension and a diffusive flux of  
32 dissolved metals between water column and sediments. As a Windows-based model, it is  
33 relatively simple to use.

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1 The last steady-state model to be considered, Metals Exposure Analysis Modeling  
2 Systems (MEXAMS), links EXAMS (a widely used fate and transport model developed by  
3 EPA) with MINTEQ, a chemical equilibrium model (Felmy et al., 1984) (see Section 4.1,  
4 Environmental Chemistry, for further discussion of MINTEQ). MEXAMS has not yet been  
5 coupled with EXAMSII (a time-variable model), so it can handle only steady-state conditions.  
6 EXAMS can be applied to a one-, two-, or three-dimensional system and includes an interactive  
7 bed layer. By transferring total metal concentrations to MINTEQ in an iterative process,  
8 EXAMS is able to calculate fate and transport of metal species. It requires the user to specify a  
9 bulk exchange coefficient to represent water-sediment exchange.

10 The remaining models listed in Table 4-13 include the capability to perform time-  
11 variable simulations. As a general rule, such models also can be used to simulate steady-state  
12 conditions, such as determining simple waste-load allocations during critical low-flow  
13 conditions. Input requirements will be relatively high for time-variable models, as the user may  
14 need to specify time variable inputs (e.g., upstream and tributary flows and concentrations, point  
15 source loads). The Exposure Analysis Modeling System II (EXAMSII, Version 2.97) is a  
16 relatively well-known model that was originally developed for steady-state applications.  
17 EXAMSII also represents bulk exchange of water and solids between the water column and  
18 sediment, rather than separate terms for particulate and diffusive fluxes of metal. It has the  
19 advantage of being very flexible with regard to execution of both simplified steady-state  
20 assumptions and more realistic (albeit more complex) time-variable analyses. It has also been  
21 widely used for many years and is well documented.

22 River Quality II (RIVEQLII) (Chapman, 1982) was developed specifically for inorganic  
23 substances. It incorporates relatively sophisticated chemical equilibrium calculations through  
24 linking with MINEQL (Westall et al., 1976) (see Section 4.1.5.1 for further discussion of  
25 MINEQL). RIVEQLII is applicable to one-dimensional rivers and streams. It assumes  
26 equilibrium conditions and incorporates particle setting to the bed, sorption reactions, and  
27 precipitation and dissolution of chemical to and from the sediment bed.

28 The next four models listed in Table 4-13, WASTOX, RCATOX, WASP5, and META4,  
29 evolved from an earlier model, the Water Quality Analysis Simulation Program (WASP)  
30 (DiToro et al., 1981). The Water Quality Analysis Simulation of Toxics (WASTOX) (Connolly  
31 and Winfield, 1984) incorporates particulate and dissolved transport processes that are very  
32 similar to SLSA and CTAP. Row-Column AESOP for Toxics (RCATOX) (HydroQual Inc.,  
33 2003) is a more recent implementation of WASTOX, one that has the potential to take advantage

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1 of parallel processing capabilities. Since RCATOX is still under development, it has not yet  
2 been released for general use.

3 The Water Quality Analysis Simulation Program, Version 5 (WASP5) (Ambrose et al.,  
4 1993) is the version of WASP that is currently supported by EPA. Essentially, WASP5  
5 incorporates many of the EXAMS capabilities, plus refined sediment transport capabilities.  
6 WASP5 is distributed with two different subroutines: EUTRO, which is used in eutrophication  
7 problems, and TOXI5 for simulating fate and transport. The WASP5 package also includes an  
8 associated food chain model. Notably, WASP5 and RCATOX are designed to interface with  
9 state-of-the-art hydrodynamic and/or sediment transport models, a feature not shared by any of  
10 the other available fate and transport models. This capability would be of use in estuarine and  
11 coastal systems having complex hydrodynamic conditions. WASP5 interfaces with the PC-  
12 compatible hydrodynamic model DYN5 (Ambrose et al., 1993). WASP5 and RCATOX can  
13 both interface with the Estuary, Coastal, Ocean Model (ECOM), a family of hydrodynamic  
14 models (Blumberg and Mellor, 1987), and with ECOMSED (HyroQual, 1998), a sediment  
15 transport model.

16 The Metal Exposure and Transformation Assessment Model (META4) model is  
17 essentially WASP5 combined with MINEQL. It is used for developing TMDL waste-load  
18 allocations and evaluating remedial actions and TMDLs (Martin and Medine, 1998). It is  
19 applicable to a variety of receiving waters, including ponds, streams, rivers, lakes, and estuaries,  
20 and can be run as one-, two-, or three-dimensional systems. META4 addresses some of the  
21 shortcomings in WASP4 to more accurately describe metal dynamics by, for example, the recent  
22 addition of subroutines to represent the interactions of dissolved metals with iron oxyhydroxides  
23 in the water column and sediment under variable pH and the ability of the model to predict future  
24 concentrations of the major cationic metals under variable regimes of water chemistry. It also  
25 handles numerous point and nonpoint loads and sequential deposition or scouring of sediment  
26 bed layers.

27 The Delft 3D model (DELFT3D), developed by Delft Hydraulics Lab in The  
28 Netherlands, is another model that is quite sophisticated with regard to its capabilities. It is a  
29 flexible integrated model that includes the following modules: hydrodynamics, water quality  
30 (including sediment transport), chemistry, and wave generation. Although the model software is  
31 proprietary, the model itself is commercially available.

32 MIKE21, developed by the Danish Hydraulic Institute (DHI), is another commercially  
33 available fate and transport model that is applicable to rivers. This three-dimensional model

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1 includes both particulate and dissolved transport processes. One of the modules of a recent  
2 release may be used to simulate the fate, transport, and bioaccumulation of metals.

3 The Hydrologic Simulation Program-FORTRAN (HSPF) (Bicknell et al., 1993) is the  
4 most recent version of a family of models originally developed in the 1960s to simulate  
5 agricultural runoff. It is unique among the models listed in Table 4-13 because that it includes  
6 hydrology and nonpoint-source terrestrial runoff modules, in addition to in-channel fate and  
7 transport capabilities. HSPF is frequently used to simulate complex watersheds and provides  
8 information for continuous input simulations or for storm events. It models well-mixed  
9 reservoirs as well as branched river systems and can include up to three sediment types. It has  
10 been widely used throughout the United States, but its high level of sophistication makes it most  
11 appropriate for use by experienced modelers.

12 RECOVERY, a model supported by the U.S. Army Corps of Engineers, has been  
13 developed with features that make it amenable to evaluating sediment capping alternatives (Ruiz  
14 et al., 2000; Boyer et al., 1994). The model is structured as a well-mixed surface-water layer  
15 overlying a vertically stratified but horizontally well-mixed sediment bed. The sediment is  
16 defined by three zones (surface, deep contaminated, and deep clean), which can be further  
17 subdivided on the basis of porosities, contaminant concentrations, and other factors.  
18 RECOVERY is particularly useful for evaluating sediment-capping scenarios and sites with old  
19 contamination (where clean sediment has layered over the contaminated bed).

20 One other model that is nearing release for use as a fate and transport model is the  
21 Environmental Fluid Dynamics Code (EFDC) (Hamrick, 2002). EFDC, originally developed as  
22 an advanced level hydrodynamic model, was subsequently modified to include sediment  
23 transport capabilities. It is a state-of-the-art model that incorporates a variety of options for  
24 representing sediment transport. Most recently, it was further refined for use with chemicals,  
25 with an initial application to metals in which two-phase linear partitioning was used (Ji et al.,  
26 2002); a second application was the ongoing analysis of polychlorinated biphenyls (PCBs) on  
27 the Housatonic River in Connecticut. The chemical fate subroutine does not currently evaluate  
28 metal speciation. EFDC is scheduled for release in the near future.

29 Water quality analyses often require probabilistic results, as the WQC in the United  
30 States are expressed in probabilistic terms, with an allowable 1-in-3-year exceedance frequency.  
31 Steady-state models cannot evaluate a return period for exceedences. When using these models,  
32 a Monte Carlo analysis can be conducted to generate a large number of model inputs and  
33 subsequent solutions that can then be analyzed probabilistically to obtain a characterization of

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1 the probability of exceeding a particular concentration. Time-variable models that generate long-  
2 term simulations (e.g., 20 years) can also be statistically analyzed to evaluate the exceedence  
3 frequencies. Several statistical models have been developed specifically for conducting these  
4 types of analyses. The Probabilistic Dilution Model (PDM) (Di Toro, 1984) is one such model.  
5 Program Monte (HydroQual, 1997) is a Monte Carlo program that generates a time series of  
6 daily concentrations. The Dynamic Toxics (DYNTOX) model (LTI, 1994) is an EPA model that  
7 uses three statistical methods to predict the frequency of water quality standard violations.  
8 RIVRISK, one of the steady-state analytical solution models described above, also contains a  
9 built-in Monte Carlo simulator.

10 Transport and fate models will vary considerably in their ease of use, with the simpler  
11 one-dimensional and analytical solution models placing relatively modest demands on the user.  
12 Although it may be a simple matter to set up and run this type of model, their proper use requires  
13 the user to exercise a considerable amount of judgment. More complex models become  
14 increasingly difficult to use and interpret, and errors associated with setting up the model are not  
15 always easy to detect. Hence, it is important that the relatively sophisticated modeling analyses  
16 be performed by an experienced analyst, one who is familiar with the details of the processes  
17 included in the model being used. Owing to the complexity of setting up and parameterizing the  
18 more complex models listed in Table 4-13, particularly for analysis of a complex problem  
19 setting, their use will often be limited to relatively refined definitive assessments.

20 Further detailed discussions about features, limitations, and example applications of the  
21 models discussed in this section are provided by Paquin et al. (2003). This review also identifies  
22 additional models designed for use in specialized regions of water bodies, such as mixing zones  
23 or plumes. The appendix includes information about where to obtain many of models discussed  
24

25 **4.4.1.1.2. Limitations.** The partition coefficient, which controls the distribution of metal  
26 between the dissolved and particulate phases, is considered to be a key model parameter in  
27 chemical/metal fate and transport evaluations (see Section 4.1.4). It is important for several  
28 reasons. First, the distribution of metal between the dissolved and particulate phases has a direct  
29 bearing on the magnitude of particulate fluxes of metal that occur in association with the settling  
30 and resuspension of sorbed metal. Second, it also controls the magnitude of diffusive fluxes of  
31 metal between the sediment interstitial water and the overlying water column, as this flux is  
32 proportional to the concentration gradient of total dissolved metal (free ionic metal + metal-DOC  
33 and metal-inorganic ligand complexes) between these compartments. Partitioning reactions also

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1 affect the metal bioavailability and the route of exposure to a metal (food vs. water). Given its  
2 importance, it is necessary to recognize that the assumption of equilibrium partitioning is a  
3 potential oversimplification in some situations (e.g., near a point source discharge to a receiving  
4 water, or immediately following a pulse exposure of such as an overflow from a storm sewer  
5 system). Under these conditions, the assumption of equilibrium between the dissolved and  
6 particulate phases may be less appropriate than is otherwise the case and additional uncertainty  
7 will necessarily be associated with the model results. If this situation exists, consideration  
8 should be given to conducting special studies (e.g., measurement of free metal concentrations  
9 over time in a water sample) to test the validity of the equilibrium assumption.

10 For neutral organic chemicals, the magnitude of the partition coefficient is often assumed  
11 to be proportional to the dissolved organic carbon content of the particles. Use of linear partition  
12 coefficients is likely to be an oversimplification in the case of metals, where a variety of sorption  
13 phases may be important. Several models are available that provide a relatively detailed  
14 representation of these metal-particle interactions (see Paquin et al., 2003, for a review).

15 Many models that were initially developed for organic substances assume first-order  
16 decay processes. These are of little importance for metals, but they sometimes are used in  
17 screening-level assessments to represent removal of sorbed material from the water column.  
18 This approach provides a relatively simplistic representation of the underlying processes and  
19 should be used with caution. Given the capacity of most computing environments today, the  
20 potential exists to use relatively sophisticated metal-specific sorption and water-sediment flux  
21 models, even for simple assessments. However, until such subroutines become more widely  
22 available, their use will be problematic, especially for an inexperienced user. Modification of  
23 existing models may be an option, but this should be attempted only by an individual who is  
24 experienced in model development and programming.

25 Modeling of mercury and metalloids, such as arsenic and selenium, is complicated by  
26 transformation processes that change the form of the metal. Methylation of mercury and arsenic  
27 or binding of selenium to amino acids (e.g., selenomethioine) changes both their physical and  
28 biological properties (see Section 4.1. for a discussion of transformation processes). Similarly,  
29 metals that readily change oxidation state (e.g., chromium) also require additional considerations  
30 in fate modeling. Many of the same transport models can be used, but input parameters will  
31 require modification. Such models are beyond the scope of this framework, which is focused on  
32 inorganic forms of metals. See, for example, the Mercury Cycling model by Hudson et al.  
33 (1994) for further guidance on fate and transport models for mercury.

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1 All modeling exercises are limited by the validity of the model framework, the accuracy  
2 of input parameters, and the experience of the analyst (Dzombak and Ali, 1993). All exposure  
3 assessments should include an explicit description of model assumptions and associated  
4 uncertainties.

#### 6 **4.4.1.2. Terrestrial**

7 Movement of metals through soils is dependent on the chemical properties controlling  
8 speciation, the presence of ligands that control complexation of metals within pore water (and  
9 ground water) and adsorption onto mineral surfaces, and the rate of water flux through the soil.  
10 Metals are lost from the soil primarily by leaching into ground water, although in particular  
11 instances uptake by plants can represent a significant loss. Section 4.1.4 reviews the processes  
12 and models that predict movement of chemicals through soils or partitioning onto mineral  
13 surfaces (i.e., partition coefficients). These serve the same role as the fate and transport models  
14 discussed above for aquatic systems and therefore are not repeated here.

#### 16 **4.4.2. Routes of Exposure to Aquatic and Terrestrial Species**

##### 17 **4.4.2.1. Aquatic Species**

18 Potential exposure routes for aquatic species include inhalation/respiration, dermal  
19 absorption, and diet (either food or incidental sediment ingestion). The extent to which a metal  
20 is taken up by any one of these exposure routes is difficult to define for all relevant routes.  
21 Inhalation/respiration, which for aquatic species means general exchange across respiratory  
22 surfaces, can involve diverse gill and lung types. Respiratory surfaces include fish gills, various  
23 molluscan and arthropod gill types, the pseudo-lungs of pulmonate gastropods, cells surrounding  
24 the sponge spongocoel, mammalian and avian lungs, and plant leaves. Absorption includes  
25 movement across the skin; the walls of such diverse structures as spongocoels, the cnidarian  
26 gastrovascular cavity, and the echinoderm water-vascular system; the filtration-based feeding  
27 structures of many crustacea, insects, and polychaetes; diverse phytoplankton cell membranes;  
28 and plant roots and leaves. Ingestion pathways can involve direct consumption or consumption  
29 after exchange through a trophic web. The diversity of potential receptors makes definition of  
30 exposure pathways more difficult for aquatic receptors than for humans.

31 The respiration/inhalation route is a particular challenge in aquatic exposure assessments  
32 because of the differing types of respiratory organs, the dynamic nature of the respiratory  
33 process in water, and the intimate contact between a receptor and metals dissolved in waters.

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1 Further complicating the issue, some respiratory organs can also be involved in locomotion,  
2 excretion, ion regulation, and food capture, sorting, and ingestion. The absorption route can  
3 involve uptake across a phytoplankton cell membrane, amphibian skin, arthropod exoskeleton,  
4 the general integument of an infaunal clam or annelid, or the egg membrane of a bird or  
5 amphibian. Absorption can occur from overlying or interstitial waters; these sources have very  
6 distinct chemistries that influence metal bioavailability.

7 Similarly, the ingestion route is difficult to define for aquatic receptors because of the  
8 diversity of feeding modes and food sources, such as sediments, suspended solids, microflora,  
9 animal tissues, and plant tissues. The combining of respiratory, locomotive, or feeding  
10 structures complicates description of the ingestion pathway for some species. The presence of  
11 life stages that feed differently also confounds exposure assessment.

12 Despite the complexities associated with quantifying exposure of aquatic animals to  
13 metals from multiple routes of uptake, the relative importance of the different uptake pathways  
14 has received considerable attention in recent years (Wang 2002; Hook and Fisher, 2001; Fisher  
15 et al., 1996; Bjerregaard et al., 1985). Bioenergetic-based kinetic models used to describe the  
16 accumulation of contaminants in aquatic animals have been developed relatively recently and  
17 have been successfully applied to a variety of organic and inorganic contaminants. These  
18 models provide a broad framework for addressing controls on contaminant bioaccumulation for  
19 diverse organisms and can be used for studying contaminant bioavailability and determining the  
20 relative importance of different routes of contaminant accumulation, including that of metals  
21 (Landrum et al., 1992; Wang et al., 1996). The models are flexible enough to incorporate  
22 environmental variability in contaminant sources, contaminant concentrations, food availability,  
23 and organism growth rates in their predictions of organism contaminant levels.

24 Applications of one-compartment biokinetic models using laboratory-based  
25 measurements of key model parameters (assimilation efficiency, uptake rates from water and  
26 food, elimination rates) have been extended to field situations for populations of marine mussels  
27 (Wang et al., 1996; Fisher et al., 1996); Ag, Cd, Co, and Se in clams (Griscom et al., 2002;  
28 Luoma et al., 1992); Po in copepods (Stewart and Fisher, 2003; Fisher et al., 2000); Se in fish  
29 (Baines et al., 2002); and freshwater mussels (Roditi et al., 2000). Site-specific model  
30 predictions for metal concentrations in animal tissues are strikingly close to independent field  
31 measurements for diverse water bodies, suggesting that it is possible to account for the major  
32 processes governing contaminant concentrations in aquatic animals and that the laboratory-  
33 derived kinetic parameters are applicable to natural conditions.

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1 Temporal aspects of exposure should be considered in  
2 assessments of any toxicant. Rapid speciation and phase  
3 changes associated with changes in pH/Eh make temporal  
4 issues particularly germane to metals. Fluctuating or pulsed  
5 exposures occur in situations such as rapid changes in pH/Eh  
6 associated with photosynthesis and respiration, hypolimnetic  
7 discharge from stratified reservoirs, biocide (e.g., copper  
8 sulfate) spraying, ingestion of prey items with seasonally  
9 high metal concentrations, surface waters receiving wastewater treatment plant effluent, urban  
10 storm water, snowmelt, and acid precipitation runoff. Transient metal concentrations may be  
11 orders of magnitude higher than background concentrations but may last for only a few hours.  
12 These episodic exposure scenarios have been poorly characterized for metals (Hoang et al.,  
13 2005). Any risk assessment for metals should clearly state all assumptions about duration of  
14 exposure.

**Temporal Aspects of Exposure**  
*Temporal aspects* of exposure are important considerations in assessing aquatic risks to metals. Organisms may respond vary different to episodic exposure than they respond to the constant exposures incorporated into classic toxicity bioassays. Sound risk analyses clearly articulate all assumptions about exposure durations.

15 ***Water pathway and respiratory route.*** For acute exposure of most water column  
16 organisms, binding of metals to the gill (respiratory) surface is the primary route of exposure.  
17 Binding of metals to the gill surface is the primary route of  
18 exposure for most water column organisms, at least for short-  
19 duration exposures. Chemical kinetics play an important role in  
20 this context, as gradients in pH and ionic composition exist at the  
21 gill surface microlayer due to respiration, excretion, and ion  
22 regulation. Dissolved aluminum toxicity to freshwater fish is a  
23 good illustration of this point (Playle and Wood, 1990). The deposition of aluminum on gills is  
24 determined largely by the rapid shift from dissolved ionic aluminum to an Al(OH)<sub>3</sub> precipitate on  
25 the gill surface due to the more alkaline state of gill microlayer water. How much aluminum  
26 speciation and phase association changes occur is a function of the initial pH of the bulk water.  
27 Exposure assessment should address metal speciation kinetics in such situations (see Section 4.1,  
28 Environmental Chemistry) or else inaccuracies will emerge in subsequent effects assessment.  
29  
30

**Primary Route of Exposure**  
*Binding of metals to the gill surface* is the primary route of exposure for most water column organisms for short-duration exposures.

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1 **4.4.2.1.1. Application.** Predicting exposure at the  
2 respiratory surface should include analysis of chemical  
3 speciation, chemical kinetics, and binding with biological  
4 ligands on the gill (or equivalent respiratory surface). The  
5 FIAM states that, in general, metal ion availability and  
6 effect are correlated with free ion concentration or activity  
7 in the water (Brown and Markich, 2000; Campbell, 1995).  
8 Consequently, knowledge of free ion concentration or  
9 activity is crucial to fully describing exposure. The BLM

**Free Ion Activity Model (FIAM)**

The *FIAM* states that, in general, metal ion availability and effect are correlated with the free ion concentration or activity in the water. It also recognizes the role of competing ions in reducing metal bioavailability ( e.g., calcium reducing zinc bioavailability). The FIAM is a powerful tool as long as it is applied with enough understanding to anticipate or recognize exceptions.

10 further suggests that the bioactivity of a metal is a result of its interaction with biological ligands  
11 (i.e., biological macromolecules on the surface of the respiratory organ) (see Section 4.5,  
12 Characterization of Ecological Effects). It follows that a dissolved metal ion's bioactivity is a  
13 function of its complexation with dissolved ligands (which determines how much free ion will be  
14 available for binding with biological ligands) and the affinity and stability of the metal  
15 complexes with the biological ligands. Further complexation of metals takes place within the  
16 cells of the respiratory organ, with the remaining free metal available for binding to transport  
17 macromolecules for delivery to the organism's circulatory system; this results in the true  
18 delivered dose.

19 Equilibrium speciation can be used to estimate the aquatic free ion concentration and its  
20 resulting activity. In some cases, the free ion can be measured directly during an exposure  
21 assessment. Combining insights from the FIAM approach with those from the BLM and HSAB  
22 (see Section 4.1, Environmental Chemistry) theories allows general prediction of metal activity  
23 on biological surfaces as different as fish gills (Janes and Playle, 1995; Reid and McDonald,  
24 1991; Pagenkopf, 1983), green algae (Parent and Campbell, 1994; Crist et al., 1988), and  
25 bacteria (Azenha et al., 1995). Entry across the integument could also be addressed with  
26 conceptual tools of the FIAM and BLM (Krantzberg and Stokes, 1988), but this application is  
27 not yet sufficiently developed for immediate use.

28 In the absence of sufficient information about speciation, exposure concentrations for  
29 dissolved metals such as cadmium, lead, and zinc can be normalized on the basis of water  
30 hardness. Most practitioners fit linear models (log of toxicity endpoint = log a + b [log of  
31 hardness] and back-transform them to produce a normalizing function:

$$\text{Toxic endpoint} = 10^a (\text{hardness})^b$$

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1 This power function contains an easily corrected back-transformation bias that should be  
2 removed from predicted hardness-adjusted exposure concentrations (Newman, 1991). The use  
3 of hardness as a normalizing function should be done only when information on speciation is  
4 lacking, as there is greater uncertainty using this method.

5  
6 **4.4.2.1.2. Limitations.** The FIAM is a powerful tool as long as it is applied with enough  
7 understanding to anticipate or recognize exceptions. As examples of important exceptions,  
8 charged uranium complexes are toxic, in addition to the free ion (Markich et al., 2000), and the  
9 neutral mercury complex  $\text{HgCl}_2^0$  is bioavailable due to its high lipophilicity (Simkiss, 1996).  
10 Small organic ligands bind metals forming nonionic complexes that are also exceptions to the  
11 FIAM. Copper bound to ascorbate was bioavailable to the green alga, *Scenedesmus sp.*  
12 (Campbell, 1995). Silver bound to glutathione or cysteine was bioavailable to *Ceriodaphnia*  
13 *dubia* (Bielmyer et al., 2002). In addition, some nonionic metal-inorganic ligand complexes,  
14 such as  $\text{NiCO}_3$ , may also be bioavailable (Hoang et al., 2004).

15 The BLM has been parameterized for copper, nickel, silver, and zinc in fish, algae, and  
16 invertebrates (*Daphnia*, *ceriodaphnia*). Some initial work has been completed for other species  
17 and metals, but rigorous parameterization/validation is limited to the organisms and metals listed  
18 above. Recent efforts in Europe have focused on extending BLM principles to describe chronic  
19 toxicity; the results should become available in the near future. At present, the BLM approach  
20 assumes that delivered dose is equivalent to the amount of metal bound to the gill (or other  
21 respiratory organ), which may overestimate exposure. Models needed to predict within-cell  
22 complexation of metals into available and nonavailable pools have not yet been developed.

#### 23 24 **4.4.2.2. Food Chain Pathway and Dietary Exposure**

25 Defining the particulars of metal exposure by ingestion is complicated by the diversity of  
26 feeding modes, digestive systems, and physiology of candidate receptors. That dietborne  
27 exposure to metals can result in accumulation in aquatic  
28 organisms is well established, although the rate and  
29 magnitude vary among organisms. What is less well  
30 understood is how best to express dietary exposure in a  
31 way that can be linked to potentially toxic effects. For  
32 nonionic organic chemicals, evidence is strong that  
33 whole-body burdens of chemical (normalized to lipid

Currently, <i>no</i> standard approaches exist to assess dietborne exposures of metals to aquatic organisms in water quality assessments, and consequently, the most significant limitation involves a lack of broad understanding of the mechanisms underlying dietborne metal toxicity.
---

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1 content) are robust metrics of toxicological dose, and these relationships appear to be  
2 independent of whether exposure was via water or diet. In fact, the greatest strength of the body  
3 residue approach for organic chemicals is that it effectively integrates different exposure  
4 pathways into a single expression of dose and toxicological potency.

5 For metals (aside from organo-selenium and methyl mercury), the situation is far more  
6 complex, and whole-body residue does not appear to be a robust indicator of dose when  
7 compared across a range of exposure scenarios and/or organisms. The reason that whole-body  
8 residue/effect relationships are not as effective for assessing metals probably stems from the fact  
9 that although distribution of nonpolar organic chemicals in organisms is influenced largely by  
10 passive partitioning, the uptake, distribution, and disposition of metals is governed by a number  
11 of active biochemical processes. For example, some organisms take up metal and sequester it  
12 into “storage” compartments in chemical forms that have little toxicological potency, whereas  
13 other organisms actively excrete excess metals. Even for a particular organism, uptake and  
14 disposition of metal may vary between waterborne and dietborne exposure (e.g., Kamunde et al.,  
15 2002; Szebedinszky et al., 2001).

16 Although these issues confound the development  
17 of simple dose/effect metrics based on whole-body metal  
18 residues, one should presume that residue/effect  
19 relationships could be established if there were better  
20 understanding of more specific concentration/response  
21 relationships for the site (or tissue) of toxic action. For  
22 example, some studies have suggested that the metal  
23 concentration in the cellular cytosol (as opposed to that bound to cell walls or sequestered in  
24 nonbioavailable metal granules) may provide a better expression of internal metal dose  
25 associated with toxic effects (Wallace and Luoma, 2003; Wallace et al., 2003; Wallace and  
26 Lopez, 1996). Others have suggested that whole-body residue residue-effect relationships are  
27 confounded because the factor that determines the effects is not whole-body concentration per  
28 se, but the rate of metal uptake in relation to metabolic capacity for detoxification and storage,  
29 and, therefore, effects are governed by factors that influence the rate of uptake. When uptake is  
30 elevated, the concentration of metabolically active metal at the site(s) of action increases (e.g.,  
31 the spillover hypothesis) and effects ensue (Rainbow, 2002).

Assessment of dietborne metal exposure is best considered in two contexts: (1) dietborne exposure leading to accumulation and exposure to higher levels in the food chain (e.g., humans, wildlife) and (2) dietborne exposure leading to direct effects on exposed organisms.
---

32 For these reasons, assessment of dietborne metal exposure should be considered in two  
33 contexts: (1) dietborne exposure leading to accumulation and exposure to higher trophic levels

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1 in the food chain (e.g., humans, wildlife) and (2) dietborne exposure leading to direct effects on  
2 exposed organisms. For the former, the primary challenges are to predict the aggregate uptake of  
3 metal from both waterborne and dietborne exposure and to express that accumulation in terms  
4 that are appropriate to assessing risk to the consuming organisms. Issues that come into play in  
5 this scenario include whether forms of metal that are sequestered and detoxified by the aquatic  
6 organism are bioavailable to organisms that then consume those aquatic organisms. The  
7 available data to date suggest that metals can be sequestered/detoxified via different  
8 mechanisms/forms in an organism, with metals sequestered as inorganic granules having greatly  
9 reduced to no bioavailability to consumer organisms, while metals detoxified via  
10 metallothioneins have relatively higher bioavailability to consumer organisms (Wallace et al.,  
11 2003; Wallace et al., 1998; Mason and Jenkins, 1995; Nott and Nicolaidu, 1994). However,  
12 owing to different digestive physiologies and other factors, changes in the bioavailability of  
13 metals sequestered/detoxified by these different mechanisms or forms may occur (Wang, 2002).  
14 With respect to the second scenario, the literature is mixed on the degree to which this is an  
15 important pathway for inducing toxicity to aquatic organisms and how best to quantify exposure  
16 in that context.

17  
18 **4.4.2.2.1. Application.** There currently are no standard approaches to assess dietborne  
19 exposures of metals to aquatic organisms in water quality assessments (Schlekat et al., 2001).  
20 For the organometallic compounds of organo-selenium and methyl mercury, dietborne exposure  
21 has been clearly shown to be a primary route of both uptake and toxic effects and should be  
22 considered in assessments of these metal compounds. Beyond those two compounds, the picture  
23 is much less clear. Toxicity to aquatic organisms from dietborne exposure to metals has been  
24 demonstrated where exposure is sufficiently high, although in some cases these concentrations  
25 are extreme (e.g., 10,000  $\mu\text{g/g}$  Cu) (Handy et al., 1993). If the dietborne exposure necessary to  
26 elicit effects is exceptionally high, it is not clear that this pathway will drive ecological risk, as  
27 the environmental concentrations necessary to produce these exposures may be so extreme that  
28 ecological risk will occur via other pathways (e.g., direct toxicity of waterborne metal).

29 In other studies, however, effects from dietborne exposure have been demonstrated at  
30 relatively low exposure concentrations, such as in zooplankton studies (Hook and Fisher, 2002;  
31 2001a, b). This raises much greater concern for metals assessment because it raises the potential  
32 for toxicologically significant exposures occurring in cases where risk via a waterborne pathway  
33 is low. That potential notwithstanding, other studies evaluating dietborne exposure with the

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1 same organisms and metals but somewhat different methods have reached different conclusions  
2 regarding the significance of dietborne exposure (De Schamphelaere and Janssen, 2004; Metzler,  
3 2003). Dietborne exposure to metals is an area of active research, and it is likely that new  
4 insights will be forthcoming to rationalize what now appear to be conflicting data into a more  
5 comprehensive understanding of dietborne metal effects. Until that time, the absence of a  
6 standard approach will require that decisions regarding the handling of dietborne metal exposure  
7 in aquatic assessments be made on a case-by-case basis. Some bioaccumulation models for  
8 metals include consideration of dietborne exposure, although few, if any, link this accumulation  
9 directly to effects.

10 In the context of screening for exposure and potential risks to consumers, the use of  
11 whole-body inorganic metal concentrations in prey species may have some utility despite the  
12 uncertainties associated with trophic transfer and bioavailability of dietary metals (i.e., in cases  
13 where whole-body residues are below dietary toxic thresholds). For more definitive  
14 assessments, further research is needed on quantifying the bioavailability and effects of inorganic  
15 dietary metals (with the exception of certain organometallics where dietary toxicity has been  
16 well established).

17 In cases where trophic relationships are not well understood, stable isotope techniques  
18 can aid in defining the trophic status of species of concern and can be used to delineate the food  
19 web (i.e., who is feeding on whom). The  $^{15}\text{N}$  concentration increases relative to  $^{14}\text{N}$   
20 concentration with each trophic exchange, and statistical models can link trophic status to metal  
21 concentration (e.g., Cabana et al., 1994). Additionally the potential for using stable (and radio)-  
22 isotopes of a metals can serve as a tools to understand dietary versus waterborne uptake,  
23 particularly in laboratory studies. Other uncertainties include how behavioral changes induced  
24 by dietborne metal exposure may affect the survival of the exposed organism (Irving et al.,  
25 2003).

26  
27 **4.4.2.2.2. Limitations.** As indicated above, the primary limitation in assessing dietborne metal  
28 exposure for aquatic organisms is the absence of a broad understanding of the mechanisms  
29 underlying dietborne metal toxicity and the consequent lack of a standard assessment approach.  
30 Establishing rigorous residue/effect relationships that integrate waterborne and dietborne metal  
31 exposure is a critical consideration. When assessing dietborne exposure through direct  
32 measurement from field samples, there are methodological issues to be resolved, such as whether  
33 to deplete the digestive system of prey organisms. Metal contained in food or sediment within

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1 the digestive system will increase measured body burdens, but it may not have the same  
2 biological availability as metal residing within organismal tissues (Chapman et al., 2003).  
3 Fractionating body burdens of metals (e.g., cytosolic metal vs. metal granules) has been  
4 suggested as a better means of identifying the most readily accumulated fraction of dietborne  
5 metal (Seebaugh and Wallace, 2004; Wallace et al., 2003; Fisher and Reinfelder, 1995;  
6 Reinfelder and Fisher, 1994), although the interpretation of this information in the context of risk  
7 assessment has not been rigorously developed.

#### 8 9 **4.4.2.3. Sediment Exposure**

10 Most situations involving dissolved metals can be addressed adequately by assuming  
11 near-equilibrium conditions between water and sediment concentrations (Hoffmann, 1981), as  
12 long as one is sufficiently aware that some conditions can lead to important nonequilibrium  
13 dynamics (Paquin et al., 2003). Bioavailability, sediment  
14 transport, and chemical speciation all affect exposure of  
15 benthic organisms to metals. Additional issues of trophic  
16 transfer and routes of exposure (gills or equivalent vs.  
17 dietary) should be considered in the same manner as  
18 discussed above for water column organisms.

<p><b>Sediment Exposure Estimates</b></p> <p>Not all benthic organisms are exposed to sediments. Some feed and respire in the overlying water column. Sediment exposure estimates apply only to those organisms that extract nutrients or oxygen from sediments and pore water.</p>
---

19 The most widely used approach for assessing metal  
20 exposure in sediment is based on EqP theory, with sulfides as the primary partitioning phase (Di  
21 Toro et al., 1990; 1991). Other approaches include sequential extractions with different media  
22 (water, weak acids, strong acids) or normalization to total iron or organic carbon. Each has its  
23 strengths and limitations. Detailed discussions are provided in Paquin et al. (2003) and Newman  
24 et al. (2004); additional information is provided in Section 4.5.9.

25  
26 **4.4.2.3.1. Application.** Several approaches have been taken for estimating exposures to  
27 sediment-associated metals that account for bioavailability differences among various sediments.  
28 The EqP approach assumes that chemical activity in the sediment, as indexed by chemical  
29 concentration in the interstitial water, is proportional to the chemical's bioavailability to  
30 sediment-dwelling organisms. In anoxic sediments, sulfides provide the primary binding phase  
31 for many cationic metals. These metal sulfides are highly insoluble and are thought to have very  
32 low toxicity. Thus, in sediments where there is more sulfide than metal, most metal should be  
33 present as sulfides and therefore relatively nontoxic. The amount of reactive sulfide is quantified

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1 by measuring the amount of sulfide liberated when sediment is extracted with 1 N HCl. This  
2 procedurally defined quantity is known as acid-volatile sulfide or (AVS). The amount of  
3 reactive metal is determined from the same extraction by measuring the metal concentration in  
4 the acid extractant. This quantity is known as simultaneously-extracted metal or SEM. The  
5 potential bioavailability of metal is determined by comparing the relative molar concentrations  
6 of the two. When SEM-AVS < 0, sufficient sulfide exists to bind all SEM and metal toxicity is  
7 not expected. When SEM-AVS > 0, metal is present beyond the binding capacity of sulfide, and  
8 toxicity may occur if there is sufficient excess metal but not sufficient other binding phases to  
9 bind the metal. Use of this SEM-AVS as exposure estimates that are correlated with toxicity of  
10 metals in sediment has been explored closely for Class B or borderline Class B metals (Berry et  
11 al., 1996; Hansen et al., 1996; Ankley et al., 1996; Ankley et al., 1991; Carlson et al., 1991; Di  
12 Toro et al., 1990). See Section 4.5.9 for further discussion.

13 Although the correspondence of SEM-AVS to  
14 toxicity was found to be strong in these studies, some  
15 question the applicability of the approach to all benthic  
16 organisms because it is based on the chemistry of bulk  
17 anoxic sediment, and many organisms live in oxygenated  
18 burrows. In addition, several studies have shown some  
19 degree of metal accumulation in organisms exposed to  
20 sediments where sulfide is in excess and metals are thought to be nonbioavailable (or at least  
21 nontoxic). A better understanding of the mechanisms of metal accumulation from sediment and  
22 their relationship to toxic effects is needed to help interpret these issues. Until such information  
23 becomes available, the SEM-AVS model can be used in exposure estimations as long as its  
24 shortcomings are acknowledged appropriately.

25 Other tools for determining the exposure  
26 concentration of sediment-bound metals include  
27 metal concentrations in chemical (Fan and Wang,  
28 2001; Babukutty and Chacko, 1995; Tessier et al.,  
29 1984), acid (Langston, 1980; Luoma and Bryan,  
30 1978), or biomimetic (Weston and Maruya, 2002;  
31 Mayer et al., 2001; Chen and Mayer, 1998) extracts.  
32 However, no consensus yet exists on their best use  
33 for different types of metals or metalloids. Several

#### **Sem-AVS**

When the molar concentrations of *acid-volatile sulfide (AVS)* in sediment exceed the amount of *simultaneously extracted metal (SEM)*, the metals are expected to associate with the solid phase and not be bioavailable.

#### **Exposure Assessments for Benthic Organisms**

The following information would enhance exposure assessments for benthic organisms: (1) improved computational or analytical methods for analyzing distribution of metal among components of the sediments, (2) improved computational methods for assessing the influences of metal form in sediments on sediment-water metal exchange, and (3) a better understanding of the processes controlling bioaccumulation of metals from solution and food by metazoan species directly exposed to the sediments.

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1 other methods have been proposed. Based on the premise that iron oxides in oxic sediments  
2 lower metal bioavailability, iron in a 1 N HCl sediment extract has been used to normalize metal  
3 exposure concentrations (Luoma and Bryan, 1978). Increasing concentrations of organic carbon  
4 can decrease metal bioavailability (Crecelus et al., 1982), so normalization of sediment metal  
5 concentrations to organic carbon content has been useful in other cases. The more readily  
6 extracted metals from sequential chemical extraction schemes tend to be the most bioavailable  
7 (Young and Harvey, 1991; Tessier et al., 1984) and can be used as exposure metal  
8 concentrations.

9  
10 **4.4.2.3.2. Limitations.** Exposure assessment for benthic receptors could be enhanced if a clearer  
11 consensus were reached about the utility of each method for different classes of metals,  
12 biological species, and sediment types. Specifically, the following information is needed to  
13 improve exposure analysis: (1) improved computational or analytical methods for analyzing  
14 distribution of metal among components of the sediments, (2) improved computational methods  
15 for assessing the influences of metal form in sediments on sediment-water metal exchange, and  
16 (3) a better understanding of the processes controlling bioaccumulation of metals from solution  
17 and food by metazoan species directly exposed to the sediments (Luoma, 1989).

18 EPA (U.S. EPA, 2002f) published a report on the application of solid phase AVS  
19 equilibrium partitioning sediment benchmarks (ESBs) and interstitial water ESBs as No-Effect  
20 guidelines to predict sediments that are acceptable for the protection of benthic organisms.  
21 Details of the SEM-AVS method are discussed in Section 4.5.9 and the metals issue papers. The  
22 method has been applied to predict toxicity of metals in sediment for Class B or borderline Class  
23 B metals. An ESB based on the difference between the concentration of SEM and AVS is  
24 appropriate for protecting benthic organisms from the direct effects of sediment-associated  
25 metals, and not for estimating metal bioaccumulation. Chapman (2003) discusses limitations  
26 concerning the AVS-SEM approach, including the degree to which the EqP approach adequately  
27 represents exposures for organisms living in microenvironments in the sediment and/or who are  
28 exposed via ingestion of sediment. Site-specific measurements, where possible, are useful for  
29 confirming that generalized approaches like SEM-AVS appropriate for specific assessment  
30 scenarios. Furthermore, one should recognize that bioaccumulation of metal may still occur  
31 even when sulfides are in excess, and the potential consequences of this accumulation should be  
32 considered.

33  
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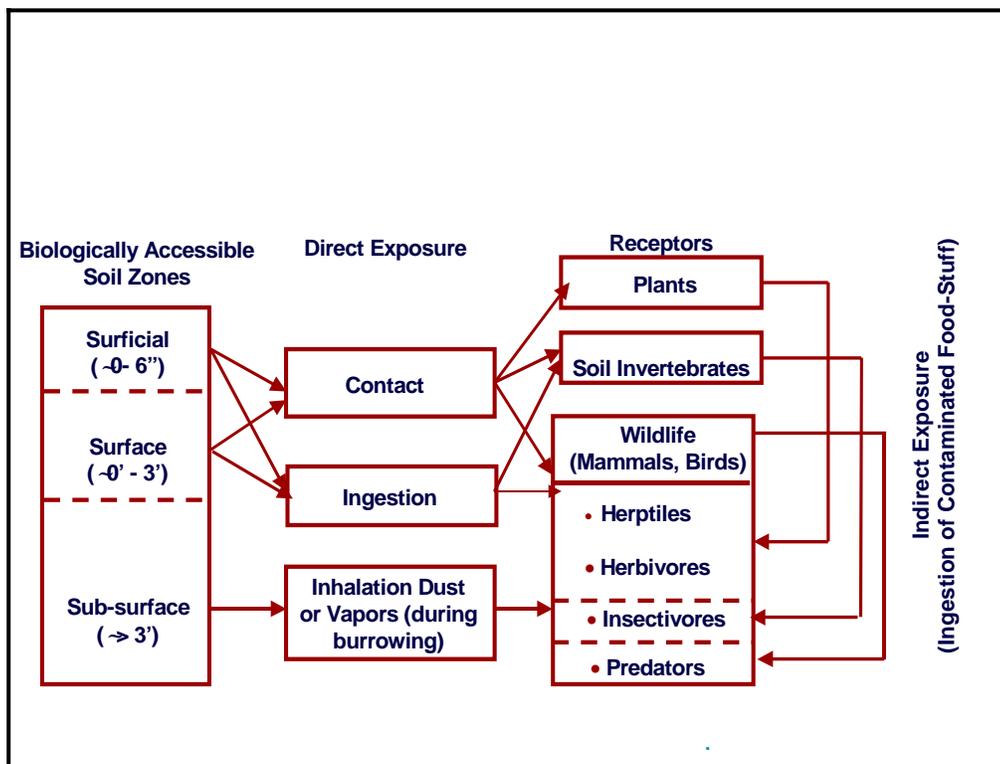
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1 **4.4.2.4. Terrestrial Species**

2 Terrestrial wildlife, plants, and invertebrates  
3 accumulate metals from direct contact with soil or sediment,  
4 from ingestion of contaminated food (plants or other animals),  
5 and from incidental soil or sediment ingestion. A conceptual  
6 model for direct and indirect exposure of terrestrial receptors  
7 to metals in soil is presented in Figure 4-6. Pathways of  
8 exposure include movement from soils through the food web,  
9 and to a lesser extent, air deposition either into soils or directly  
10 onto terrestrial receptors (e.g., plants). Because of significant differences in exposure patterns, it  
11 is more convenient to discuss methods by receptor group (invertebrates, plants, wildlife) rather  
12 than by pathways or environmental compartments.

<p><b>Pathway of Exposure for Terrestrial Organisms</b></p> <p><i>Pathways of exposure for terrestrial organisms</i> to metals include movement from soils through the food web, and to a lesser extent, air deposition either into soils or directly onto terrestrial receptors (e.g., plants).</p>
--

13  
14 **4.4.2.4.1. Soil invertebrates.** The soil ecosystem includes a complex food web of soil  
15 invertebrates (both hard- and soft-bodied) that feed on each other, decaying plant material, and  
16 bacteria or fungi. For risk assessment purposes, however, exposure is described as a function of  
17 soil concentration rather than a detailed analysis of movement of metals through the food web.  
18 This is a reasonable approximation for soft-bodied invertebrates (e.g., earthworms) whose  
19 exposure is primarily through soil pore water (from both dermal absorption and soil ingestion)  
20 (Allen, 2002). There is more uncertainty in correlating soil concentrations with effects in hard-  
21 bodied invertebrates because they are primarily exposed through ingestion of food and incidental  
22 amounts of soil. (Sample and Arenal, 2001). Regardless, for all types of soil invertebrates,  
23 exposure estimates should account for differences in bioavailability among soil types, which  
24 include differential partitioning to soil particles and pore water, metal speciation, and aging.  
25 Each of these processes is discussed in detail in Section 4.1, Environmental Chemistry.



**Figure 4-6. Conceptual model for direct and indirect exposure of ecological receptors to metals in soil zones.**

Source: Menzie and Little (2000).

1 **4.4.2.4.1.1. Application.** Currently, soil invertebrate exposure is calculated on the basis of total  
 2 metal concentration in bulk soils collected in the top 0–12 cm of soil (U.S. EPA, 1989c). In  
 3 detailed, higher level assessments, the organic matter on top of the soil (the “duff”) may be  
 4 analyzed separately to provide further detail on exposure to detritivores (such as *Collembola*)  
 5 and deeper-soil-dwelling organisms (e.g., various species of earthworms). However, such  
 6 measures of exposure are limited, as they do not account for differences among soils in  
 7 bioavailability factors.

8 Cation exchange capacity (CEC) recently has been shown to be an important factor  
 9 modifying zinc bioavailability in soils for both invertebrates and plants, and presumably it will  
 10 be important for other cationic metals as well. However, CEC is strongly dependent on the type  
 11 and amount of organic material and oxyhydroxides present in the soil and is strongly pH

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1 dependent. Surface charge on organic material and oxyhydroxides increases with increasing pH,  
2 thereby increasing their sorptive capacity for metals (thus decreasing metal bioavailability).  
3 Conversely, positive surface charges increase as the pH drops, increasing sorption of anions  
4 (e.g., arsenic or selenium) under low pH conditions and decreasing sorption of cation ionic  
5 metals. Clays, on the other hand (except for kaolinite), have a surface charge that is largely  
6 independent of pH. Therefore, normalization of toxicity data to CEC can be done only within  
7 specific soil types and pH ranges, which frequently are not specified either in laboratory  
8 bioassays or many field studies. Furthermore, it is important to note that most published values  
9 of CEC are measured at pH 7.

10 Soil chemical models are being developed to predict how aging will modify bulk soil  
11 concentrations when soils are amended with soluble salts. Aging reduces the bioavailable  
12 fraction of metals over time (see Section 4.1.6.3 for a discussion of aging in soils). Preliminary  
13 studies suggest that consideration of aging may result in estimates of the bioavailable fraction as  
14 low as  $0.1 \times$  bulk soil concentrations. Until the data become available for metals of concern,  
15 toxicity values derived from soluble-salt amended soils (which have not simulated aging) cannot  
16 be reliably corrected to approximate aged metals in field situations. Appropriate adjustments  
17 should be included in toxicity test protocols to simulate aging (McLaughlin et al., 2002), except  
18 when assessing acute (short-term) risks of spills.

19  
20 **4.4.2.4.1.2. *Limitations.*** Data on CEC for field soils are often available, but similar information  
21 from laboratory studies of the toxicity of metal-spiked soil currently is not. Furthermore, the  
22 dependency of CEC on soil type (amount and type of organic matter, type of clay, and pH) also  
23 complicates the comparison among studies. Therefore, although exposure concentrations can be  
24 adjusted across field locations of similar soil type and pH, it is more difficult to make  
25 appropriate comparisons of field exposures with laboratory-generated concentration-response  
26 functions. Expressing exposure on the basis of pore water concentration is the goal, to reduce  
27 the variability in toxicity among sites; however, there are currently significant limitations to  
28 collecting and interpreting metal-related data from soil pore waters. Such information is not  
29 available in the published literature and, therefore, should be estimated using EqP theory (as with  
30 sediment pore water analyses). Published soil binding coefficients ( $K_d$ s) can be used, although  
31 these values are inherently uncertain as well (published value depends on derivation method, soil  
32 type, etc.; see Section 4.1.4 for a discussion on the limitations of  $K_d$ s).

33  
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1 **4.4.2.4.2. *Plants.*** Plants access metals through the pore water,  
2 although mycorrhizae, protons, and phytosiderophores released by  
3 the root can significantly influence the microenvironment and  
4 change uptake rates of metals (George et al., 1994; Sharma et al.,  
5 1994; Laurie and Manthey, 1994; Arnold and Kapustka, 1993).

<p><b>Metal Accumulation in Plants</b></p> <p>The highest accumulation of metals in plants generally occurs in the roots.</p>
---

6 Furthermore, plants have both active and passive mechanisms for taking up or excluding metals,  
7 depending on internal concentrations and whether or not the metal is an essential micronutrient,  
8 or whether it is mistaken for an essential micronutrient. Plants can be exposed to metals via  
9 aerial deposition onto leaf surfaces, trapping metals in hairs or rough cuticular surfaces. This  
10 might provide an exposure route for herbivores; it may also provide an exposure route for plants,  
11 as there are ion channels through the cuticle that are able to transport ionic metals from the leaf  
12 surface to other locations in the plant, depending on the inherent mobility of the metal in the  
13 xylem and phloem (Marschner, 1995).

14  
15 **4.4.2.4.2.1. *Application.*** All plant species take up metals from soil through their roots via  
16 various mechanisms (Raskin et al., 1994; Cataldo and Wildung, 1978). The default approach to  
17 estimating exposure of plants to metal is measuring metal concentrations in bulk soil. However,  
18 as with soil invertebrates, this overestimates exposure because it does not account for differential  
19 bioavailability that results from complexation. Furthermore, with time, the bioavailability of soil  
20 metals may change due to dissolution or complexation; thus, “point-in-time” measurements of a  
21 soil may not reflect the future bioavailability. For example, Pb sulfide spilled onto soil (relative  
22 bioavailability 1–5%) with time would weather to Pb sulfate (relative bioavailability 50%),  
23 which could further evolve to sorb onto Fe oxides or phosphates (relative bioavailability 10–15,  
24 30–59%, respectively). Soil pH, organic matter, and cation exchange capacity are the most  
25 important variables influencing bioavailability (see Section 4.5, Characterization of Ecological  
26 Effects). However, CEC and clay content are not consistently reported in the literature and  
27 therefore cannot be used to define relative bioaccessability and toxicity of metals. General  
28 categories of uptake based on soil pH and organic matter are shown in Tables 4-17 and 4-18 in  
29 Section 4.5 for ranges typically found in soils. These tables are a useful qualitative guide to  
30 identify soils with increased (or decreased) metal bioavailability. It is very clear that strongly  
31 acidic soils increase plant uptake of Zn, Cd, Ni, Mn, and Co and increase the potential for  
32 phytotoxicity from Cu, Zn, and Ni. Alkaline soil pH increases uptake of Mo and Se, while Pb  
33 and Cr are not absorbed to any significant extent at any pH (Chaney and Ryan, 1993).

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1  
2 **4.4.2.4.2.2. *Limitations.*** Qualitative relationships between soil chemistry and bioavailability  
3 are appropriate for national-scale application. However, for site-specific or metals-specific  
4 applications, quantitative methods are preferred. It has been suggested that only uncomplexed,  
5 free ionic species of cations can be taken up by roots, and this has been described using a FIAM  
6 similar to the model used in aquatic systems (Parker and Pedler, 1997; Lund, 1990). Studies  
7 have been conducted to generate models to predict free copper activity from total metal content,  
8 pH, and organic matter content (Sauve et al., 1997a, b, 1995; McBride et al., 1997), and for lead,  
9 empirical models were generated using only total metal levels and pH. However, exceptions to  
10 the free-ion model have been identified. Ionic or organometallic complexes that increase the  
11 total concentration of elements at the root surface have been correlated with increased uptake,  
12 either through disassociated ions or through uptake of intact complexes (Parker et al., 2001;  
13 McLaughlin et al., 1994). In addition, it is not clear how well plants can distinguish between  
14 ions of similar size and charge. Plant uptake of macronutrients is much better understood than is  
15 uptake of micronutrients or contaminants, with the primary work on uptake of micronutrients  
16 focusing on iron (Welch, 1995). Different mechanisms have been identified that control  
17 macronutrient uptake by plants, providing a means through which contaminants can enter root  
18 tissue.

19  
20 **4.4.2.4.3. *Wildlife.*** The relative importance of exposure pathways  
21 and routes varies by species of animal as well as by metal,  
22 although, in general, wildlife exposure is primarily through diet  
23 and incidental ingestion of soils or sediments. The EPA has  
24 concluded that there are certain chemicals and exposure situations  
25 for which inhalation or dermal pathways are important, but under  
26 most situations they can safely be considered to be insignificant contributors to total metal loads  
27 (U.S. EPA, 2003c).

<p><b>Exposure Pathway for Terrestrial Wildlife</b></p> <p>Food and the incidental ingestion of soil are the two most important exposure pathways for terrestrial wildlife.</p>
---

28 Wildlife food chain exposures for metals are controlled by bioavailability,  
29 bioaccessibility, and bioaccumulation. The availability of metals in soils depends on whether  
30 exposure occurs via pore water or other pathways external to the organisms. Bioaccessibility of  
31 metals to animals and plants that live on or in the soils can be influenced by a number of soil  
32 parameters, such as pH, CEC, and organic carbon. These soil factors tend to be less important  
33 for soils that are incidentally ingested by wildlife species. For further review of soil

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1 bioavailability factors that are important in wildlife exposure, see Section 3.4.6.3, Ecological  
2 Effects (Wildlife).

3  
4 **4.4.2.4.3.1. *Application.*** In the absence of site-specific information, the following  
5 generalizations can be used:

- 6  
7 • Incidental soil ingestion is a proportionally more important pathway for herbivores  
8 than for carnivores or invertivores.
- 9  
10 • Uptake into soil invertebrates (e.g., earthworms) is a proportionally more important  
11 pathway for animals that feed on these organisms. (Note: This assessment reflects  
12 work done with earthworms and may not apply to hard-bodied soil invertebrates such  
13 as *Colembolla*.)

14  
15 The relative importance of exposure pathways (soil vs. food chain) is dictated by the  
16 fraction of metal-contaminated soil in the diet and the amount of accumulation of metal in food  
17 items. Figure 4-7 provides a simple scheme for judging the relative contribution of food and soil  
18 before accounting for bioavailability. Incidental ingestion of soil becomes proportionally more  
19 important for exposure to wildlife when (1) the bioaccumulation factor (BAF) from soil to food  
20 (e.g., to plants or soil invertebrates) is less than 1 and (2) the fraction of soil in the diet is greater  
21 than 1%.

22  
23 **4.4.2.4.3.2. *Limitations.*** Experience at metals-contaminated sites indicates that the above  
24 generalizations should be viewed with caution. As site-specific information is acquired, the  
25 relative importance of pathways may change. For example, site-specific data may show that the  
26 accumulation of a chemical into plants or soil invertebrates is much lower than indicated by the  
27 default assumptions. In such cases, incidental ingestion of soil would become proportionally  
28 more important. The bioavailability of metals in incidentally ingested soil is also variable, as  
29 discussed later. Therefore, when the exposure is being driven by incidental ingestion,  
30 refinements of exposure estimates can benefit from a better understanding of bioavailability.  
31 Attention should be paid to the bioavailability of metals for which incidental soil ingestion is the  
32 predominant pathway and where ecological risk is indicated, although very little information is  
33 available on this for most wildlife species.

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Both the bioaccumulation factor and life history components interact to control the relative importance of soil or food as the primary metal transfer medium. In general, the following statements can be made about dietary uptake of metals from soil versus from food:

- If bioaccumulation is low ( $\ll 1$ ), importance of soil ingestion versus diet for metal exposure increases.
- When bioaccumulation is greater ( $\sim 1$  or higher), the food pathway should dominate.
- The closer the association an animal has to the ground, the greater the importance of soil ingestion. This association may be due to ground foraging, burrowing habits, etc.
- The looser the association with the ground (e.g., piscivores, aerial/arboreal insectivores, raptors), the lower the importance of soil ingestion.

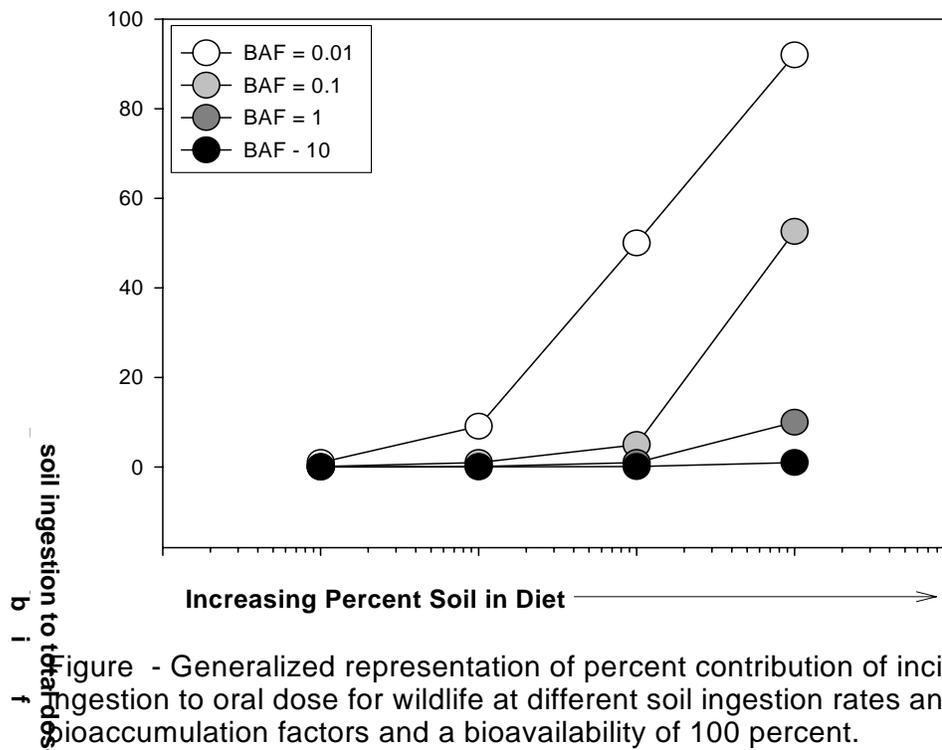


Figure - Generalized representation of percent contribution of incidental soil ingestion to oral dose for wildlife at different soil ingestion rates and bioaccumulation factors and a bioavailability of 100 percent.

Figure 4-7. Relative contribution of incidental soil ingestion to oral dose for wildlife at different soil ingestion rates and bioaccumulation factors, and a bioavailability of 100%.

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1 Extrapolation from models developed for estimation of bioavailability of metals in soils  
2 for incidental human exposures may not be broadly applicable to all wildlife species owing to  
3 the influence of differences in digestive physiology and anatomy across the broad and diverse  
4 range  
5 of mammalian and avian species (Menzie-Cura and TN&A, 2000). For example, metals present  
6 in soils may be more or less bioavailable within the gut of an herbivore that relies on  
7 fermentation as compared to the simpler gut of a carnivore that is designed to break down  
8 proteins. These gut systems differ in chemistry (including pH) and residence time. For example,  
9 ruminants such as deer, antelope, and other hoofed stock initially process food through microbial  
10 digestion in the rumen, and their gut pH is general neutral or slightly acidic. Furthermore, end  
11 products of microbial digestion in domestic sheep bind to copper and enhance its uptake in the  
12 proximal intestine, resulting in a very low tolerance of these animals to dietary copper  
13 (NAS/NRC, 1980). Hind gut fermentors such as horses, rabbits, and granivorous birds (e.g.,  
14 grouse or pheasants) also rely on microbial processes for digestion of lignen and other plant parts  
15 but have a more acidic foregut than do ruminants. Most insectivores and carnivores, on the other  
16 hand, have relatively acidic digestive systems with a significant amount of protein present.  
17 While the low pH may alter the metal speciation (or dissociate ions from attached ligands), most  
18 metals require active transport to move through the gut wall and into the circulation. This may  
19 be accomplished through binding with transporter proteins present either in the diet or within  
20 gastrointestinal cells, which likely differ among the wildlife species (Hill, 1979). Taken  
21 together, these physiological differences are significant and make it difficult to accurately  
22 extrapolate dietary toxicity thresholds across species. However, thoughtful application of this  
23 information will allow appropriate inferences to be made across species with similar physiology  
24 and may explain instances where statistical projections of toxic thresholds (e.g., species  
25 sensitivity distributions) appear to not be predictive of actual effects.  
26

#### 27 **4.4.3. Food Chain Modeling for Wildlife**

##### 28 **4.4.3.1. Application**

29 Food chain modeling can be used to estimate the exposure of wildlife to metals based on  
30 ingestion of soil, food, and water. The basic format of the model is the same as that for organic  
31 substances and is shown in Figure 4-8; detailed explanations are available in several related  
32 documents (e.g., U.S. EPA, 2003c [EcoSSLs]; U.S. EPA, 1997d [ECOFRAM]; Sample et al.,  
33 1997). Measured, or predicted/estimated, concentrations of metals in soil, surface water, and

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1 food items can be used in the model, or concentrations in food can be modeled using trophic  
2 transfer factors. TRIM.FaTE ([http://www.epa.gov/ttn/fera/human\\_apex.html](http://www.epa.gov/ttn/fera/human_apex.html)) can be used for  
3 modeling media concentrations of metals as a result of aerial deposition. Information on diet,  
4 foraging area, and the like can be found in CH2M HILL (2001) and Sample et al. (1997). The  
5 absorbed fraction variable accounts for differences in relative bioavailability (RBA), and is  
6 either 1 (default value) or an appropriate estimate. Note that there is no relative absorption  
7 factor (RAF) included for food, because the default assumption is that site bioavailability equals  
8 that from the toxic form used in the toxicity tests

9 This approach is the same as that used in risk  
10 assessments of organic substances except when trophic  
11 transfer rates are used to model food concentrations only  
12 on the basis of soil measurements (rather than using direct  
13 measures of concentration of metals in food items), in

14 which case metal-specific transfer rates are required. As with aquatic organisms, trophic transfer  
15 values for metals in terrestrial systems are an inverse function of soil concentrations. Therefore,  
16 it is inappropriate to use constants for this term. Sources, use, and limitations of function of soil  
17 where apparent uptake ratios are greater at the lowest and highest concentrations of metals in  
18 soils as compared with the middle concentrations of metals in soils and where tissue metal  
19 concentrations remain stable over a wide range of soil metal concentrations.

20 Sample et al. (1998a) developed uptake models to predict concentrations in earthworms  
21 from soil concentrations. These models can be used to estimate the exposure of both the worms  
22 themselves and of vermivorous wildlife (e.g., song birds, voles, and shrews). For selected metals  
23 (arsenic, cadmium, copper, mercury, manganese, lead, and zinc), the best estimate of tissue  
24 concentration in earthworms is a simple ln-ln regression. The addition of soil pH data to the  
25 regression model did not markedly improve fit. If soil calcium concentration was incorporated  
26 into the regression model, a better fit could be obtained for cadmium and lead but not for other  
27 metals. Tissue concentrations were inaccurately estimated for the transition metals nickel and  
28 chromium by both simple and multiple regression models. For general estimates, log-linear  
29 regression models may be used as bioaccumulation models for arsenic, cadmium, copper,  
30 mercury, manganese, lead, and zinc in earthworms. For site-specific assessments, it is  
31 recommended that location-relevant bioaccumulation models be developed through direct  
32 measurements of local soil and tissue concentrations.

Trophic Transfer Values

*Trophic transfer values* for metals in terrestrial systems are an inverse function of soil concentrations; therefore, it is inappropriate to use constants for this term.

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**Oral intake (mg/kg/day) =**

$$[Soil_j \times P_s \times FIR \times AF_{js}] + \left[ \sum_{I=1}^N B_i \times P_i \times FIR \times AF_{ij} \right] \times AUF + [Water_j \times WIR]$$

where

Soil<sub>j</sub> = Concentration for contaminant (j) in soil (mg/kg dw)

P<sub>s</sub> = Soil ingestion rate (proportion of FIR)

FIR = Food ingestion rate (kg/kg-body weight/day, dw)

AF = Absorbed fraction of contaminant (j) in biota type (i)

N = Number of different types of biota in the diet

B<sub>i</sub> = Contaminant concentration in biota type (i) (mg/kg dw)

Note: (Soil<sub>j</sub> × T<sub>ij</sub>) can be substituted for B<sub>i</sub> where T<sub>ij</sub> = soil-to-biota trophic transfer factor (TTF) [as dry weight to dry weight] for contaminant (j) and biota type (I)

P<sub>i</sub> = Proportion of biota type (i) in diet (unitless)

AUF = Area use factor; proportion of available habitat for a wildlife species within the area of concern (%)

Water<sub>j</sub> = Concentration for contaminant (j) in water (mg/L)

WIR = Water ingestion rate (mL/kg/day)

1 **Figure 4-8. Wildlife Oral Exposure Model.**

2 **Source: U.S. EPA (2003c).**

3  
4 There is no compilation of plant biota/sediment accumulation factors (BSAFs), but U.S.  
5 EPA (2003c) (EcoSSLs) provides some data on select metals. For national-level assessments,  
6 uptake factors for plants provided by Efroymson et al. (2001) should be used. The highest  
7 accumulation of metals in plants occurs in the roots, although other parts of the plant also  
8 accumulate metals to varying degrees (Mitrofanov,1993; Greszta, 1982). With the exception of  
9 a few hyperaccumulator species, most plant species do not bioconcentrate metals (i.e., BAFs <1).  
10 Lead, arsenic, chromium, and cobalt are not taken up by plants in measurable quantities, and the  
11 small amount that is taken up is mostly confined to root tissues (Chaney et al., 2000; McGrath,  
12 1995; Chaney and Ryan, 1994; Xu and Thornton, 1985). Exceptions to this exist, notably  
13 including, but not limited to, Se and Mo: uptake of these metals into the edible portion of plant

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1 tissues can be quite high, generally not sufficient to cause plant toxicities (McGrath, 1995;  
2 Bingham et al., 1986; Foy et al., 1978), but may cause significant food chain toxicities .

3 In contrast, plants are quite sensitive to some metals (manganese, zinc, copper, for  
4 example) and may die before achieving high levels that pose a threat to animals via food chain  
5 transfer.

#### 7 **4.4.3.2. Limitations**

8 Food chain modeling for wildlife is similar for both metals and organic substances. All  
9 estimates are limited by the information available about the receptor(s) of concern in terms of  
10 dietary preferences, relative amounts consumed of various items, relative bioavailability between  
11 laboratory and field, and other factors (see Figure 4-8). For metals, the largest uncertainty in the  
12 dietary uptake model is in the estimates of trophic transfer factors. As discussed above, these  
13 generally are not constants and therefore require the use of uptake equations (i.e., tissue  
14 concentrations vary as a function of soil concentration). This necessitates the use of at least  
15 quasi-probabilistic modeling, rather than a more simple determination of a single, deterministic  
16 oral uptake value.

17 Use of either measured or modeled tissue metal accumulation levels as an indicator of  
18 potential toxicity is limited by the requirement that the accumulated amount be related to a  
19 benchmark effect level (i.e., Critical Body Residue, CBR). Very little information is available  
20 for metal CBRs in terrestrial wildlife. This is particularly problematic when whole-body tissue  
21 levels are reported because what really matters is the effective metal concentration at the site of  
22 action of toxicity. If the concentration at the proximate site of action of toxicity is proportional  
23 to the whole-body concentration, then this is a lesser concern. However, if the concentration at  
24 the site of action is not proportional to the whole-body concentrations, then direct measures of  
25 metal concentrations at the site of action is required, especially at higher assessment levels.

26 Furthermore, the absolute level of metal  
27 accumulation is not as important as the rate of uptake  
28 (Hook and Fisher, 2002; Hook, 2001; Roesijadi, 1992).  
29 High uptake rates overwhelm the ability of organisms to  
30 sequester or excrete the metal, leaving larger proportions of  
31 the accumulated metal in a more bioavailable form. Uptake is believed to occur because of the  
32 ability of some organisms to sequester metals that enter the cell (e.g., by inducing the synthesis  
33 of metallothionein [MT] or granule formation). Adverse effects are avoided as long as the rate

If the concentration at the site of action is not proportional to the whole-body concentration, then direct measures of metal concentrations at the site of action is required, especially when conducting detailed, higher level assessments.
--

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1 of metal uptake does not exceed the rate at which the organism is able to bind the metal, thereby  
2 preventing unacceptable increases in cytosolic levels of bioreactive forms of the metal. If the  
3 rate of uptake is too great, the complexation capacity of the binding ligand (e.g., MT) could be  
4 exceeded; cytosolic metal levels then become unacceptably high, and adverse effects could  
5 ensue. Because measures of uptake rates are not available, static concentrations are used instead.

6 Measurement of the form of the metal that is present in a tissue may be a more  
7 predictable indicator of the potential for effects than is total metal concentration (Mason and  
8 Jenkins, 1995; Roesijadi, 1992). Metals also may be bound and sequestered by  
9 organophosphorus granules, thereby rendering them nonavailable to bind with other intracellular  
10 target enzymes (George, 1982; Coombs and George, 1978). Although a wealth of available data  
11 exist on measured total tissue levels of metals, there are few data on intracellular speciation or  
12 sequestration of the metals. Although models that can be used to perform this sort of evaluation  
13 are currently under development, the ability of such models to be used as a tool in exposure  
14 assessment remains to be demonstrated. Therefore, measurement of total metal in plant (or  
15 animal) tissue remains the accepted default approach.

16 In sum, the following factors contribute significantly to uncertainty in food chain models:

- 17 • Soil ingestion rates are highly uncertain.
- 18 • Diet composition can be highly variable, and diet composition has a significant effect  
19 on exposure.
- 20 • Relative bioavailability from foods is completely unknown.
- 21 • Modeling over a number of trophic levels propagates uncertainty.
- 22
- 23
- 24
- 25
- 26

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1 **4.5. CHARACTERIZATION OF ECOLOGICAL EFFECTS**

2 Metals are naturally occurring substances, and organisms have evolved mechanisms for  
3 maintaining homeostasis in the presence of expected exposure levels. However, areas of metals  
4 enrichment, particularly when generated by anthropogenic activities, can pose challenges to  
5 organisms. Toxicity assessment for ecological receptors exposed to metals requires an  
6 understanding of both the natural mechanisms for tolerance for (or, in the case of micronutrients,  
7 the use of) metals and the toxicological responses that occur when exposure exceeds the capacity  
8 of the organism to regulate its body burdens. Interactions between metals in either their uptake  
9 or toxicity (such as Cd/Ca/Zn, Hg/Se, Cu/Mo) also should be considered in toxicity assessments.  
10 Risk assessments for metals are further complicated by the need to express the dose-response (or  
11 concentration-response) functions of bioavailable units that are functionally equivalent to  
12 measures of exposure. This section provides tools and approaches for addressing issues of  
13 essentiality, appropriate toxicity tests, novel endpoints (e.g., gene expression), and acclimation  
14 or adaptation to continued exposures.

15  
16 **4.5.1. Essentiality**

17 Essentiality, or the requirement for normal  
18 organism metabolic function, of many metals is one of the  
19 primary factors that differentiates risk assessment for  
20 metals and metal compounds from that of synthetic organic  
21 chemicals (Janssen and Muysen, 2001). Some trace  
22 elements, such as cobalt, copper, iron, manganese,  
23 selenium, molybdenum, and zinc, are necessary for the normal development of plants and  
24 animals. In many cases, these metals are added to animal feed and pharmaceutical products  
25 (SRWG, 2002) or to plant fertilizers. Other metals, such as arsenic, cadmium, lead, and  
26 mercury, are not known to be essential to plant and animal growth and development. Trace  
27 elements can be divided into three groups:

<p style="text-align: center;"><b>Essentiality</b></p> <p><i>Essentiality</i>, or the requirement for normal organism metabolic function, of many metals is one of the primary factors that differentiates risk assessment for metals and metal compounds from that of synthetic organic chemicals.</p>
---

- 28
- 29 • Those known to be essential.
- 30
- 31 • Those that have beneficial metabolic effects but have not been shown to be essential.
- 32
- 33 • Those that occur widely in living organisms but seem to be only incidental
- 34 contaminants and are not known to be beneficial (Mertz, 1981).
- 35

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1 Table 4-14 classifies the metals addressed in this framework by their known essentiality  
2 to organisms. The concept that many metals are required for organism health at one range of  
3 concentrations and are toxic in quantities outside of that range has been referred to as the  
4 “window of essentiality” (Hopkin, 1989) or the “optimal concentration range” for essential  
5 elements (Alloway, 1995; Fairbrother and Kapustka, 1997; Van Assche et al., 1997). For  
6 essential elements that exhibit biphasic dose-response curves (Figure 4-8, above), adverse effects  
7 resulting from deficiency should be considered, as well as those that result from excessive  
8 exposure. Recognition of the window of essentiality, as well as consideration of the biochemical  
9 and physiological processes that regulate metals within living organisms, are both important  
10 components of ecological effects assessment (Abernathy et al., 1993).

#### 11 **4.5.1.1. Application**

12 The optimal concentration range (or safe intake range) for essential elements should  
13 ensure that effects thresholds such as Toxicity Reference Values (TRVs) are not lower than the  
14 nutritional requirements for the particular plant or animal species being evaluated. Where TRVs  
15 or other effects concentrations (or doses) are intended as thresholds for detrimental effects due to  
16 excessive intake), care should be taken to ensure that these toxicity thresholds for essential  
17 metals are at the upper end of the optimum range or sufficiency range (at the point where toxic  
18 effects begin to occur). If set too low (i.e., in the range where deficiency can occur), the  
19 determination of risk will be erroneous. For wildlife, the literature on dietary requirements of  
20 essential elements for livestock can be consulted. The NAS/NRC has published useful  
21 summaries (NAS/NRC, 1994, 1980), and McDowell  
22 (2003) updates this information. Minimum  
23 concentrations required for plant growth are  
24 summarized in Marschner (1995).

#### 25 **4.5.1.2. Limitations**

26 Because of differences in test conditions  
27 among published studies, it may be difficult to  
28 directly compare toxicity threshold values with  
29 recommended dietary requirements of essential  
30 elements. Extrapolation of data among species (e.g.,  
31 from livestock to wildlife species) also may  
32  
33

#### **Threshold Values**

For essential elements, it is important to ensure that effects thresholds, such as Toxicity Reference Values (TRVs), are not lower than the nutritional requirements for the plant or animal species being evaluated. It may be difficult, however, to directly compare toxicity threshold values with recommended dietary requirements because of differences in test conditions among published studies.

In screening-level assessments, toxicity threshold values are advised for application, if they are not lower than estimated requirements. Detailed, higher level assessments may require additional bioassays to characterize the biphasic dose-response curve and determination of both required and excessive threshold levels.

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1 add uncertainty to the effects assessment. Furthermore, addition of safety factors when deriving  
 2 protective values often results in concentrations significantly below required intake. Derived  
 3 toxicity threshold values should be used in screening-level risk assessments if they are not lower  
 4 than estimated requirements. Uncertainty in toxicity thresholds or estimated requirements  
 5 should be addressed as part of the risk management process. Higher level assessments, where  
 6 more accurate estimates of effects thresholds are expected, may require additional bioassays to  
 7 characterize the biphasic dose-response curve and determination of both required and excessive  
 8 threshold levels. See the following sections for considerations of bioavailability factors,  
 9 mixtures of multiple metals, and interspecific extrapolations.

10 **Table 4-14. Metals classified by their known essentiality**

Metal	Essential (known requirement for health and function)		Beneficial (but not known to be essential)		Nonessential (and not known to be beneficial)
	Plants	Animals	Plants	Animals	
Aluminum (Al)					x
Antimony (Sb)					x
Arsenic (As)				x	
Barium (Ba)					x
Beryllium (Be)					x
Cadmium (Cd)					x
Chromium (Cr)		x			
Cobalt (Co)		x	x		
Copper (Cu)	x	x			
Lead (Pb)					x
Manganese (Mn)	x	x			
Mercury (Hg)					x
Molybdenum	x	x			
Nickel (Ni)	x	x			
Selenium (Se)		x	x		
Silver (Ag)					x
Strontium (Sr)					x
Thallium (Tl)					x
Vanadium (V)				x	
Zinc (Zn)	x	x			

Source: Adapted from a table presented in SRWG (2002) and incorporating data from NAS/NRC (1980) and Barak (1999). Fairbrother and Kapustka (1997) discussed the roots of essentiality of naturally occurring elements.

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## 4.5.2. Acclimation and Adaptation

Organisms have developed various mechanisms to cope with variable background metal concentrations, particularly for those metals that are essential elements (see Table 4-14). If the amount is less than required, passive or active uptake mechanisms are used to enhance internal concentrations, whereas during periods of excessive amounts exclusion mechanisms come into play. These various approaches to homeostasis are discussed in detail in Kapustka et al. (2004). Additionally, organisms can acclimate to suboptimal metal levels by changing various physiological functions, or populations can undergo genetic adaptation and develop increased tolerance to different levels (Rusk et al., 2004; Wallace and Srb, 1961).

The genetic makeup of an organism defines its ability to cope with environmental conditions. Genes can be expressed or remain “silent,” and shifts in gene expression can occur when the environment changes. Furthermore, organisms use different portions of their total array of genetic information in different life stages. This shifting of tolerance within the genetically defined limit of the organism is known as *acclimation*. Physiological changes induced by acclimation may be reversed if the environment reverts to the original conditions (Posthuma and Van Straalen, 1993; Prosser, 1986). Tolerance acquired through physiological acclimation processes is not always passed on to offspring; however, the same genetic information that allowed acclimation to occur in the parents will be passed on, so the offspring will retain the ability to acclimate in a similar fashion. If the offspring develop in the altered environment, they will express the set of genes most appropriate for tolerance of those conditions.

Genetic *adaptation* results from increased survival of tolerant genotypes and subsequent changes in gene frequencies. However, linking these genetic changes to increased tolerance in the field and identifying the specific mechanisms responsible has proven challenging. Laboratory experiments conducted with F<sub>1</sub> generations obtained from metal-contaminated habitats provide the strongest evidence to support a genetic basis of tolerance (Klerks and Levinton, 1993), and new methods in toxicogenomics (e.g., microarrays) are providing additional insights.

### **Tolerance, Acclimation, and Adaption**

***Tolerance*** is the ability of an organism to maintain homeostasis under a variety of environmental conditions, such as variable metal concentrations.

***Acclimation*** is how an individual develops tolerance during its lifetime, and it may be gained or lost. Acclimation is also called ***phenotypic plasticity***.

***Adaptation*** is a genetic change over multiple generations as a response to natural selection. Traits are not lost during single life times. Adaptation is also known as ***genotypic plasticity***.

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1 Evidence of adaptation (convergent evolution)  
2 for metal tolerance in plants comes from the fact that  
3 plants of diverse taxonomic relationships grow on soils  
4 high in metals. Metal-tolerant flora have been  
5 described for soils high in zinc, nickel, chromium, and  
6 copper (Antonovics et al., 1971; Brooks, 1972). These  
7 reviews indicate that some species are restricted to the  
8 high-metal soils, but other species exist across a broad  
9 concentration range. These represent differences in  
10 niche breadth (i.e., those restricted to high-metal soils  
11 vs. those occurring in soils that have either high or low  
12 concentrations of metals).

#### Costs of Metal Tolerance

Although considerable evidence supports the hypothesis that previously exposed populations will be tolerant to metals, both physiological acclimation and adaptation to contaminants may have specific costs. For example, induction of metal-binding proteins increases metals tolerance but also uses energy normally available for other metabolic processes (e.g., growth, reproduction). Similarly, genetic changes associated with metals exposure might harm populations. Reduced genetic diversity has been reported in populations exposed to contaminants and may result in population bottlenecks as well as increased susceptibility to other stressors.

13 Metal adaptation in plants often is accompanied by metal adaptation in co-located  
14 animals, as selection for metal tolerance is expected to improve fitness in exposed conditions  
15 (Posthuma and Van Straalen, 1993). Compared with other environmental parameters, metal  
16 exposure is regarded as a strong and stable selective force, and it can lead to rapid  
17 evolution of tolerance (Posthuma and Janssen, 1995). Metal-tolerant animals tend to grow fast,  
18 mature early, and have a high excretion efficiency (Posthuma and Janssen, 1995).

19 Metal adaptation in natural populations of terrestrial invertebrates has been demonstrated  
20 conclusively for several animals: the terrestrial isopod *Porcellio scaber* and the springtails  
21 *Orchesella cincta*, *Isotoma notabilis*, and *Onychiurus armatus* (Posthuma and Van Straalen,  
22 1993). Metal tolerance also has been demonstrated in ticks and a fly species in response to the  
23 application of a metal-based pesticide (Posthuma and Van Straalen, 1993). There is evidence for  
24 increased metal tolerance in other species, but acclimation and adaptation could not be  
25 distinguished (Posthuma and Van Straalen, 1993).

26 Although there is considerable evidence to support the hypothesis that previously  
27 exposed populations will be tolerant to metals, both physiological acclimation and adaptation to  
28 contaminants may have specific costs. For example, although induction of metal-binding  
29 proteins increases tolerance to subsequent metal exposure, it also uses energy normally available  
30 for other metabolic processes (e.g., growth, reproduction). Similarly, genetic changes associated  
31 with exposure to contaminants might harm populations. Reduced genetic diversity has been  
32 reported in populations exposed to contaminants and may result in population bottlenecks.  
33 Furthermore, as tolerant genotypes are eliminated from a population, the reduced genetic  
34 diversity may increase the susceptibility of this population to other stressors.

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1 **4.5.2.1. Application**

2 Metal tolerances of plants and livestock are summarized in Table 4-15. It should be  
3 noted that the NAS/NRC (1980) committee that identified the maximum levels of metals  
4 tolerated by domestic livestock based its conclusions on data from toxicological-type feeding  
5 studies in which soluble metal salts had been mixed with practical or purified diets to examine  
6 animal response to the dietary metals. If soil is incorporated into diets, metal solubility and  
7 bioavailability may be much more limited than in the tests relied on by the NRC. For example, it  
8 has been noted that until soil exceeds about 300 mg Pb kg<sup>-1</sup>, animals show no increased body  
9 burden from ingesting the soil (Chaney and Ryan, 1993). Other metals in equilibrium with  
10 poorly soluble minerals or strongly adsorbed in soils are often much less bioavailable than they  
11 would be if they occurred in more soluble salts.

12 It has been shown in studies with *Daphnia* and algae that acclimation of test organisms to  
13 culture conditions will significantly influence the outcome of toxicity tests (Janssen and  
14 Heijerick, 2003; Bossuyt and Janssen, 2002).

15 Presumably, similar effects occur with terrestrial  
16 organisms (e.g., earthworms or springtails) (see  
17 Fairbrother et al., 2002). Thus, organisms should  
18 be acclimated to the same waters or soils that will  
19 be used in the toxicity studies for several weeks  
20 before study initiation. This will avoid shocking  
21 the organisms by placing them in test conditions  
22 that have significantly depleted or elevated levels of  
23 endogenous metals (e.g., those that are not the  
24 subject of the toxicity study) and ensure that any  
25 adverse effects are actually due to the metal in  
26 question. However, it is not possible to conduct  
27 pre-acclimation for soil microbial studies because  
28 in situ populations of microorganisms from  
29 previously collected soils are used. This calls into question the applicability of results from  
30 microbial function studies, and until these issues are resolved, results should not be used for  
31 setting soil criteria (U.S. EPA, 2003e).  
32

**PICT**

For site-specific assessments, the concept of *pollution-induced community tolerance (PICT)* has been proposed as a tool to assess community-level effects. PICT is tested by comparing responses of communities collected from polluted and reference sites to contaminant exposures under controlled conditions. The increase in community tolerance at a polluted site that results from the elimination of sensitive species is considered evidence that this restructuring was caused by the pollutant.

Extrapolation of these results to communities of terrestrial plants and animals should be undertaken with caution, since most of the original research on PICT has been conducted using small organisms with relatively fast life cycles (e.g., benthic invertebrates, soil microbial communities).

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**Table 4-15. Maximum tolerable levels of dietary minerals for domestic livestock compared with levels in forages**

Element	Soil-plant barrier <sup>a</sup>	Level in plant foliage <sup>b</sup>		Maximum levels chronically tolerated <sup>c</sup>			
		Normal	Phytotoxic	Cattle	Sheep	Swine	Chicken
		mg/kg dry foliage		mg/kg dry diet			
As (inorg.)	Yes	0.01–1	3–10	50	50	50	50
B	Yes	7–75	75	150	~150	~150	~150
Cd <sup>d</sup>	Fails	0.1–1	5–700	0.5	0.5	0.5	0.5
Cr <sup>3+</sup>	Yes	0.1–1	20	~3000	~3000	~3000	~3000
Co	Fail?	0.01–0.3	25–100	10	10	10	10
Cu	Yes	3–20	25–40	100	25	250	300
F	Yes?	1–5	–	40	60	150	200
Fe	Yes	30–300	–	1000	500	3000	1000
Mn	?	15–150	400–2000	1000	1000	400	2000
Mo	Fails	0.1–3	100	10	10	20	100
Ni	Yes	0.1–5	50–100	50	~50	~100	~300
Pb <sup>d</sup>	Yes	2–5	–	30	30	30	30
Se	Fails	0.1–2	100	~10	~10	10	10
V	Yes?	0.1–1	10	50	50	~10	10
Zn	Yes	15–150	500–1500	500	300	1000	1000

<sup>a</sup>Indicates whether a plant will die at concentrations lower those that which will affect animals.

<sup>b</sup>Based on literature summarized in Chaney et al. (1983).

<sup>c</sup>Based on NAS/NRC (1980). Continuous long-term feeding of minerals at the maximum tolerable levels may cause adverse effects. NRC estimates levels by extrapolating between animal species when data are not available.

<sup>d</sup>NRC based the maximum levels tolerated of Cd or Pb in liver, kidney, and bone in foods for humans rather than simple tolerance by the animals. Because of the simultaneous presence of Zn, Cd in animal tissues is less bioavailable than Cd salts added to diets.

Yes = Plant dies; Fails = Plant does not act as a barrier and concentrates metal to levels for toxic animals; ? = Unknown.

1 Most research examining tolerance to metals has focused on population-level responses,  
 2 although consequences at the community level also occur. The most common explanations for  
 3 increased tolerance of populations within communities include acclimation and selection for  
 4 resistant genotypes. However, because communities consist of large numbers of interacting  
 5 species, it is likely that other mechanisms unique to these systems will contribute to increased

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1 tolerance. For example, the replacement of sensitive species by tolerant species, termed  
2 “interspecific selection” (Blanck et al., 1988), is a common response in polluted systems and one  
3 of the most consistent indicators of metal pollution. Pollution-induced community tolerance  
4 (PICT) has been proposed as an ecotoxicological tool to assess effects of contaminants on  
5 communities (Blanck et al., 1988). PICT is tested by comparing responses of communities  
6 collected from polluted and reference sites to contaminant exposures under controlled  
7 conditions. The increase in community tolerance at a polluted site that results from the  
8 elimination of sensitive species is considered evidence that this restructuring was caused by the  
9 pollutant.

10 Naturally occurring levels of metals play an important role in biogeographic distributions  
11 of plants and animals and may, in fact, be limiting factors in species distributions or use of  
12 landscapes. Therefore, it is difficult to make generalizations about effects levels that are  
13 applicable and consistent to all organisms in all habitats, and it becomes very important to  
14 clearly define the geospatial location of the area to which the assessment results will apply.  
15 For site-specific assessments, the assessment results will be directly applicable to the entire  
16 range of species that may be found on that site (although for assessments conducted over areas  
17 larger than several square miles, it is possible that multiple soil types and other local landforms  
18 may result in significant differences in metal bioavailability and plant communities.

19 For assessments conducted for regional or national assessments, criteria development, or  
20 ranking purposes, it should be acknowledged that results will be based on organisms and soil  
21 types that result in greatest bioavailability and sensitivity. Care should be taken, however, that  
22 the organism-environment combinations that are assessed are, in fact, compatible with real-  
23 world conditions. For example, benthic organisms generally associated with slow-moving,  
24 warm waters would not be expected to tolerate conditions of high metal bioavailability such as  
25 those occur that in faster-moving, colder waters that have little organic matter. Thus, for site-  
26 specific assessments, species tested and water (or sediment) used in the test system should be  
27 similar to conditions at the site. In the absence of such information, data from standard test  
28 species and conditions could be used, but uncertainty factors may be warranted to adjust the final  
29 toxicity value accordingly.

30 More appropriately, single-result assessments for the entire country should be avoided.  
31 Rather, such assessments should be subdivided into metal-related ecoregions known as  
32 “metalloregions” (McLaughlin and Smolders 2001) so that protection levels, mitigation goals,  
33 and ranking results will be appropriate for the suite of species naturally present within each type  
34 of controlling environment. This is directly analogous to the use of ecoregions when

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1 establishing WQC (Griffith et al., 1999). The value of metalloregions is that they provide the  
2 conceptual framework to account not only for the broad regional parameters affecting metal  
3 availability in soils and waters, but also for the differences in organism response to added metal.

#### 4 5 **4.5.2.2. Limitations**

6 Although there is considerable evidence to  
7 support the hypothesis that previously exposed  
8 populations will be tolerant to metals, both physiological  
9 acclimation and adaptation to contaminants may have  
10 specific costs (Wilson, 1988). For example, although  
11 induction of metal-binding proteins increases tolerance  
12 to subsequent metal exposure, it also uses energy  
13 normally available for other metabolic processes (e.g.,  
14 growth, reproduction). Similarly, genetic changes

15 associated with exposure to contaminants might harm populations. Reduced genetic diversity  
16 has been reported in populations exposed to contaminants and may result in population  
17 bottlenecks. Furthermore, as tolerant genotypes are eliminated from a population, the reduced  
18 genetic diversity may increase the susceptibility of this population to other stressors. There is  
19 theoretical support for the hypothesis that populations adapted to contaminants have higher  
20 metabolic costs or are more susceptible to other stressors (Hoffman and Parsons, 1997; Mulvey  
21 and Diamond, 1991); however, few empirical studies have demonstrated increased costs.

22 One of the assumptions behind the use of PICT as an ecotoxicological tool is that  
23 differences in tolerance among communities can be detected using short-term experiments. This  
24 significantly constrains the application of PICT as an assessment tool. Although tolerance at the  
25 population level can be assessed using a variety of species, logistical considerations will limit the  
26 types of communities where tolerance can be investigated experimentally. Most of the original  
27 research on PICT has been conducted using small organisms with relatively fast life cycles (e.g.,  
28 benthic invertebrates, soil microbial communities). Therefore, extrapolation of these results to  
29 communities of terrestrial plants and animals should be done with caution.

30 The metalloregion concept (McLaughlin and Smolders, 2001), although intuitively  
31 appropriate, has not yet been fully developed for the United States. The country has been  
32 divided into ecoregions for both aquatic and terrestrial systems (Bailey et al., 1994; Bailey,  
33 1983). These are based on climactic and vegetation factors and form the basis of metalloregions.  
34 EPA is still working to complete ecoregion maps at much finer scales for each state (see EPA

#### **Metalloregions**

For national-scale assessments, the entire country can be subdivided into metal-related ecoregions (known as “metalloregions”) to help ensure that protection levels, mitigation goals, and ranking results will be appropriate for the suite of species naturally present within each type of controlling environment. However, the metalloregion concept has not yet been applied across

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1 Web site at <http://www.epa.gov/wed/pages/ecoregions/ecoregions.htm>). To complete the  
2 metalloregion concept, soil properties that affect bioavailability (e.g., pH, cation exchange  
3 capacity (CEC), and OM) should be overlaid on the ecoregions, along with soil type (e.g.,  
4 sandy loam, clay loam) and background metal concentrations of metals. Similar information is  
5 needed for water bodies. Although this type of information is fairly current and available, soil  
6 data have not been updated since the mid-1970s, which may limit their usefulness to some  
7 extent. Nevertheless, work is under way to develop metalloregions, although it is likely to be  
8 several years from the time of this writing before they are available for use in a decision-making  
9 capacity.

### 11 **4.5.3. Metals Mixtures**

12 Mixtures of metals (including metalloids and other contaminants) are commonly  
13 encountered in the natural environment as a result of anthropogenic inputs. Metal interactions,  
14 according to Calamari and Alabaster (1980), occur at three levels:

- 15 • Chemical interactions with other constituents in the media;
- 16 • Interactions with the physiological processes of the organism during uptake; and
- 17 • Interactions at the site of toxic action.

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22 Much of the difficulty in interpreting the available information on the toxic effects of  
23 metal mixtures is due to differing measures and definitions of the bioavailable fraction of metals,  
24 whether it is the fraction that is available for uptake from the environment or at the site of toxic  
25 action. Some measure of the bioavailable metal fraction in the exposure media is needed to  
26 accurately predict the effects of metals and metal mixtures (Di Toro et al., 2001; Sauvé et al.,  
27 1998; Weltje, 1998; Posthuma et al., 1997; Ankley et al., 1996). Characterization of effects of  
28 metal mixtures has also been reported to be concentration dependent (Mowat and Bundy, 2002;  
29 Fargašová, 2001; Herkovits et al., 1999; Spehar and Fiandt, 1986).

#### 31 **4.5.3.1. Studies of Metal Mixtures**

32 Few controlled studies exist on the toxicologic interactions of metals found in  
33 environmental contamination scenarios. ATSDR has compiled and evaluated interaction studies  
34 involving various metals: methyl mercury, arsenic, cadmium, chromium, lead manganese, zinc,  
35 copper, cesium, cobalt, strontium, and uranium (draft interaction profiles available online at

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1 <http://www.atsdr.cdc.gov/iphome.html> ). Few studies allowed any quantification of interaction  
2 magnitude, whether using the authors' definitions of toxicologic interaction or EPA's definitions  
3 based on dose and response addition. The summaries below indicate some of the qualitative  
4 conclusions available on interaction potential.

5 A study of a mixture of cadmium, lead, and zinc in rats found slightly more marked  
6 adverse hematological effects with the ternary mixture exposure than with the cadmium-lead,  
7 cadmium-zinc, or lead-zinc mixtures (Thawley et al., 1977); inconsistencies in dietary levels of  
8 calcium and vitamin D in this study, however, may have made comparisons problematic. A  
9 well-controlled rat study has reported significant synergism between cadmium and lead  
10 regarding testicular atrophy (Saxena et al., 1989). That study also demonstrated protective  
11 effects of high dietary levels of zinc, which effectively reduced the testicular effects of the  
12 cadmium-lead mixture to control levels. No studies have been located that would allow  
13 extrapolation of those high exposure results to more common, lower environmental levels. In  
14 another study (Fowler and Mahaffey, 1978), a relatively wide range of endpoints were  
15 investigated in studies that covered each metal singly and all possible binary and ternary  
16 mixtures. Body weight gain was depressed equally by the ternary mixture and the cadmium-lead  
17 mixture, and to a lesser extent by the arsenic-lead and cadmium-lead mixtures, whereas food  
18 utilization was depressed more by the ternary and arsenic-cadmium mixtures than by the other  
19 binary mixtures. In general, the biological parameters studied in this report indicated changes of  
20 smaller magnitude and inconsistency in direction for binary mixtures compared with ternary  
21 mixtures.

22 The data regarding interactions of environmental metals usually are not adequate for  
23 predicting the magnitudes of interactions. Interaction profiles by ATSDR of metal-metal  
24 interactions have considered the following combinations: arsenic, cadmium, chromium and lead;  
25 lead, manganese, zinc, and copper; and cesium, cobalt, and strontium (see above Web site for  
26 draft reports). Experimental efforts to identify and quantify interaction mechanisms among these  
27 metals are needed. For some endpoints, the data are not robust enough to show even the  
28 direction of interaction (i.e., whether the joint action will be dose additive or greater than or less  
29 than additive). The animal studies discussed briefly in this report used commercial diets or semi-  
30 purified diets that may have higher or lower levels of essential metals than human diets. Much  
31 higher doses of the metals appear to be required to elicit effects when commercial diets are used  
32 than when semi-purified diets are used. At the other extreme, effects are seen at very low doses  
33 when deficient diets are used. Comparisons among studies are therefore problematic,  
34 particularly when the diets are not specified.

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### **4.5.3.2. Mixtures**

Similar to human health risk assessments, ecological risk assessments commonly involve mixtures of metals. However, Agency guidance is less developed for how to estimate metal mixture effects for wildlife. Binary interactions of dietary metals in livestock are discussed in NAS/NRC (1994, 1980) and McDowell (2003). These references can be used as sources of information from which at least qualitative estimates of additivity, synergy, or antagonism can be developed. Additionally, see section 4.3.6.3. for discussion of human health assessment for mixtures, which provide information useful in consideration of wildlife effects.

### **4.5.3.3. Application**

Two key questions should be addressed by effects assessments related to metal mixtures:

- To what extent does each metal contribute to any observed effect?
- Are the effects significantly greater than or lesser than the sum of the individual component effects?

The answers to these questions also have the potential to affect water quality guidelines (WQGs), EcoSSL values, cleanup targets, and other similar management decisions.

Methodologies (graphical and statistical) to predict impacts of metal mixtures and interactions of individual metals within mixtures can be broadly classified as either Concentration Addition models or Effects Addition models. Both models use metal water concentrations to generate concentration-response curves for individual metals, and these data are then used to generate specific critical concentrations for mixture models. Similar models can be developed for soils or sediments.

In the Concentration Addition model, all metals in a mixture are added together to predict toxicity; differing potencies are taken into account by converting chemical concentrations to an equitoxic dose (e.g., Toxic Units (TUs) or Toxicity Equivalence Factors (TEFs), which convert all metals to one metal concentration). In the Effects Addition model, differing potencies are ignored, and the effect of each metal's concentration in a mixture is combined to predict mixture toxicity. Only the Concentration Addition model allows detection of toxicity greater than predicted (more than additive); the Effects Addition model can only predict strict additivity.

The TU approach involves deriving dimensionless units for each metal in a mixture by dividing individual concentrations by their individual toxic concentrations (such as LC<sub>50</sub> values).

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1 The TUs for all the metals in the test mixture are then summed. A value of  $1 \pm 0.1$  in the case of  
2 an  $LC_{50}$  would predict 50% mortality (or another effect if TUs are based on that effect rather than  
3 on lethality). A value significantly greater than 1 would predict more than 50% mortality. A  
4 value significantly less than 1 would predict less than 50% mortality.

5 The TU approach can be used with any endpoints, for instance  $LC_{50}$  or  $EC_{50}$  values.  
6 However, it assumes the same mode of action for all the chemicals so will only predict additive  
7 effects. The toxic concentration can be derived from guideline values (e.g., WQGs), from  
8 literature toxicity data, or from specific experiments. This approach can be used when setting  
9 WQC. It has, for example, been recommended for use as part of the Australia and New Zealand  
10 WQGs (ANZECC and ARMCANZ, 2000). These guidelines employ a concentration addition  
11 approach using WQG concentrations of metals as TUs:

$$12 \quad TTM = \sum(C_i/WQG_i)$$

13  
14  
15 where TTM is the predicted total toxicity of the mixture,  $C_i$  is the concentration of the  
16 component, and  $WQG_i$  is the guideline for that component. If TTM exceeds 1, then the mixture  
17 has exceeded the water quality guideline. It is important to note that this has been developed for  
18 water, and significant limitations may be associated with applying it to soil systems.

19 Norwood et al. (2003) conducted a literature review on the effects of metal mixtures.  
20 Mixtures varied from 2 to 11 metals. The investigators determined that the TU approach is  
21 presently the most appropriate model for predicting effects of metal mixtures, based on currently  
22 available data (e.g., effect concentrations,  $EC_x$  values). Effects addition models, especially if  
23 based on body or tissue concentrations, might be more accurate in the future, but they require  
24 reliable dose-response and bioaccumulation curves for all single metals (not just  $EC_x$  values)  
25 and then careful testing of the models (research on tissue concentration effect levels is ongoing).

26 **Application of HSAB.** The QICAR approach (described in Section 4.1, Environmental  
27 Chemistry) might also be applicable to predict the potential for interactions of metals in mixture.  
28 Unsatisfied with the qualitative conclusions of Newman and McCloskey (1996), Ownby and  
29 Newman (2003) fit binary metal mixture data derived from the Microtox assay to the following  
30 model of joint independent action (Finney, 1947):

$$31 \quad \text{Predicted } P_{A+B} = P_A + P_B - P_A P_B$$

32 where:  
33  
34

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1  $P_{A+B}$  = The biological response to the metal mixture, expressed as a proportion

2  $P_A$  and  $P_B$  = The biological responses to A or B when present singly

3  
4 This relationship would hold if the actions of the paired metals were independent.  
5 However, if the metals were not independent, the  $P_A P_B$  term will deviate from its expected value  
6 of  $z P_A P_B$  where  $z = 1$ . Assuming that paired metals with very similar binding tendencies are  
7 more likely to interact with the same biological ligands than are metals with very dissimilar  
8 tendencies, deviations of the  $z$  coefficient from 1 would reflect departure from complete  
9 independence of metal action. HSAB theory allowed prediction of metal interactions in this  
10 model system. Although Microtox is considered to be a useful tool for organic contaminants, it  
11 is very sensitive for metals (Willemson et al., 1995).

#### 13 **4.5.3.4. Limitations**

14 At present, the most appropriate approach for determining the toxicity of metal mixtures  
15 and addressing the two key questions listed above is to use the TU approach as a screening-level  
16 assessment. This approach cannot, however, be used beyond screening because it does not  
17 provide enough certainty. Norwood et al. (2003) found that of 191 case studies examined, 70%  
18 were additive or less than additive. Thus, this approach was primarily either appropriate or  
19 overprotective, but 30% of the case studies indicated  
20 that this approach would be underprotective. For  
21 aquatic organisms acutely exposed to cationic metals,  
22 the assumption of additivity is sufficient, particularly if  
23 bioavailability adjustments are made using the BLM  
24 (see Section 3.4.8).

25 Currently, there are no realistic means of ranking  
26 mixtures of metals or individual metals within mixtures.  
27 National criteria for mixtures also are not possible at  
28 this time. Furthermore, the concentration addition (TU)  
29 approach is recommended only for application to  
30 mixtures with fewer than six components. This issue remains site specific because interaction  
31 responses are dependent on both the actual metal mixture combinations (metals and ratios) and  
32 the exposed organisms.

33 Researchers are presently attempting to predict the effects of metal mixtures on the basis  
34 of critical body concentrations of metals. Ongoing research is attempting to integrate the effect

#### **Toxic Unit Approach**

Currently, the most appropriate approach for determining the toxicity of metal mixtures is to use the *Toxic Unit (TU) approach* as a screening-level assessment tool. This approach cannot, however, presently be used beyond screening because it does not provide adequate certainty since combined effects depend on the relative amounts of each metal. Furthermore, the TU approach is recommended for applications with mixtures containing less than six components.

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1 accumulation functions of each metal into a metal mixture model in which an effect addition  
2 formula will be compared with a concentration addition formula, both based on body  
3 concentration. Current work is focused on aquatic invertebrates, and similar research is required  
4 for other organisms.

5 It is possible that the BLM may be expanded in the future to include mixtures. In theory,  
6 if two metals compete for binding to the same site of toxic action on an organism, it should be  
7 possible to model the total metal bound to that site and hence predict metal toxicity using a  
8 mechanistic BLM approach in an Effects Addition model. Alternatively, if two metals do not  
9 compete for the same binding site on the organism, then the BLM may provide more reliable  
10 estimates of individual metal bioavailability, and these estimates can then be combined in more  
11 accurate Effects Addition models. However, at present, these possibilities remain theoretical and  
12 need testing. However, this possibility, while improving the ability to assess the effects of metal  
13 mixtures, does not include temporal aspects (i.e., “time-to-response” versus concentration).  
14

#### 15 **4.5.4. Background**

16 Background is defined as the amount of metals  
17 occurring in soils, water, or air as a result of  
18 anthropogenic and natural processes. Anthropogenic  
19 contributions are limited to those that are not influenced  
20 by current, direct releases (i.e., emissions, discharges, or  
21 disposal) from a source or site of concern. This  
22 includes metals that may arise from manmade  
23 substances (particularly metalloids) or from natural  
24 substances (metallic ores) present in the environment as  
25 a result of human activity that are not specifically  
26 related to the release in question (U.S. EPA, 2003c).

27 Background should be defined in a specific spatial and  
28 temporal aspect that is related to the scope of the  
29 particular hazard or risk assessment. Background  
30 concentrations can vary by as much as five orders of  
31 magnitude, depending on soil type, geography, and other factors (Chapman and Wang, 2000).

32 Background may exacerbate toxicological effects and accumulations of metals from  
33 direct emissions or other regulated sources or, conversely, it may result in adaptation of  
34 organisms to higher metal concentrations and result in increased tolerance to emissions (see

#### **Background**

*Background* is defined as the amount of metals occurring in soils, water, or air as a result of anthropogenic and natural processes. Anthropogenic contributions are limited to those that are not influenced by current, direct releases (i.e., emissions, discharges, or disposal) from a source or site of concern. This includes metals that may arise from manmade substances (particularly metalloids) or from natural substances (metallic ores) present in the environment as a result of human activity that are not specifically related to the release in question. Background can exacerbate toxicological effects and accumulations from direct emissions or other sources, or conversely it may result in adaptation of organisms to higher metal concentrations and result in increased tolerance to emissions.

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1 above section). Furthermore, because metals occur naturally, and some are essential macro- or  
2 micronutrients, they are at least partially responsible for how plants and animals are distributed  
3 within various ecoregions. The distribution of plants and animals, local species diversity,  
4 species survival, and the vitality of individuals can be profoundly affected by background levels  
5 of metals in an area. Humans, on the other hand, are distributed throughout the world,  
6 irrespective of naturally occurring levels of metals, so knowledge of background levels is not as  
7 significant.

#### 8 9 **4.5.4.1. Application**

10 Concentrations of metals in soils and waters of the United States vary tremendously.  
11 Thus, use of a single number to represent all areas within the United States is to be discouraged.  
12 Statewide averages (U.S. EPA, 2003c) provide somewhat better resolution, but even these are  
13 constrained by political boundaries, not by geochemical characteristics. Additional information  
14 on concentration of metals in soils at smaller spatial resolutions is provided in Shacklette and  
15 Boerngen (1984). Some metals (e.g., Fe, Cu, and Zn) are included in the State Soil Geographic  
16 Database (STATSGO) available at [www.nrcs.usda.gov/technial/techtools/stat\\_browser.html](http://www.nrcs.usda.gov/technial/techtools/stat_browser.html).  
17 These data can be grouped at whatever spatial scale is required, but they are not screened for  
18 whether they represent true background concentrations. Similarly, data on water concentrations  
19 can be retrieved from EPA's STORET database ([www.epa.gov/storet/index.html](http://www.epa.gov/storet/index.html)). These data  
20 should be used with caution, however, as this is a voluntary-entry database, and there is no  
21 consistent method for measurement or for quality assurance/quality control (QA/QC) of the data.  
22 Like the STATSGO information, STORET data do not necessarily represent true background  
23 levels; additionally, there is incomplete coverage across the United States.

24 Similar data for sediments are available from the EPA's National Sediment Quality  
25 Survey database, which is available in the form of an MS Access 1997 database and can be  
26 obtained on a CD from the EPA's Office of Water, Office of Science and Technology (EPA,  
27 OW, OST). It contains survey data from 1980 to 1999, including sediment chemistry data, tissue  
28 residue data from selected organisms, and toxicity data (lethal and sublethal effects on various  
29 test organisms). Overall, there are data from more than 50,000 stations and 4.6 million  
30 analytical observations. The data were compiled from a variety of sources, but mostly from state  
31 and federal monitoring programs, and sampling and analysis strategies varied among sources.  
32 All data have "data qualifiers" associated with them in the database. However, this was not a  
33 statistically designed survey, and it is heavily biased toward contaminated sites, so inferences to  
34 areas that were not sampled should be made with caution.

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1 A more appropriate approach is to define average (and ranges) of background  
2 concentrations for various ecoregions (Hargrove and Hoffman, 1999; Bailey, 1998; Omernick,  
3 1986). Work is under way in this regard, but such values currently are not available. Therefore,  
4 the recommended default is to use state averages where possible and always to define the range  
5 that might be encountered within the spatial scale being considered. It is also important to  
6 recognize that background concentrations in water are highly variable through time because  
7 flooding or drought conditions substantially change the relative concentrations of metals in a  
8 water body. Again, it is suggested that ranges rather than single number averages be used.

9 Several practical issues should be considered when evaluating the contribution of  
10 background to hazard or risk and its implications for various risk management options when  
11 conducting site-specific assessments. First, a physical and/or temporal boundary should be  
12 defined for the analysis. Next, background should be described, estimated, or measured. U.S.  
13 EPA (2002b, c) provides detailed guidance on how to estimate local background concentrations  
14 and notes that locations of background samples should be areas that could not have received  
15 contamination from the site but that have the same characteristics as the medium of concern (i.e.,  
16 water, soil, or air).

17 Another reason to include background in the assessment is to evaluate the effect of the  
18 remedial options. Although some areas of the site may have elevated concentrations for certain  
19 metals, other metals may not be elevated. Remedial actions could cause these naturally  
20 occurring metals to become more bioavailable, thereby resulting in unintended toxicity. The  
21 HSAB theories and QICAR models (see Section 4.1.2) can be applied to derive useful estimates  
22 of biological activity for metals to aid in making decisions regarding remedial actions.

23 **Incorporating background.** How the information on background concentrations is used  
24 in the final risk assessment depends on how data on toxic responses were generated and the  
25 relationship of the bioavailability characteristic of the naturally occurring material to the source-  
26 related additional metal. For aquatic organisms, toxicity tests generally are conducted in waters  
27 that are relatively low in background metals and are of moderate hardness. Therefore, the  
28 toxicity thresholds described (e.g., in WQC) represent the total amount of metal in the water, not  
29 the amount that can be added to the natural background levels. If, however, the tests are run on  
30 site-specific waters where metal background and bioavailability may differ significantly from  
31 standard waters and where organisms have been acclimated to such conditions, then the toxicity  
32 threshold reported should include both the background and the amount that is added. When  
33 using this approach, caution should be exercised, because the background levels of metals in the  
34 aquatic system may be highly variable over short time periods. The primary consideration is that

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1 the *bioavailable fraction* of the natural background concentrations should be added to the  
2 *bioavailable fraction* of the added metal.

3 In terrestrial systems, toxicity bioassays for soil organisms (plants and invertebrates) are  
4 conducted either in artificial soils or in actual soils, both of which contain background  
5 concentrations of metals (particularly of essential elements). Therefore, the toxic levels reported  
6 are the amount added during the study. Unfortunately, the background amounts frequently are  
7 not reported, so it may not be feasible to take a true “added risk” approach. Furthermore, the  
8 amount of metal added to the test system generally is in a much more bioavailable form (e.g., a  
9 metal salt) than is the background material. Therefore, for a site-specific release of a highly  
10 bioavailable form of the metal, the background concentration may not contribute significantly to  
11 total metal uptake. Thus, it becomes extremely important to measure exposure in terms of the  
12 *bioavailable fraction*, so field exposures can be expressed in a manner comparable to the highly  
13 bioavailable material used in toxicity tests for threshold setting.

14 For human health and wildlife assessments, the amount of metal in food material should  
15 be taken into consideration in a manner similar to that discussed above for soil organisms.  
16 Again, differences in bioavailability of food-incorporated metals and top-dressed metal salts  
17 should be considered. Additionally, natural uptake and the amount of metals in forage and other  
18 food items will vary, depending on the amount of metal in soil and the particular species of  
19 plant/animal present in the area. Site-specific assessments can take this into consideration. For  
20 national or large-scale assessments, a default assumption can be made that food items contain  
21 sufficient amounts of micronutrients to meet dietary needs and that the toxic threshold value  
22 represents the *bioavailable fraction* that is added above these values.

#### 23 24 **4.5.4.2. Limitations**

25 National databases of metal concentrations in soil or water do not currently differentiate  
26 between naturally occurring levels and levels that are elevated owing to anthropogenic sources.  
27 However, the databases can be screened to ascertain whether specific areas are affected by point  
28 source emissions and so can be used as reasonable estimates for large-scale background levels.  
29 More important, however, is that background concentrations are most frequently reported as total  
30 amount of metal, without specifying the bioavailable fraction or chemical speciation or the data  
31 (e.g., pH or redox potential) needed to estimate bioavailability. Information on probable mid- to  
32 long-term changes in soil or water properties that might enhance bioavailability of background  
33 metals also should be provided for accurate assessments of future risk.

34  
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1 **4.5.5. Indirect Effects of Metals**

2 Metals can initiate ecological changes by directly  
3 affecting individual organisms (i.e., through toxic  
4 responses). Organisms can die, fail to reproduce, or have  
5 altered behavioral patterns. As a consequence of such  
6 actions on individuals, other organisms within the  
7 community will be indirectly affected through reduced  
8 number of prey items or predators or changes in  
9 competition for resources. Additionally, indirect interactions between organisms can and do  
10 occur independently of initiating effects from toxicity due to metals exposures (described by Dill  
11 et al., 2003). Although “initiators” may be biotic, physical, or chemical, in all cases there is an  
12 effect to a species (the “transmitter”) that has an effect on another species (the “receiver”).

13 Indirect effects to organisms initiated by metals toxicity can be negative (density  
14 dependent or independent) or positive (density dependent) and can occur between species or  
15 within the same species. Some examples are summarized in Table 4-16; additional examples are  
16 provided by Chapman et al. (2003). Interactive effects can also occur due to the combined  
17 effects of environmental stressors and metals toxicity. For instance, Liess et al. (2001) found  
18 that the toxicity of copper to an Antarctic amphipod was increased by food shortage and  
19 excessive ultraviolet-B radiation. Some authors have considered such interactive effects to be  
20 indirect; however, they can be addressed as direct effects in risk assessments and hence are not  
21 considered further.

**Indirect Effects**

*Indirect effects* to organisms initiated by metals toxicity can be negative (density dependent or independent) or positive (density dependent) and can occur between species or within the same species. Table 4-16 summarizes some of these relationships.

22  
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1 **Table 4-16. Examples of Indirect Effects of Metal Toxicity**

2

3 <b>Transmitter organism</b>	<b>Receiver organism</b>	<b>Comment</b>
4 Prey populations reduced	Predator populations reduced	Negative indirect effects due to reduced prey (food)
5 Predator populations reduced	Prey populations increased	Positive indirect effects due to reduced predation
6 Competitor populations reduced	Competitor populations increased	Positive indirect effects due to reduced competition
7 Toxicity to some larvae of a species	Increased adult-to-larval survival, increased growth and biomass of adults of the same species	Positive indirect effects due to density-dependent compensation

8

9

#### 10 **4.5.5.1. Application**

11 Functional redundancy (species having similar roles in ecosystem processes) is a well-  
12 known but arguably not well-understood phenomenon in ecosystems. It is often assumed that if  
13 an organism's primary prey item is reduced or eliminated, then the organism can switch to  
14 another prey item due to functional redundancy. However, in the case of food web-mediated  
15 effects, the following sequence can and does apply: chronic metal exposure reduces the food  
16 abundance of certain dietary components; as a result there are increased energetic costs of  
17 feeding and an associated reduced growth efficiency. It is important that all components of the  
18 food web are exposed to metals; thus, the determination of the final effect should include the  
19 concurrent changes to all elements.

20 A determination of whether indirect effects such as loss of preferred prey can occur  
21 requires three components. First, appropriate conceptual diagrams should be developed in the  
22 problem formulation phase of the risk assessment and subsequently refined. Such diagrams  
23 should incorporate sufficient detail regarding key biotic interactions (e.g., competition and  
24 predation) and the ecological context in which the species exist and pollution occurs. As noted  
25 by Chapman et al. (2003), seasonal and life-stage changes in feeding patterns can occur and will  
26 "require iterative temporal diagrams showing the various reasonable possibilities." If the factor  
27 is limiting, then the organism will exhibit a response; however, if it is not a limiting factor, the  
28 likelihood that it will lead to organismal changes decreases.

29 Second, risk assessors should focus proactively on the first three possibilities in Table 4-  
30 16 because it is extremely unlikely that indirect effects due to density-dependent compensation  
31 will be detectable without extraordinary effort. Increased numbers of receiver organisms due to  
32 reduced competition or predation are more likely to be detected and more likely to have overall

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1 adverse implications for communities and/or ecosystems, despite their apparent positive aspects  
2 (to the particular receiver species).

3 Third, risk assessors should focus on any cases of reduced growth of individuals,  
4 particularly where the individuals affected are relatively tolerant to metals toxicity *and* their  
5 normal feeding patterns are disrupted. These are clear indications (although not conclusive  
6 proof) of indirect effects of metals toxicity. An example of negative indirect effects due to  
7 reduced prey is provided by Campbell et al. (2003). These authors summarize extensive field  
8 research into yellow perch in metal-impacted lakes in eastern Canada. Yellow perch in lakes  
9 shift from feeding on zooplankton to feeding on littoral macrobenthos during their second year  
10 of growth and then begin to include a significant amount of fish in their diet during their third to  
11 fifth year of growth. Fish in the most metal-impacted lakes did not undergo this normal  
12 sequence of diet shifts. Instead, they continued to utilize smaller prey throughout their lives  
13 owing to the loss of their primary prey species to direct metal toxicity. A bioenergetic  
14 bottleneck developed because the perch's growth efficiency was reduced by the need to catch  
15 and eat smaller prey. The perch were more tolerant to metals toxicity, and thus there were no  
16 major direct effects on their survival (though multiple physiological effects were recorded).  
17 However, the loss of their primary prey species resulted in smaller or stunted perch, a major  
18 indirect effect of metals toxicity. Many methods exist in the ecological literature to gather  
19 population, community, or ecosystem function data that may be achieved without significant  
20 levels of effort. Some elegant analyses of population demographics can be accomplished within  
21 a single sampling effort (e.g., Wilson et al., 1996; Heyer et al., 1994; Davis, 1982).

#### 23 **4.5.5.2. Limitations**

24 It is difficult to predict natural (e.g.,  
25 behaviorally mediated), indirect interactions in nature  
26 (Dill et al., 2003). It is also difficult to predict indirect  
27 interactions due to abiotic factors such as metals  
28 toxicity. Accurate predictions require good  
29 understanding of the functional interactions within and  
30 between populations, particularly along food chains, as  
31 well as density-dependent and density-independent  
32 processes between and within species.

Presently there is no realistic means of ranking metals on the basis of indirect effects, nor are national criteria possible. This issue remains a site-specific one because interactions between the initiator (metal toxicity), the transmitter, and receptor organisms are dependent on both the level and type of toxicity and the sensitivities of individual organisms within structurally and functionally unique populations, communities, and food chains.

33 Presently there is no realistic means of ranking metals on the basis of indirect effects, nor  
34 are national criteria possible. This remains a site-specific issue because interactions between the

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1 initiator (metal toxicity), the transmitter, and receptor organisms are dependent on both the level  
2 and type of toxicity and the sensitivities of individual organisms within structurally and  
3 functionally unique populations, communities, and food chains.

4 As noted by Campbell et al. (2003) and Dill et al. (2003), indirect effects of stressors on  
5 organisms probably are not uncommon occurrences. However, there are few documented cases  
6 of this occurring, and it is likely that most cases go unrecognized. It would be useful to review  
7 previous detailed-level ecological risk assessments involving metals toxicity to determine and  
8 document previous cases.

9 Indirect effects from metals toxicity are primarily associated with ecosystem function,  
10 not structure. However, ecological risk assessments typically are focused on determining risk to  
11 structure, not function. The assumption is made that measuring structure protects function  
12 because of functional redundancy. Clearly, given the reality of indirect effects, this is at best a  
13 questionable assumption that needs to be tested.

#### 14 15 **4.5.6. Bioavailability in Terrestrial Systems**

16 Bioavailability of soil metals to terrestrial  
17 organisms is closely linked to dynamic soil physical and  
18 chemical parameters, and biotic processes. As discussed  
19 in Section 4.1, there are qualitative and quantitative  
20 methods and models for considering soil chemistry issues  
21 and aging of metals in soil that modify metal  
22 bioavailability and its toxicity in soil. Unless specific  
23 bioavailability data exist, even with measures of soil  
24 physical and chemical parameters such as soil loading  
25 capacity, aging of metals, and speciation, accurate  
26 estimates of exposure to the terrestrial biota cannot be made. In situations where information is  
27 not available, bulk soil chemistry is typically used with a default of 100% relative  
28 bioavailability.

Unless specific bioavailability data exist, even with measures of soil physical and chemical parameters such as soil loading capacity, aging of metals, and speciation, accurate estimates of exposure to terrestrial biota are not possible. In situations where information is unavailable, bulk soil chemistry is typically used with a default of 100% relative bioavailability (i.e., the bioavailability of the chemical in soil is presumed to be the same as the bioavailability in the tests used to develop the toxicity data).
---

##### 29 30 **4.5.6.1. Soil Organisms: Invertebrates and Microorganisms**

31 Metal speciation is the primary consideration in assessing the bioavailability of metals to  
32 soil invertebrates and microbes. Major assumptions regarding metal exposure in aquatic  
33 systems, such as a the relatively homogeneous dissolution of metals in the exposure water, may  
34 not be applicable or may apply at different scales in soil systems. Although soil microbes may

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1 be immersed in soil solution films surrounding soil particles, few invertebrates are exposed to  
2 metals in this manner. Exposure usually consists of partial contact of soil solution films with the  
3 surfaces of the invertebrates that are capable of absorbing metals (e.g., earthworm dermal  
4 surfaces). Direct contact with membranes across which metal uptake can occur does not take  
5 place for many hard-bodied soil invertebrates (e.g., arthropods), and metal uptake is almost  
6 entirely through the ingestion of metal associated with particle matter or soil solution. For these  
7 reasons, exposure and relative bioavailability cannot be expressed similarly for each organism in  
8 the soil ecosystem, and an understanding of primary routes and mechanisms of metal exposure  
9 should be established for species or groups of similar  
10 organisms.

Exposure and relative bioavailability cannot be expressed similarly for each organism in the soil ecosystem, and an understanding of primary routes and mechanisms of metal exposure should be established for species or groups of similar organisms.

11 Often, direct toxicity testing of the soil of concern  
12 is the best method for assessing bioavailability and  
13 toxicity-to-soil biota (Fairbrother et al., 2002). Issues such  
14 as spiking of metals solutions onto soils, aging, and  
15 laboratory-to-field extrapolation should be considered. In  
16 the ecological soil screening document (U.S. EPA, 2003c), published literature was evaluated  
17 using primary soil parameters affecting lability of metals in soils in a matrix to qualitatively  
18 indicate metal bioavailability. Further information on this topic and factors relating soil  
19 chemistry to soil biota toxicity are discussed in Section 4.1.6. A terrestrial BLM method  
20 currently under development (Allen, 2002) may provide a useful tool to link bioavailability, soil  
21 chemistry, and toxicity to soil biota.

#### 22 23 **4.5.6.2. Plants**

24 The most common route of metal exposure in  
25 plants is through the roots. Ions and organic molecules  
26 contact roots via the transpiration stream, diffusive  
27 transport, and microbe-facilitated transport. At the root  
28 surface, soluble contaminants have the potential to enter  
29 the root tissue through the transpiration stream or through  
30 a range of mechanisms designed to facilitate nutrient uptake. In general, it is thought that only  
31 uncomplexed, free ionic species of cations and ions can be taken up by roots. This has been  
32 described using a FIAM (Parker and Pedler, 1997; Lund, 1990). Sauvé et al. (1998) put forward

Ionic or organometallic complexes that increase the total concentration of elements at the root surface have been correlated with increased uptake, either through disassociated ions or through uptake of intact complexes. Thus, research into how to quantify metal complexes in soils relative to their lability is important.

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1 a method to calculate free metal activity levels for copper and lead to derive a pH-dependent soil  
2 criterion for soil biota. The soil criteria paper was based in part on previous studies (Sauvé et al.,  
3 1997a, b, 1995; McBride et al., 1997) to generate models to predict free copper activity from  
4 total metal content, pH, and organic matter content or, in the case of lead, using only total metal  
5 levels and pH. The equations were generated from more than 60 soils using ion selective  
6 electrodes and standard methods for determining pH, organic matter content, and total metal  
7 levels. However, many exceptions to the FIAM have been identified, such that it has been  
8 abandoned in favor of its close cousin, the BLM. Ionic or organometallic complexes that  
9 increase the total concentration of elements at the root surface have been correlated with  
10 increased uptake, either through disassociated ions or through uptake of intact complexes (Parker  
11 et al., 2001; McLaughlin et al., 1994). In addition, it is clear that plants do not distinguish well  
12 between many pairs of ions of similar charge and size (e.g., As and P or Cd and Zn).

13 Plant bioassays can be used to measure the relative bioavailability of metals in various  
14 soil types. Results can be used to determine either the direct or the indirect value of  
15 bioavailability of contaminants in plants and to extrapolate an indirect estimate of relative  
16 bioavailability to organisms that consume the plants (assuming a correlation between plant and  
17 animal uptake). This type of testing has been routinely done in agriculture for decades and has  
18 been used to validate extraction tests. Tests have most often focused on identifying plant  
19 deficiencies of particular elements, but they are easily adapted to evaluate toxicities (Gettier et  
20 al., 1985).

21 In the absence of test data, relative bioavailability can be estimated qualitatively based on  
22 relative pH and organic matter content (Tables 4-17 and 4-18), although other soil factors can be  
23 significant, most notably CEC. However, combinations of these two soil parameters and their  
24 ranges are sufficient as a qualitative guide for assessors to identify soils where metals may have  
25 increased (or decreased) availability to plants. This is particularly noted when soil pH exceeds 7  
26 and is a mechanism to obtain essential elements under conditions of low availability.

27  
28  
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1 **Table 4-17. Qualitative bioavailability of metal cations in natural soils to plants and**  
 2 **soil invertebrates**  
 3

Soil type	Soil pH		
	Low organic matter (<2%)	Medium organic matter (2 to <6%)	High organic matter (6 to 10%)
4 ≤ Soil pH ≤ 5.5	Very high	High	Medium
5.5 < Soil pH ≤ 7	High	Medium	Low
7 ≤ Soil pH ≤ 8.5	Medium	Low	Very low

10  
11  
12 **Table 4-18. Qualitative bioavailability of metal anions in natural soils to plants and soil**  
 13 **invertebrates**  
 14

Soil type	Soil pH		
	Low organic matter (<2%)	Medium organic matter (2 to <6%)	High organic matter (6 to 10%)
4 ≤ Soil pH ≤ 5.5	Medium	Low	Very Low
5.5 < Soil pH < 7	High	Medium	Low
7 ≤ Soil pH ≤ 8.5	Very High	High	Medium

15  
16  
17  
18  
19  
20  
21 Source: U.S. EPA (2003e).

22  
23 **4.5.6.2.1. Critical Tissue Residues.** When used appropriately, plant tissue analysis can provide  
 24 an indirect, semiquantitative assessment of bioavailability processes (NAS/NRC, 2002). For  
 25 phytotoxic metal concentrations to be effectively used as an indirect measure of bioavailability,  
 26 it is important that the threshold values of the plant tested be well understood. In addition,  
 27 toxicities of certain metal elements are associated with deficiencies of others. For example,  
 28 increased zinc, copper, and nickel toxicities can be associated with iron deficiencies (Bingham et  
 29 al., 1986), and increased lead and zinc toxicities can also be related to phosphorus deficiencies

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1 (Brown et al., 2000, 1999a; Laperche et al., 1997). The behavior of plant species in response to  
2 nutrient deficiencies varies, and this behavior can affect the uptake of metal elements  
3 (Marschner, 1998). Because of these multiple confounding factors, the bioavailability of metals  
4 in plants (as well as to consumers) is more accurately and reliably measured directly as the  
5 edible plant tissue concentrations of the metal in association with soil metal concentrations in the  
6 root zone (NAS/NRC, 2002).

#### 7 8 **4.5.6.3. Wildlife**

9 For most metals, the dietary intake pathway is the main route of exposure for wildlife  
10 (NAS/NRC, 2002; see also Section 4.3). However, the incidental ingestion of soil can often  
11 contribute a large portion to the majority of the exposure to a wildlife consumer. Because many  
12 inorganic metals do not readily accumulate in food (organometallic compounds are excluded  
13 from discussion here), highly contaminated soil may result in higher exposures to metals through  
14 activities such as grooming fur, preening feathers, consuming soiled prey or forage, burrowing,  
15 and taking dust baths. However, canopy feeders would be anticipated to have less incidental soil  
16 ingestion and therefore less exposure to inorganic metals than wildlife that consume food that is  
17 in more intimate contact with the ground.

18 The relative importance of the dietary and incidental soil ingestion pathways is dictated  
19 by (1) the fraction of total metal available in soil versus that in food and (2) the relative  
20 bioavailability of the metal in the soil as compared to metal in food items. Figure 4-7 in Section  
21 4.2.3. shows the relative contribution of food and soil to total metal exposure before accounting  
22 for bioavailability. Understanding of the bioavailability of metals in incidentally ingested soils  
23 becomes necessary when there is a high amount of metal in the soil that is not taken up by soil  
24 organisms (plants or invertebrates). However, the same variables that restrict uptake by plants or  
25 other soil organisms act to reduce bioavailability to wildlife that ingest soil directly. Therefore, a  
26 qualitative estimate of low relative bioavailability could be made for these soils. Furthermore,  
27 data that are generated for human health studies could be used to estimate soil bioavailability for  
28 wildlife, acknowledging the uncertainty inherent in such interspecies extrapolations.

29 **Dietary bioavailability.** Very little information is available on dietary bioavailability for  
30 most wildlife species (see Menzie-Cura and TN&A, 2000), for a review, cited in NAS/NRC,  
31 2002). One of the most significant challenges is that the bioavailability of metals may be  
32 influenced by differences in digestive physiology and anatomy across the broad and diverse  
33 range of mammalian and avian species. For example, metals present in soils or food may be

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1 more or less bioavailable within the gut of an herbivore that relies on fermentation as compared  
2 to the comparatively simpler gut of a carnivore that is designed to break down proteins. These  
3 gut systems differ in chemistry (including pH) and residence time. The potential differences  
4 between species have not yet been explored rigorously. In general, ruminants absorb lower  
5 amounts of metals than do monogastric animals such as rats (NAS/NRC, 1980). Some general  
6 guidelines are provided for some metals (e.g., lead in NAS/NRC (1980)), and human-derived  
7 values can be used as default values in the absence of species-specific data.

8 **Critical body residues (CBRs).** CBRs are internal concentrations of chemicals that are  
9 correlated with the onset of a toxic response (Conder et al., 2002; Lanno et al., 1998). The use of  
10 CBRs reduces uncertainties in ecological risk assessment procedures because they account for  
11 site-specific bioavailability and multipathway issues (Van Straalen, 1996; Van Wensem et al.,  
12 1994 ). CBRs can be based on whole-body residues (see below for discussion of this approach  
13 in soil invertebrates) or concentrations in specific tissues. Tissue-specific critical loads have been  
14 established for several species of vertebrate wildlife for lead in liver, cadmium in kidney, and  
15 selenium in eggs. See Beyer et al., 1996, for these figures.

#### 16 17 **4.5.7. Bioavailability in Aquatic Systems**

18 Many factors influence the bioavailability of inorganic metals in aquatic systems.  
19 Abiotic (e.g., organic carbon, pH, cations) and biotic (e.g., uptake and metabolism) modifying  
20 factors determine the amount of metal that interacts at biological surfaces (e.g., at the gill) and  
21 subsequently is taken up. In the dissolved phase, metals can exist as free ions as well as in a  
22 variety of complexed forms. These forms, or species, are of key importance in understanding  
23 potential impacts because they have differing bioavailabilities, and therefore water quality can  
24 dramatically influence the proportion of bioreactive forms of a metal. For many metals in  
25 aquatic systems, it is the free ionic form which is believed to be responsible for toxicity. For  
26 example,  $\text{Cu}^{2+}$  has been directly linked to toxicity in fish and invertebrates, while Cu complexed  
27 by dissolved organic matter does not induce toxicity to the same degree (Ma et al., 1999;  
28 Erickson et al., 1996) because bioavailability for uptake is reduced.

29 The relationship between speciation and bioavailability is expressed through the FIAM  
30 (Campbell, 1995). However, the FIAM is not without limitations, as links between metal  
31 speciation and toxicity are complicated and the free metal ion is not always the only bioreactive  
32 form. For example, complexed metal, including Cu bound to DOC can be taken up and  
33 contribute to toxic impacts and effects (McGeer et al., 2002; Erickson et al., 1996). Although

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1 the link between bioavailability of metals and factors influencing speciation (such as pH,  
2 temperature, and organic and inorganic anionic complexation) are of prime importance, other  
3 abiotic factors, particularly cations, influence metal bioaccumulation and toxicity. Dissolved  
4 cations such as  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ , and  $\text{Mg}^{2+}$  can competitively inhibit metal uptake. BLMs, the  
5 recently developed integrated toxicity prediction models, have successfully combined abiotic  
6 speciation, cationic competition, and bioaccumulation at the presumed site of toxic action  
7 (reviewed by Paquin et al., 2002a). The influence of exposure conditions on metal  
8 bioavailability is beginning to be understood for some metals, but speciation of metals within the  
9 organism is generally less well understood. However, speciation within the organism is likely  
10 important for storage, metabolism, elimination, and delivery to and bioavailability at the site of  
11 toxic action (Mason and Jenkins, 1995).

12 It is important to account for bioavailability in relation to exposure conditions when  
13 comparing toxicity studies and relating these to natural environments. This includes knowing  
14 the species of metal that is likely to be present in the environment. A number of other variables  
15 also can influence the outcome of a laboratory toxicity assay, including the acclimation of test  
16 animals to the culture conditions (see Section 4.5.2, Acclimation, for further discussion), the  
17 natural background concentrations of the metals (including, but not limited to, the metal of  
18 interest) in either the test water or the site of concern (see Section 4.5.4, Background), potential  
19 interactions of the various metals (see Section 4.5.3, Mixtures), and the potential of the metal to  
20 transform to a bioreactive species (see Section 4.1, Environmental Chemistry). The relative  
21 compositions of the pretest culture medium and the underlying test medium also should be  
22 examined because sudden changes in ionic composition have been shown to cause sufficient  
23 stress to sensitize organisms and enhance toxicity (Taylor et al., 1990). This is a necessary  
24 prerequisite for successful comparison of effect levels derived from laboratory tests to predicted  
25 exposures.

26 The bioavailability of metals in aquatic systems can vary significantly among taxonomic  
27 groups. For example, although most mayflies (*Ephemeroptera*) are generally sensitive to metals,  
28 caddisflies (*Trichoptera*) and many stoneflies (*Plecoptera*) are relatively tolerant (Clements et  
29 al., 1992). In fact, these species-specific differences in sensitivity to contaminants have  
30 motivated the development of numerous indices of water quality that are based on composition  
31 of benthic communities (Barbour et al., 1992; U.S. EPA 1989a; Hilsenhoff 1987). Because  
32 responses of aquatic organisms to chemical disturbances are usually contaminant specific

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1 (Slooff, 1983), field assessments of metal effects may be difficult when more than one  
2 contaminant is present, and caution is required when using biotic indices to assess effects of  
3 complex effluents. In some situations, it may be necessary to either develop chemical-specific  
4 indices on the basis of sensitivity to specific classes of contaminants or calibrate metrics used in  
5 individual field assessments. For example, metals discharged into a system dominated by highly  
6 sensitive taxa (e.g., certain heptageniid mayflies) will have greater effects than the same effluent  
7 discharged into a system dominated by tolerant organisms (e.g., certain chironomids). Kiffney  
8 and Clements (1996) showed that benthic communities from headwater streams were more  
9 sensitive to metals than were communities from mid-elevation streams.

#### 11 **4.5.7.1. Application**

12 A variety of methodologies (e.g., hardness adjustments, FIAM, Water Effect Ratio  
13 (WER), and aquatic BLMs ) can be used to account for differences in concentrations of  
14 bioavailable metal species when assessing the effects of metals in aquatic systems (U.S. EPA,  
15 1999b). These bioavailability considerations are important in understanding the exposure  
16 conditions of interest and how these relate to toxicity study results. WER determinations can  
17 account for site-specific (i.e., dependent on the water quality at a specific location)  
18 bioavailability (U.S. EPA, 1994c), although they require the development of animal test data.  
19 Hardness adjustments were among the first computational methods to account for bioavailability  
20 when applying WQC. FIAM approaches that produce speciation profiles of a metal in an aquatic  
21 system provide insight into the relative bioavailabilities of the different forms of metal and the  
22 importance of complexation. Several models are available for the calculation of metal speciation  
23 in natural waters, including MINEQL (Schecher and McAvoy, 1994; Westall et al., 1976),  
24 MINTEQA2 (Brown and Allison, 1987), CHESS (Santore and Driscoll, 1995), WHAM  
25 (Tipping, 1994) and PHREEQ (Parkhurst, et al., 1980). The BLM approach further extends  
26 bioavailability considerations because it applies the latest information on the chemistry and  
27 physiological effects of metals in aquatic environments (Di Toro et al., 2001; Santore et al.,  
28 2001; Paquin et al., 1999).

29 Quantitative Ion Character Activity Relationships (QICARs) can be used to extrapolate  
30 from availability and/or toxic data for a metal for which there are data to a metal for which data  
31 are limited. The approach is similar to the Quantitative Structure-Activity Relationships  
32 (QSARs) that have been used for organic compounds. The QICAR approach was recently

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1 integrated into the general context of modern QSAR practices by McKinney et al. (2000), and  
2 Weltje (2002) successfully applied it to the lanthanides. The QICAR approach is best applied  
3 with a full understanding of the system under study, and the approaches should be validated  
4 before application. This approach is but one example of the types of tools that are available but  
5 that still need further development to support metals risks assessments for aquatic systems.

6 In the BLM, chemical speciation is simulated as an equilibrium system that includes  
7 complexation of inorganic ions and NOM using the CHEMICAL Equilibria in Soils and Solutions  
8 (CHESS) model (Santore and Driscoll, 1995). It includes a description of metal interactions  
9 with NOM based on the WHAM (Tipping, 1994). The simulation of biological interactions is  
10 based on the binding affinity characteristics measured in gill-loading experiments, originally  
11 described by Playle et al. (1993a, b). The biotic ligand  
12 (i.e. the gill) is represented as having a characteristic  
13 binding site density and conditional stability constants  
14 for each of the dissolved chemical species with which it  
15 reacts. Predictions of metal toxicity are made by  
16 assuming that the dissolved metal LC50, which varies  
17 with water chemistry, is always associated with a fixed  
18 critical level of metal accumulation at the biotic ligand.  
19 This fixed level of accumulation at 50% mortality is  
20 referred to as the LA50. The LA50 is assumed to be  
21 constant, regardless of the chemical characteristics of  
22 the water (Meyer et al., 1999, 2002).

The FIAM and BLM approaches both consider:

- Chemical speciation of the metal and the relative amounts of complexation to inorganic and organic ligands.
- The relative bioavailability of metal species (for example but not limited to the free metal ion) and interactions with other cations that may compete for binding sites of uptake or toxicity.
- Accumulation of metal by the organisms at the site of toxic action.

23 The BLM approach has been applied successfully to combine the influences of speciation  
24 and cationic competition on metal toxicity. (Di Toro et al., 2001; Santore et al., 2001; McGeer et  
25 al., 2000; de Schamphelaere and Janssen, 2002, 2004; de Schamphelaere et al., 2002, 2003,  
26 2004; Heijerick et al., 2002a, 2002b, also see example application for Cu below)., The model can  
27 distinguish, at least conceptually, metal that will bioaccumulate and cause toxicity from the total  
28 metal pools in an organism and the bioavailable metal pool in the exposure media. Themodeling  
29 approach has been extended to species such as algae (Heijerick et al., 2002; de Schamphelaere et  
30 al., 2003) and *Daphnia* (de Schamphelaere and Janssen, 2002, 2004; de Schamphelaere et al.,  
31 2002, 2004), which are toxicologically relevant but more difficult to characterize in terms of  
32 accumulation at the site of toxic action.

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1 While the focus of currently existing BLMs is acute toxicity, the Ion Balance Model  
2 (Paquin et al., 2002a, b,c) can be used to extend the applicability of the BLM to evaluate chronic  
3 toxicity. The Ion Balance model uses the BLM to predict accumulation levels at the biotic ligand  
4 and then explicitly represent the degree of the physiological response of the organism to metal  
5 exposure over time (i.e., disruption of ionoregulation and gradual loss of plasma sodium).  
6 Although the approach, which was initially applied to silver, may ultimately provide a way to  
7 predict effects due to metals over varying exposure durations, further development and testing is  
8 required. Chronic toxicity models have recently been developed for the impacts of Cu on  
9 *Daphnia magna* (de Schamphelaere and Janssen, 2004) and of Zn on fish, *Daphnia* and algae in  
10 European Union risk assessments on zinc.

11 Overall, the BLM approach has wide application in terms of understanding  
12 bioavailability in relation to toxicity. It incorporates the influence of speciation in the exposure  
13 medium, bioaccumulation and toxic impacts in a robust approach, and has been applied in a  
14 variety of contexts. For example, the BLM has recently been incorporated into draft revisions to  
15 EPA's water quality criteria for Cu, it has been used in risk assessment and it is being applied as  
16 an alternative to the Water Effect Ratio approaches for setting site specific discharge objectives.  
17 When considering the application of this approach, as with all models, care should be taken to  
18 understand and explicitly account for the assumptions and potential sources of uncertainty.

#### 19 20 **4.5.7.2. Limitations**

21 Novel applications such as the FIAM and BLM have  
22 been shown to offer dramatic improvements over traditional  
23 approaches such as the hardness equations which depend on  
24 the empirical relationship between water hardness and  
25 toxicity and the correlation of water quality variables with  
26 hardness. While these new approaches offer improvements,

EPA has developed a draft criteria document for copper that incorporates the BLM and will undergo peer review. The draft criterion document is available on the EPA/OST Web page at [www.epa.gov/waterscience](http://www.epa.gov/waterscience).

27 there are still many unknowns and uncertainties in relation to bioavailability and the biotic and  
28 abiotic influences on metal toxicity. For example, the role of natural dissolved organic matter in  
29 bioavailability and moderating toxicity is not well understood. There are considerable research  
30 efforts ongoing currently, and it is likely that our understanding of metal bioavailability and the  
31 method for integrating this knowledge into prediction models will improve quickly in the coming  
32 years.

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1           The BLMs that have been and are being developed for a subset of metals (e.g. Cu, Ni,  
2 Cd, Ag, Pb, Zn) are based on a limited number of species. BLMs are generally focused on  
3 predicting acute toxicity, although there are a few examples of BLMs that predict chronic  
4 toxicity, as discussed previously. The development of BLMs to predict chronic toxicity will  
5 likely require new knowledge about the combined effects of dietary and waterborne exposure,  
6 the mode of toxicity, and the accumulation of metal or dose delivered to the biological site of  
7 toxic action. Similarly, acclimation responses, which can influence bioaccumulation at the site  
8 of toxicity, are not well understood, and plant and animal species can differ considerably with  
9 regard to the forms of metal taken up as well as their relative toxicities. Although these and  
10 other issues add complexity to the evaluation, the BLM approach considers metal  
11 biogeochemistry; and therefore represents a viable avenue toward understanding and predicting  
12 the toxicity of metals.

#### 14 **4.5.7.3. Example: BLM Application to Development of Copper Aquatic Life Criteria**

15           The BLM's ability to incorporate metal speciation reactions and organism interactions  
16 allows for the prediction of metal toxicity to a variety of organisms over a wide range of water  
17 quality conditions. Accordingly, the BLM is an attractive tool for deriving WQC in EPA's water  
18 program. Application of the BLM may eliminate the need for site-specific criteria modifications,  
19 such as water effect ratios, which are currently used to account for site-specific chemistry  
20 influences on metal toxicity. EPA currently is using the BLM to develop a freshwater aquatic  
21 life criteria criterion maximum concentration (CMC) for copper. The BLM accounts for  
22 inorganic and organic ligand interactions of copper and also considers competitive interactions  
23 that influence binding of copper at the site of toxicity. Although a new model is being used, the  
24 criterion derivation is still based on the principles set forth in the 1985 guidelines (Stephen et al.  
25 1985). To develop a BLM-based criterion, model predictions of critical accumulations on the  
26 biotic ligand (LA50 values) and either LC50 or EC50 values are needed to calculate species  
27 mean acute values (SMAVs) and genus mean acute values (GMAVs) as well as to derive a  
28 species sensitivity distribution.

29  
30 **4.5.7.3.1. Model Input Parameters.** Much of the aquatic toxicity literature reviewed for the  
31 derivation of the copper criterion neither measured nor reported many of the key BLM input  
32 parameters. In these cases, the input parameters were estimated. A detailed description of the

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1 methods used to obtain or estimate these input parameters is included in U.S. EPA (2003a).  
2 Briefly, when critical water chemistry parameters were not available, a variety of strategies were  
3 employed to find the additional or surrogate data (e.g., authors or lab personnel were contacted,  
4 and alternative sources such as studies with similar water quality U.S. Geological Survey's  
5 National Stream Quality Accounting Network (NASQAN; <http://water.usgs.gov/nasqan/>) and the  
6 EPA STOrage and RETrieval (STORET; <http://www.epa.gov/STORET/>) were used). Where  
7 sources could not be used to develop geochemical input parameters for the BLM, data were  
8 generated using the reported water hardness and regression relationships constructed from  
9 NASQAN data.

10  
11 **4.5.7.3.2. *Quality Ranking of Water Chemistry Input Parameters.*** A ranking system of 1 to 6  
12 was devised to evaluate the quality of the chemical characterization of the test water (but not the  
13 overall quality of the study). Studies that included all of the necessary BLM input parameters,  
14 based on measurements from either the test chambers or the source water, were assigned a  
15 ranking of 1. Rankings of 2 to 4 were assigned to studies that did not measure all parameters but  
16 provided reliable estimates of ion concentrations. Studies were assigned a ranking of 5 to 6  
17 when one of the key parameters (DOC, Ca, pH, or alkalinity) was not measured and could not be  
18 reliably estimated or if two or more key parameters (DOC, Ca, pH, or alkalinity) were not  
19 measured. Only those studies with a rank of 1 to 4 were used to derive the criterion.

20 As with any modeling effort, the reliability of model output depends on the reliability of  
21 the input. Although the input data have been extensively scrutinized and filtered, the reliability  
22 of the BLM-derived values developed for copper in this project are subject to the limitations of  
23 the input measurements/estimation procedures described above.

24  
25 **4.5.7.3.3. *Criteria Generation.*** To calculate an acute criterion or CMC, reported acute toxicity  
26 values (e.g., LC50s) and individual test water chemistry parameters were used to calculate LA50  
27 values by running the model in the speciation mode. These LA50 values were then  
28 “normalized” to a standard water condition by running the model in the toxicity mode and  
29 specifying user-defined LA50s. These normalized LC50s were used to calculate SMAVs,  
30 GMAVs, and a final acute value (FAV) pursuant to the 1985 guidelines procedure. The FAV  
31 represents a hypothetical genus more sensitive than 95% of the tested genera. The FAV was  
32 derived from the four GMAVs that have cumulative probabilities closest to the 5th percentile

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1 toxicity value for all the tested genera. Inputting this FAV as an LC50 concentration and  
2 running the model in speciation mode determines the lethal accumulation associated with the  
3 FAV in the standard test water.

4 The Cu BLM assumes that most of the parameters are invariant for different organisms.  
5 Despite the complexity of the modeling framework, the thermodynamic constants used to  
6 simulate the inorganic and organic chemical equilibrium reactions are determined by the  
7 characteristics of the metal and available ligands. As such, the constants do not change for  
8 simulations involving different organisms. Although most BLM parameter values (including the  
9 biotic ligand binding constants and site densities) are consistent across organism types,  
10 differences in sensitivity across organisms types should still be accounted for. This is  
11 accomplished by adjusting the critical biotic ligand concentration (e.g., LA50) values for each  
12 species.

13 This criterion LA50 is programmed into the model as a constant. To derive a criterion  
14 for a specific site, the site water chemistry data are input to the model. The model then uses an  
15 iterative approach to determine the dissolved copper concentration needed to achieve a Cu-biotic  
16 ligand concentration equal to the criterion LA50. This dissolved Cu concentration is, in effect,  
17 the FAV based on site water chemistry. The site-specific CMC is this predicted dissolved metal  
18 concentration (or criterion FAV) divided by two. The site-specific CCC is the CMC divided by  
19 the final acute-chronic ratio.

20  
21 **4.5.7.3.4. Next Steps.** EPA has developed a draft criteria document that will undergo peer  
22 review. When EPA solicits scientific views from the public on the draft criterion document, the  
23 model will be made available on the Office of Water web page at [www.epa.gov/waterscience](http://www.epa.gov/waterscience).  
24 Until the peer review is completed and a final copper criteria document is published by EPA, the  
25 procedures described here are draft and subject to change, and the criteria are not considered to  
26 be available for use. After completing the copper criteria update, EPA will consider  
27 incorporating the BLM into derivation procedures for other metal criteria, such as for silver,  
28 cadmium, and zinc. Although the BLM is currently appropriate for use in deriving an updated  
29 freshwater copper CMC, further development is required before using it to evaluate a saltwater  
30 copper CMC, a CCC, or a chronic value.

31 Alternative approaches may be considered to establish the database of input parameters  
32 for the acute toxicity studies, because the approach described in this document is labor intensive.

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1 Alternatives may include either developing a few high-quality data sets that satisfy the minimum  
2 data requirements of the guidelines for a limited set of organisms or developing data sets for  
3 known sensitive species. Estimating missing input parameters by relying on statistical  
4 techniques or Monte Carlo approaches may also be explored.  
5

#### 6 **4.5.8. Bioaccumulation and Bioconcentration in Aquatic Organisms**

7 Bioaccumulation can be defined as the net accumulation of a metal in a tissue of interest  
8 or a whole organism that results from exposure from all relevant sources (e.g. water, food, and  
9 sediment). Metal bioaccumulation can apply to the entire organism, including both metal  
10 adsorbed to surfaces or absorbed by the organism or to specific tissue. It is usually expressed on  
11 a weight (dry or wet)- adjusted basis. Bioaccumulation that occurs under steady-state conditions  
12 (i.e., where accumulation remains relatively constant due to uptake being offset by elimination )  
13 is often of primary concern in risk assessments. Bioaccumulation of metals is a concern with  
14 respect to the accumulating organism when it occurs in the toxicological form(s) and  
15 concentrations at the site(s) of toxic action. It also can be indicative of dietary exposure to  
16 aquatic organisms at higher trophic levels. While inorganic metals can bioaccumulate in animal  
17 and plant tissues, biomagnification across three or more trophic levels is rare.

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1 The simplest tools for estimating  
2 bioaccumulation include direct measures of  
3 tissue levels and the derivation of simple  
4 relationships between tissue levels and  
5 environmental concentrations. These  
6 simple empirical relationships have often  
7 been expressed as bioaccumulation factors  
8 (BAF) or bioconcentration factors (BCF)  
9 and various data bases summarize values  
10 reported in the literature. It is well  
11 recognized, however, that there can be  
12 considerable uncertainty associated with  
13 the application of literature-derived,  
14 bioaccumulation or bioconcentration  
15 factors to specific risk assessment  
16 situations. In other cases, efforts have been  
17 made to establish more detailed  
18 mathematical relationships between  
19 exposure concentrations and tissue levels.  
20 For example, empirical approaches that  
21 extend beyond simple factors include the  
22 use of regression equations that describe  
23 the relationship between exposure and  
24 accumulation. Significant advances are  
25 also being made on the application of  
26 kinetic or steady-state uptake models for  
27 describing and predicting bioaccumulation  
28 (Reinfelder et al., 1998; Chang and  
29 Reinfelder, 2002; Kahle and Zauke, 2003;  
30 Wang and Zauke, 2004; ). These offer  
31 promise for understanding the contribution of differing routes of exposure (e.g., water vs. diet).  
32 There is considerable experience in using mechanistic bioaccumulation models for estimating

**Biomagnification** is defined as an increase in the concentration in an organism from a lower trophic level to a higher trophic level within the same food web due to bioaccumulation from the diet (also see trophic transfer). Biomagnification is expressed as the ratio of the concentration in the organisms of the higher trophic level to the concentration in the organisms of the lower trophic level (i.e., biomagnification factor). Note, however, that biomagnification that is quantified using field data makes an explicit assumption that bioaccumulation results from the diet only, when actually multiple sources may be involved (e.g., water or sediment). Biomagnification factors greater than one indicate biomagnification, while factors less than one indicate no biomagnification or biodilution (see below). Inorganic forms of metals rarely biomagnify across three or more trophic levels.

**Biodilution**, a decrease in organism concentration with increasing trophic level has generally been more commonly observed when concentrations are evaluated across three or more trophic levels.

**Trophic transfer.** The transfer of a bioaccumulated substance in a prey species to a predator species via dietary exposure. When the concentration in the predator species is increased relative to the prey, trophic transfer is a form of biomagnification. Trophic transfer is important due to its relationship to dietary toxicity (also called secondary poisoning) in which toxicity is manifest through accumulation in prey species and subsequent dietary exposure to predatory species. Importantly, however, dietary toxicity cannot be directly inferred from an evaluation of trophic transfer alone, but rather the combination of trophic transfer and toxicity information for predator/prey species within an ecologically-linked food web.

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1 risks of organic chemicals in aquatic systems and it is anticipated that such models will have an  
2 increasing role for risk assessments of metals.

3 Because the simpler empirical approaches such as the BCF and BAF have received much  
4 use and attention in the past, they are discussed in further below with respect to applications and  
5 limitations. However, the kinetic and steady-state uptake and bioaccumulation models will  
6 likely emerge as important approaches.

#### 7 8 **4.5.8.1. *Scientific Issues with BCF/BAF***

9 The BCF is the ratio of contaminant concentration in an organism to its concentration in  
10 water at steady state conditions and water-only exposures. Metal concentrations are usually  
11 expressed on a weight-adjusted, whole-organism basis, and waterborne metals may be expressed  
12 as total or dissolved metals. BCFs have been developed primarily with hydrophobic organic  
13 chemicals in aquatic systems, but similar such accumulation factors have been applied to other  
14 matrices (e.g., sediment and soils) for both organic chemicals and metals. Strictly speaking,  
15 metal bioconcentration in sediment and soil systems is the net accumulation of a metal in or on  
16 an organism from pore water only. Hence, in sediment and soil, the denominator for the BCF  
17 ratio should comprise the pore water concentration of metal, not the total metal concentration in  
18 the sediment or soil. In the broadest context, the BAF is the ratio of a contaminant concentration  
19 in an organism to that in a specified medium at steady state in situations where the organism is  
20 exposed to multiple environmental media.

21 Although BAFs and BCFs are calculated in a similar manner, the interpretation is slightly  
22 different with metal accumulation in organisms arising from water only for BCFs and from both  
23 water and dietary sources for BAFs. For aquatic organisms, BAFs are derived from  
24 measurements in natural environments, and BCFs are nearly always measured under laboratory  
25 conditions where exposure can be effectively limited to the water column. Unless metal  
26 concentrations in pore water serve as the denominator for the ratio, soil and sediment BAFs are  
27 usually termed BSAFs. Concentrations are usually measured on a total-metal and weight-  
28 adjusted whole-organism (or tissue) basis.

29 Toxicological bioaccumulation is the fraction of the metal that bioaccumulates and is  
30 distributed to receptors at sites of toxic action (Figure 2-2). For metals, this would include  
31 reactions with target proteins or other receptors that result in toxicity but not interactions with  
32 metallothionein and other metal-binding ligands or incorporation into granules that make metals

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1 unavailable for interactions with target molecules. This fraction is conceptual in nature and  
2 difficult to measure in practice; it is akin to the minimal effective dose measured in blood that is  
3 often used in medicine for assessing therapeutic effects. It could be conceptually defined as a  
4 toxicological bioaccumulation fraction or the ratio of total metal concentration in an organism to  
5 the metal concentration at the site(s) of toxic action.

6 As discussed in more detail in McGeer et al 2004, there are fundamental differences in  
7 the physical, chemical and toxicological properties of inorganic metal substances and organic  
8 substances, such that the BAF/BCF model would not apply to the former. The success of the  
9 BAF/BCF model as a valid indicator of the environmental and toxicological behavior of neutral  
10 organic substances is due to their hydrophobic/lipophilic chemical properties, and this has  
11 important consequences for application to inorganic metals. Many of the assumptions and  
12 characteristics of the BAF/BCF model openly conflict with the physical, chemical, biological,  
13 and toxicological realities associated with inorganic metal substances. For example, the  
14 BAF/BCF model works well for neutral organic substances because uptake of lipophilic  
15 substances into biota occurs via simple passive diffusion. However, uptake of the vast majority  
16 of inorganic metals is a physiological process, which occurs via a number of specific routes,  
17 most of which involve saturable transport kinetics. The degree of uptake and ultimate internal  
18 fate of inorganic metals is strongly influenced by ligand binding and receptor site competitive  
19 interactions which control metal availability and/or transfer processes, and cannot be reliably  
20 represented by a simple partitioning between water and organism. As a result of the differences  
21 explained below, simple application of these models is meaningless in assessing acute and  
22 chronic toxicity (adapted from McGeer et al., 2004):

- 23
- 24 • The principal theoretical features of the BAF/BCF model that make it applicable to  
25 neutral organic substances also make it inapplicable to inorganic metal substances.  
26 These factors produce an inverse relationship between BAF/BCF and exposure  
27 concentration; this has been observed for both essential metals and nonessential  
28 metals.
- 29
- 30 • The approach of using one simplified bioaccumulation model (BCF and BAF) and  
31 applying it to inorganic metals ignores the basic physical and chemical differences

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1 between organic and inorganic substances and is not supported by theoretical and  
2 empirical weight of evidence.

- 3
- 4 • Based on the inherent assumptions of the BCF and BAF model and on the  
5 environmental and toxicological behavior of the organic substances from which they  
6 were developed and validated, for the vast majority of inorganic metals evaluated, the  
7 scientific basis for broad application of the BAF/BCF model is lacking in the context  
8 of hazard assessment.
  - 9
  - 10 • The complexity of assessing and predicting the bioavailability and bioaccumulation  
11 of metals in aquatic systems arises from many factors including:
    - 12 - Essentiality of some metals resulting in a “U”-shaped dose response curve.
    - 13 - Variation in assimilation efficiency for different species of metal, for different  
14 biota and at different sites of uptake.
    - 15 - Ability to modulate uptake at the various sites of uptake.
    - 16 - Contributions of different routes of entry to the metal body burden and effects.
    - 17 - Ability to sequester, store, detoxify and eliminate bioaccumulated metals.
    - 18 - All metals will bioaccumulate to some degree without impacts as a result of  
19 exposure to natural background concentrations
  - 20
  - 21 • Based on reviews (e.g., McGeer et al., 2003), it would appear that for the vast  
22 majority of the metal/taxonomic group combinations assessed, the assumptions  
23 regarding the independence of BCF/BAF with exposure concentration and  
24 proportionality of hazard with increasing BCF/BAF do not hold true.
  - 25
  - 26 • The latest scientific data on bioaccumulation do not currently support the use of BCF  
27 and BAF data when applied as generic threshold criteria for the hazard potential of  
28 inorganic metals (e.g., for classification as a “PBT” chemical).
  - 29
  - 30 • In cases where the use of BCF/BAF data are being considered, a careful evaluation of  
31 existing data for metals that formally documents the extent to which values represent  
32 bioaccumulation linked to toxic impact (both direct and indirect) and the resulting

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1 uncertainty introduced by concentration dependency and other aspects of the  
2 BCF/BAF data should be done.

- 3
- 4 • Moving from the current situation to a revised, improved, and validated set of criteria  
5 and/or methodologies for assessing metal hazards and risks presents a number of  
6 options and challenges, as currently there would appear to be no clear alternatives  
7 ready for application.
- 8
- 9 • BCFs for metals will not be absolute values, but will vary widely with both the  
10 specific exposure circumstances/conditions as well as the status/age/condition of the  
11 particular organism measured. This applies to all substances, organic and inorganic,  
12 although the relative scale of the uncertainty added is greater for inorganic  
13 substances.

14 The Organization for Economic Co-operation and Development (OECD) has published  
15 guidance for the hazard classification of chemical substances (OECD, 2001). The hazard  
16 classification schemes presented in the OECD guidance incorporates, among other parameters,  
17 evidence of bioaccumulation as a basis for hazard ranking. The OECD guidance recognizes the  
18 shortcomings associated with the use of BAF/BCF data as a surrogate for the hazard potential of  
19 metals, and therefore cautions that metals should be assessed on a case-by-case basis rather than  
20 recommending the application of the simple bioaccumulation model (BAF/BCF). The principle  
21 features of the BAF/BCF model that make it applicable as a surrogate for acute and chronic  
22 toxicity to neutral organic substances also make it inapplicable to inorganic metal substances as  
23 discussed above and in McGeer et al. (2004).

24 The phenomena described above regarding the limitations of the BCF/BAF model also  
25 have a bearing on the use of BCFs and BAFs in national-level assessments (e.g., water quality  
26 criteria and national-scale risk assessments) in addition to site-specific assessments. For  
27 example, variability in BCFs and BAFs caused by an inverse relationship of BCF with exposure  
28 concentration can confound the application of BCFs in risk assessments, the extent to which  
29 depends in part on the magnitude of this relationship (i.e., its slope) and the extent of  
30 extrapolations made (i.e., extrapolation of BCFs across differing exposure concentrations and  
31 species). For organic chemicals, generic normalizing factors have been shown to reduce  
32 variance in BAFs due to bioavailability differences (e.g., normalizing to organic carbon and

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1 lipid; see Burkhard et al., 2003). However, analogous normalizing factors have yet to be  
2 developed and widely applied in the context of BAFs or BCFs for metals, although some  
3 progress has been made with relating accumulation parameters to body weight (Hendriks and  
4 Heikens, 2001) or permeable surface area in the case of the amphipod, *Gammarus zaddachi*  
5 (Wang and Zauke, 2004). Simple normalizing factors can be confounded by the fact that  
6 animals and plants have evolved physiological and anatomical means for regulating and or  
7 storing internal concentrations of metals and the species-dependent specificity of these  
8 mechanisms. Regulation of metals by biota can actually result in decreased apparent BAF/BCF  
9 factors with increasing exposure concentrations. Thus, a fundamental understanding of such  
10 mechanisms is important for reducing the uncertainties associated with interpreting tissue  
11 residues. The mere presence of a metal in a plant or animal does not mean it is at a site of toxic  
12 action. In summary, caution should be exercised when using published BAF and BCF values to  
13 make judgements about resultant tissue levels of metals in animals and plants. Where this is  
14 found to be necessary as an initial, screening level of assessment, the associated uncertainties  
15 should be identified and discussed with respect to their potential impact on the risk estimates and  
16 resultant decisions.

17 In situations where a decision has been made to employ a BAF or BCF factor, there may  
18 ways to reduce some of the uncertainty. One idea involved subtracting “normal” accumulation  
19 from the calculation of the BCF. This involves separating the portion of metal that  
20 bioaccumulates from exposure under “normal” or background conditions from that portion that  
21 occurs as a result of exposure to elevated levels of metals (McGeer et al., 2003). The ACF value  
22 also accounts for the accumulation of essential metals required for physiological function. These  
23 ACF values were dramatically lower than BCF values (illustrating the importance of normal  
24 bioaccumulation) for some metals, particularly essential metals. However, for most metals  
25 ACFs still varied with exposure concentration and were not viewed as a sufficient replacement to  
26 the BCF (McGeer et al., 2003).

27 Another approach that may reduce uncertainty caused by concentration dependency of  
28 BCFs and BAFs involves limiting the selection and application of BCF and BAF values to  
29 concentration ranges that more closely correspond to toxicological thresholds. For example,  
30 limiting the calculation of BAF and BCF values to concentrations that approximate the  
31 applicable water quality criterion has also been suggested as a method for reducing the  
32 uncertainty around BCF and BAF values in situations where concentration dependency is

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1 evident. This would account for bioaccumulation at an exposure level where concern over  
2 bioaccumulation might be expected. This approach has limitations (McGeer et al., 2004) and  
3 does not appear to reduce the overall variability associated with BCF and BAF measurements  
4 when evaluated across broad classes of organisms. An additional issue for this approach is that  
5 WQC reflect some of the more sensitive organisms, whereas the BCF and BAF measurements  
6 are not necessarily from these same organisms and include data from biota that may not be as  
7 sensitive to chronic effects. Therefore, as a modifier for broad-based application, this variation  
8 of the BCF/BAF methodology does not appear to explain variability. However, on a site-  
9 specific basis where toxicity thresholds and species are better characterized, this approach may  
10 have value in reducing uncertainty. As already noted, caution should be exercised with any of  
11 the BAF or BCF applications.

#### 12 13 **4.5.8.2. Bioaccumulation in Relation to Dietary Toxicity**

14 Discriminating between metals that have the potential to cause effects via trophic transfer  
15 and metals that do not is another approach that might be useful in distinguishing between metals,  
16 based on bioaccumulation and impacts. Metals taken up and stored within an organism may not  
17 cause direct effect to that organism but may be bioavailable to organisms in the next trophic  
18 level that feed on it. Bioaccumulation of metals in prey organisms may be quite high, especially  
19 in organisms such as high volume filter feeders and those that accumulate elevated levels of  
20 metals, for example, those that store detoxified forms. Metals that bioaccumulate to levels in  
21 prey organisms that cause impacts in predatory organisms are clearly important issues to address  
22 however, there is a general lack of understanding of the potential for toxicity.

23 A prey organism with a high concentration of a particular metal represents a potential  
24 opportunity for the trophic transfer of the metal from an enriched source to a predator at the next  
25 trophic level. The form of detoxified storage of that accumulated metal in the prey species has a  
26 significant effect on the potential assimilation of that metal by the predator (Wang and Fisher,  
27 1999). For example, Nott and Nicolaidou (1990) have shown that the bioavailability to  
28 neogastropod mollusc predators of metals present in detoxified metalliferous granules in prey  
29 varies among metals and with type of granule; thus the zinc-rich pyrophosphate granules  
30 accumulated in barnacles are not digested in the digestive tract of the predator *Nucella lapillus*  
31 and are therefore not bioavailable to that predator. Similarly the physico-chemical form of  
32 accumulated cadmium in the oligochaete *Limnodrilus hoffmeisteri* is critical in the assimilation

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1 of cadmium by a predator, in this case the decapod *Palaemonetes pugio* (Wallace et al., 1998;  
2 Wallace and Lopez, 1997). Other studies have indicated that lead bioaccumulated in mussels  
3 and stored in a detoxified form within the soft tissues of the mussels is bioavailable and may  
4 cause impacts (Regoli and Orlando, 1994). The potential effects of food preparation (cooking)  
5 on metal bioavailability should be considered when assessing risks to human consumers.

6 Currently there are no standardized tools for incorporating the potential impact of dietary  
7 exposure into assessments. Research efforts are focusing on the relative importance of: dietary  
8 pathways in relation to waterborne exposures, transfer from sediments into food chains, the  
9 potential of bioaccumulated to cause impacts in consumer organisms. One example of an  
10 approach that illustrates how exposure, bioaccumulation and trophic transfer can be linked is  
11 illustrated below.

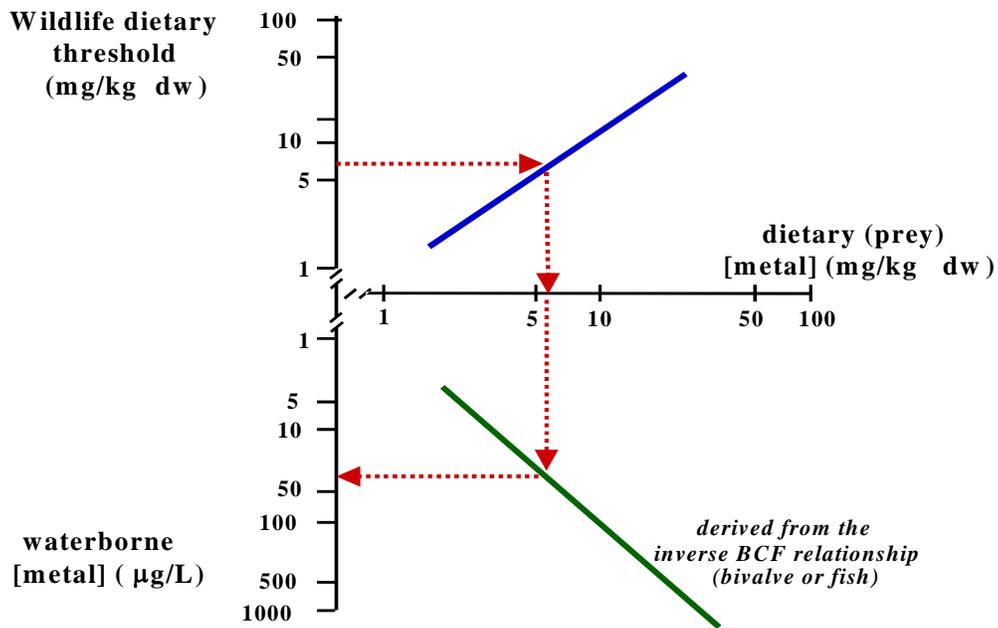
12 A general approach to account for potential impacts arising from trophic transfer would  
13 be to link bioaccumulation in prey items to exposure in the water column as well as potential  
14 impacts in consumers. One methodology to achieve this is to integrate tissue burden to toxicity  
15 relationships (prey: predator interactions) with exposure to bioaccumulation relationships (e.g.,  
16 BCF and BAF values). A theoretical example of this approach, developed by Brix et al.  
17 ([www.epa.gov/ncea/raf/pdfs/metals/sumryrprt\\_metals.pdf](http://www.epa.gov/ncea/raf/pdfs/metals/sumryrprt_metals.pdf)) is shown in Figure 4-9.

18 Within the context of this approach, it would be necessary for relevance to ensure that the  
19 species being considered were linked within trophic food webs (i.e., the exposure to  
20 bioaccumulation relationship was for prey items being consumed by the predators that are  
21 sensitive to trophic transfer). Some data already exists to begin these evaluations. For example,  
22 it is possible to derive the water concentrations necessary to produce impacts via dietary  
23 exposure (see Brix et al. reference above) and site-specific case studies would be valuable in  
24 illustrating, testing, and validating these relationships. The potential benefits from this approach  
25 would be reducing the uncertainty from extrapolations across exposure concentrations currently  
26 being made with metals BCF/BAF data. Also, there would be an ability to link impacts through  
27 to waterborne metal concentrations. However, particularly if BCF and BAF data is used, this  
28 approach would bring with it the inherent uncertainty associated with predicting tissue metals  
29 burdens (e.g., the high variability) and the inability to account for geochemical influences on  
30 uptake and accumulation. Ensuring that the local site specific factors that influence the  
31 bioaccumulation and the expression of impacts are understood and recognized would be  
32 important, to ensure for example that criteria and guideline values were not set at existing

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1 background levels. Two regression relationships, one for exposure and bioaccumulation (green  
2 line) and the other for bioaccumulation to dietary toxicity thresholds (blue line) are used to line  
3 exposure and dietary impacts.



22 **Figure 4-9. Linkages between dietary toxicity threshold, bioaccumulation in prey**  
23 **organisms and waterborne exposure.**

24  
25  
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1 **4.5.8.3. Alternatives to Tissue Burdens and Bioaccumulation**

2 A key parameter that a bioaccumulation measure should be validated against is chronic  
3 toxicity. Because bioaccumulation criteria within the context of persistent, bioaccumulative,  
4 toxic substances are used as indicators of chronic toxicity (Franke et al., 1994; OECD, 2001),  
5 validation of linkages to chronic metal toxicity would provide confidence in their use and  
6 application. A number of key issues should be addressed when considering bioaccumulation of  
7 metals in relation to the potential for chronic impacts, and these add uncertainty to the  
8 interpretation of data. However, unlike the substances that the PBT concept was originally  
9 developed for, there is often substantial information on the chronic toxicity of metals.

10 In some regards, our ability to understand and interpret chronic metal toxicity is as advanced,  
11 or possibly more advanced, than metal bioaccumulation. Therefore, rather than trying to derive  
12 and validate a surrogate for the chronic impacts of metals, it might, in some cases, be feasible to  
13 eliminate bioaccumulation and only consider chronic toxicity data. The development of a  
14 criterion based on chronic toxicity could be based on a variety of approaches, all of which will  
15 require a modified framework for consideration as BCFs, BAFs, and bioaccumulation would be  
16 replaced. Despite the challenges associated with this degree of change, it is worthy of  
17 consideration.

18  
19 **4.5.9. Bioaccumulation in Terrestrial Organisms**

20 For terrestrial ecosystems, the concept of bioaccumulation is intended to capture the potential  
21 for two ecologically important outcomes: (1) direct toxicity to plants and wildlife and (2)  
22 secondary toxicity to animals feeding on contaminated plants and animals. This approach  
23 captures the potential for trophic transfer of metals through the food web, so total exposure can  
24 be calculated, including dietary intake as well as intake from contaminated environmental media  
25 (soil and water). For vegetation, the BAF (or BSAF) is defined as field measurements of metal  
26 concentration in plant tissues divided by metal concentration in soil (or soil solution); the BCF is  
27 defined as the same measurement carried out in the laboratory (Smolders et al., 2003).

28 Data applicability is directly related to which tissue is sampled and how it is processed.  
29 BAFs for plants include metals aerially deposited on leaves as well as those in soil particles  
30 adhering to roots. Such metals will not be part of BCFs, which frequently are determined in  
31 hydroponic culture. Similar differences between BCFs and BAFs apply for earthworms exposed  
32 in laboratory studies using the filter paper substrate protocols. Furthermore, BCFs with

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1 earthworms may not include additional feeding of the animals during the study. Additionally,  
2 field studies are reflective of chronic exposures, whereas BCFs may be calculated from shorter  
3 time frames. For birds and mammals, whole-body BAFs generally are not calculated, except for  
4 small mammals such as rodents (Sample et al., 1998b). Rather, concentrations in target tissues  
5 are measured for comparison with critically toxic levels (Beyer et al., 1996).

6 The BAFs for metals by soil invertebrates and most plants are typically less than 1, although  
7 they usually are based on the total metal in soil and tissue. BSAFs expressed in this manner may  
8 be suitable for comparisons of metal uptake within the same soil type, but they would be  
9 misleading if soil bioavailability factors (e.g., pH and organic carbon) differ. Unfortunately, the  
10 literature database is populated almost entirely with BAFs derived from measurements of total  
11 metal. Furthermore, bioaccumulation is not a simple linear relationship. Uptake is nonlinear,  
12 increasing at a decreasing rate, as medium concentration increases. Reliance on incorrect  
13 relationships of the bioavailable portion will be trivial compared to the error associated with  
14 BAFs at high concentrations. In the future, a ratio of total metal in the organism to some  
15 measure of the bioavailable fraction of metal in the soil (e.g., free ion concentration or weak salt  
16 extractable) should be used for expressing a BSAF that allows comparison among different soils.  
17 An alternative approach currently under study is to use a multivariate statistical model to look  
18 for patterns of uptake of multiple metals to predict the potential bioconcentration of one metal of  
19 particular interest (Scott-Fordsmand and Odegard, 2002).

#### 21 **4.5.9.1. Models for Bioaccumulation in Soil Invertebrates**

##### 22 **4.5.9.1.1. Application**

23 **4.5.9.1.1.1. *Univariate models.*** The bioaccumulation of metals in soil organisms cannot  
24 reasonably be modeled from information based  
25 solely on soil concentrations. Therefore, models for  
26 the prediction of metal bioaccumulation by soil  
27 invertebrates are primarily empirical in nature,  
28 describing relationships between metal body burdens  
29 in oligochaetes and collembola, soil metal  
30 concentrations, and soil physical/chemical  
31 characteristics. Statistical relationships have been  
32 established using univariate and multiple regression

The bioaccumulation of metals in soil organisms cannot be modeled from information based solely on soil concentrations. Two types of models are available to help describe the relationships between metals body burden and soil parameters: (1) *univariate models* that describe statistical relationships between body burdens, soil metal concentrations, and soil physical and chemical properties; and (2) *multivariate models* that explain BAFs as a function of soil properties.

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1 approaches. Sample et al. (1998a) and Peijnenburg et al. (1999b) have each developed  
2 univariate uptake models for earthworms that are based on empirical data (metal concentrations  
3 in worms vs. the natural log of amount of metal in soils) that are widely used as a first  
4 approximation. However, these models are not specific to soil type and therefore do not account  
5 for bioavailability factors such as pH, clay content, or cation exchange. Furthermore, they do  
6 not adequately predict Cr or Ni uptake.

7  
8 **4.5.9.1.1.2. *Multivariate models.*** Multivariate models also are available (Peijnenburg et al.,  
9 1999a, b) for *Eisenia andrei* and *Enchytraeus crypticus* that explain BAF as a function of soil  
10 characteristics. The soil parameters that generally contributed the most to explaining the  
11 variance between uptake rate constants and BAFs were pH (for Cd, Zn) and also CEC (for Pb)  
12 and clay content (for Cd). Similar studies are needed for describing relationships between soil  
13 physical/chemical characteristics and metal bioaccumulation in other groups of soil invertebrates  
14 such as collembola and isopoda. Until these are available, the models for earthworms and  
15 enchytrids can be applied to other groups, although the added uncertainty should be  
16 acknowledged.

17 Path analysis has been suggested as an alternative for multiple regression in describing these  
18 relationships. It partitions simple correlations into direct and indirect effects, providing a  
19 numerical value for each direct and indirect effect and indicates the relative strength of that  
20 correlation or causal influence (Basta et al., 1993). Bradham (2002) used path analysis and  
21 backwards stepwise regression analysis to derive statistical models capable of predicting uptake  
22 and effects of As, Cd, Pb, and Zn in earthworms as a function of soil properties.

23 Saxe et al. (2001) described a model for predicting whole-body concentrations of Cd, Cu, Pb,  
24 and Zn in *Eisenia andrei* as a function of pH, soluble metals in the soil at gut and environmental  
25 pH, and soluble organic carbon in soil extracts. The model also includes parameters that  
26 characterize the ability of worms to regulate the metal body burden, whether metal uptake is via  
27 the epidermal or gut surface and whether the metal is essential. The model has been validated  
28 against a series of Dutch soils and is very good at correctly predicting metal accumulation.

29  
30 **4.5.9.1.2. *Limitations.*** Bioaccumulation of organic substances is typically modeled using a  
31 One-Compartment, First-Order Kinetics (1CFOK) model. However, most of the assumptions of  
32 the model are violated when applied to bioaccumulation of metals by soil invertebrates. Soil

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1 invertebrates are not exposed to a constant concentration of metals in the soil over space and  
2 time, making it difficult to accurately define exposure. Sufficient data exist on the metabolism  
3 of metals to show that all pools of metal taken up by the soil invertebrates are not equally  
4 available for depuration (some are actually never depurated and are released only when the  
5 organism dies), making a 1CFOK model an inaccurate approximation.

6 Metal concentrations do not reach a steady state that is proportional to external (i.e., soil)  
7 concentrations. This is similar to the situation in aquatic systems where metal bioaccumulation  
8 is a function of water concentration. Internal essential metal concentrations are regulated and  
9 remain relatively constant over a wide range of soil metal concentrations. Only when normal  
10 regulatory mechanisms are overwhelmed do internal levels of essential metals increase.  
11 Accumulation of nonessential metals also violates the assumption of steady state, as organisms  
12 have evolved mechanisms for the detoxification of nonessential metals that involve the internal  
13 accumulation of the metal in forms that are not toxic to the organism (e.g., incorporation into  
14 inorganic granules or binding to organic molecules to form metal ligands such as  
15 metallothioneins).

#### 16 17 **4.5.9.2. Critical Body Residues**

18 CBRs are an extension of the concept of  
19 bioaccumulation to internal concentrations of  
20 metals that are correlated with some toxic response  
21 and hence represent toxicological bioavailability  
22 (Conder et al., 2002; Lanno et al., 1998). Use of  
23 CBRs in appropriate species may reduce  
24 uncertainties in ecological risk assessment  
25 procedures (Van Straalen, 1996; Van Wensem et  
26 al., 1994). However, only a few CBRs have been  
27 developed in soil invertebrates for metals.  
28 Crommentuijn et al. (1997, 1994) and Smit (1997)  
29 established CBRs for sublethal effects for Cd and  
30 Zn, respectively, in the springtail (*Folsomia*  
31 *candida*). Conder et al. (2002) demonstrated that  
32 effects of Cd in earthworms (*Eisenia fetida*) are

Knowing the species of metal that is likely to be present in the environment is important for extrapolating results from laboratory tests to natural settings. Laboratory studies are often done under simplified environmental conditions (compared to the natural environment) that may enhance aquatic bioavailability. Conversely, these studies frequently do not include potential effects of dietary metal exposure.

In addition to abiotic factors that can influence metal bioavailability, other variables may potentially affect the outcome of laboratory toxicity assays and should be considered in interpretation of data. These include acclimation of test animals to culture conditions, natural background concentrations of the metal (including, but not limited to the metal of interest), potential interactions of the various metals, and potential transformations to bioreactive species. The similarity of the pretest culture medium to the underlying testing medium also should be considered.

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1 correlated with concentrations of the metal in the solid phase of the worms (i.e., the pellet  
2 fraction, following homogenization and centrifugation). If future research can isolate the fraction  
3 of an invertebrate that represents toxicological bioavailability, then it may be possible to  
4 estimate a toxicological BSAF representing a relationship between a specific fraction of metal  
5 that accumulates in the organism and a measure of chemical bioavailability in the soil. Until  
6 then, CBRs based on whole-organism analyses are a reasonable approximation for use in  
7 ecological risk assessments.

#### 8 9 **4.5.10. Sediment Toxicity - Equilibrium Partitioning Approach for Metals**

##### 10 **4.5.10.1. Rationale for Use of EqP Benchmarks**

11 Toxic pollutants in bottom sediments of the nation's lakes, rivers, wetlands, estuaries,  
12 and marine coastal waters create potential for continued environmental degradation even where  
13 water column concentrations comply with established human health and aquatic life WQC. In  
14 addition, contaminated sediments can be a significant pollutant source that may cause water  
15 quality degradation to persist even when other pollutant sources are stopped (U.S. EPA  
16 1997d,e,f; Larsson, 1985, Salomons et al., 1987; Burgess and Scott, 1992).

17 Because of their widespread use and associated environmental releases, metals such as  
18 cadmium, copper, lead, nickel, silver, and zinc are commonly elevated in aquatic sediments.  
19 Various types of sediment guidelines have been proposed for assessing the potential effects of  
20 these metals on benthic invertebrate communities. Many of these involve empirical correlations  
21 of metal concentrations in sediment with associated biological effects (e.g., sediment toxicity);  
22 these include ER-M, ER-L, TEL and PEL, SLC, AET and others (Sullivan et al., 1985; Persaud  
23 et al., 1989; Long and Morgan, 1990; Ingersoll et al., 1996; MacDonald et al., 1996). Most of  
24 these approaches use measures of total metal in sediment, and do not account for differences in  
25 bioavailability of metals among sediments. Nevertheless, when examined across large data sets,  
26 these empirical guidelines do show relationships between concentrations of total metal and  
27 biological effects. However, a limitation to these empirical approaches is that the causal linkage  
28 between the measured concentration of metals and the observed toxicity cannot be established, in  
29 part because of the procedures used to derive correlative values and because values derived are  
30 based on total rather than bioavailable metal concentrations. That is, for any given total metal  
31 concentration, adverse toxicological effects may or may not occur, depending on the

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1 physicochemical characteristics of the sediment of concern (Tessier and Campbell, 1987;  
2 Luoma, 1989; Di Toro et al., 1990).

3 An alternative to empirical approaches is equilibrium partitioning (EqP) theory. The EqP  
4 approach is intended to predict concentrations that will or will not cause adverse effects based on  
5 an understanding of the factors that control metal bioavailability in sediments and the  
6 relationship between that bioavailability and biological effects. For cationic metals (e.g.,  
7 cadmium, copper, lead, nickel, silver, and zinc), these factors include the presence of acid  
8 volatile sulfides (AVS), which form insoluble metal sulfides that are believed to have low  
9 biological availability. Beyond AVS, particulate organic carbon (POC) and iron and manganese  
10 hydroxides in sediment, and dissolved organic matter (DOM) in interstitial water are also  
11 believed to influence metal bioavailability in sediment. By recognizing differences in  
12 bioavailability among sediments, the intent of the EqP approach is to produce guideline values  
13 that are applicable across a wide variety of sediments and represent causal relationships between  
14 specific chemicals and sediment toxicity.

15 In 1987, EPA reviewed proposed approaches to developing numerical sediment quality  
16 guidelines (Chapman, 1987). All of the approaches reviewed had strengths and weaknesses, and  
17 no single approach was found to be applicable for the derivation of sediment benchmarks in all  
18 situations (U.S. EPA, 1989d). The EqP approach was selected for further development because  
19 it presented the greatest promise for generating defensible national chemical-specific sediment  
20 benchmarks applicable across a broad range of sediment types. While the Agency has never  
21 adopted formal sediment quality criteria, the technical development of sediment quality  
22 guidelines was pursued and has resulted in the publication of several numerical guidelines (See  
23 <http://www.epa.gov/nheerl/publications/>). These guidelines are called “EqP sediment  
24 benchmarks” (ESBs) and are numerical concentrations for individual chemicals or mixtures of  
25 chemicals derived using the EqP approach. As described below, these are intended to be  
26 concentrations below which there should not be direct toxicity to most benthic organisms as a  
27 result of the chemical or chemicals addressed in the guideline.

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1 **4.5.10.2. Application of the EqP Approach**

2 As originally proposed, the EqP approach for copper, cadmium, lead, zinc, nickel, and  
3 silver focused on AVS as the principal partitioning phase. Because these metals form sulfides  
4 that are highly insoluble, toxicity from these metals is not expected if there is sufficient sulfide  
5 available to bind all available metal. Because some metals in sediment are present in mineral  
6 forms that are not highly reactive, the metal concentrations to which the sulfide concentration is  
7 compared is not the total metal in sediment, but the metal extracted simultaneously with the  
8 sulfide, and is therefore referred to as simultaneously extracted metal (SEM). Because cadmium,  
9 copper, lead, nickel, and zinc are divalent metals, 1 mol of each metal can bind with 1 mol of  
10 AVS. The molar concentrations of these metals are compared with AVS on a one-to-one basis.  
11 Silver, however, exists predominantly as a monovalent metal, so that silver monosulfide (Ag<sub>2</sub>S)  
12 binds 2 mols of silver for each mol of AVS. Therefore, SEM Ag will represent the molar  
13 concentration of silver divided by two, [Ag]/2, which is compared with the molar AVS  
14 concentration. Thus, the solid-phase AVS ESB is defined as:

15  
16 
$$\sum_i [\text{SEM}_i] \leq [\text{AVS}]$$

17 where:

18  
19 
$$\sum_i [\text{SEM}_i] = [\text{SEM}_{\text{Cd}}] + [\text{SEM}_{\text{Cu}}] + [\text{SEM}_{\text{Pb}}] + [\text{SEM}_{\text{Ni}}] + [\text{SEM}_{\text{Zn}}] + 1/2[\text{SEM}_{\text{Ag}}]$$

20  
21 When this sum is less than the molar concentration of AVS, no toxicity is expected  
22 because there is sufficient sulfide to bind all SEM. Results of calculations using chemical  
23 equilibrium models indicate that metals act in a competitive manner when binding to AVS. That  
24 is, the six metals will bind with AVS to form their respective sulfides in the order of their  
25 increasing solubility: silver, copper, lead, cadmium, zinc, and nickel. Therefore, they should be  
26 considered together. There cannot be a guideline just for nickel, for example, because all the  
27 other metals may be present as metal sulfides and therefore, to some extent, as AVSs. If these  
28 other metals are not measured as a mixture, then the  $\sum \text{SEM}$  will be misleadingly small, and it  
29 might appear that  $\sum [\text{SEM}] < [\text{AVS}]$  when in fact this would not be true if all the metals were  
30 considered together. It should be noted that EPA currently restricts this discussion to the six  
31 metals listed above; however, in situations where other sulfide-forming metals (e.g., mercury)  
32 are present at high concentrations, they also should be considered.

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1 While the SEM-AVS comparison has been found effective for predicting when metals in  
 2 sediment will not cause toxicity, the presence of excess SEM above AVS does not always  
 3 portend toxicity. This is, at least in part, because other components of sediments can also reduce  
 4 the bioavailability and toxicity of metals. To address these additional factors, a second  
 5 component of the metal mixture ESB was developed: the interstitial water benchmark. The  
 6 concept behind the interstitial water component is that even if there is excess SEM above AVS in  
 7 a sediment, if there is sufficient additional binding available to maintain non-toxic  
 8 concentrations of metals in the interstitial water, then the sediment should not be toxic, despite  
 9 the excess SEM. The aggregate toxicity of metals in the interstitial water is assessed by dividing  
 10 the concentration of dissolved metal in interstitial water by the final chronic value from the EPA  
 11 water quality criterion for the protection of aquatic life. This ratio is called the Interstitial Water  
 12 Benchmark Unit (IWBU), and these are summed across all of the metals (using an assumption of  
 13 additivity, as described in Section 4.5.3); if the sum is less than 1, then the interstitial water-phase  
 14 ESB is met, and toxicity is not expected. Mathematically, this equates to:

$$\sum [M_{i,d}] / [FCV_{i,d}] \leq 1$$

15  
 16 where:

$$\sum_i ([M_{i,d}] / FCV_{i,d}) = \sum_i ([M_{Cd,d}] / FCV_{Cd,d}) + \sum_i ([M_{Cu,d}] / FCV_{Cu,d}) + \sum_i ([M_{Pb,d}] / FCV_{Pb,d}) +$$

$$\sum_i ([M_{Ni,d}] / FCV_{Ni,d}) + \sum_i ([M_{Zn,d}] / FCV_{Zn,d})$$

17  
 18  
 19  
 20  
 21 For freshwater sediments, the FCVs are hardness dependent for all of the divalent metals  
 22 under consideration, and thus should be adjusted to the hardness of the interstitial water of the  
 23 sediment being considered. Because there are no FCVs for silver in freshwater or saltwater, this  
 24 approach is not applicable to sediments containing significant concentrations of silver (i.e.,  
 25  $\Sigma SEM > AVS$ ). Because silver has the smallest solubility product and the greatest affinity for  
 26 AVS, it would be the last metal to be released from the AVS or the first metal to bind with AVS.  
 27 Therefore, it is unlikely that silver would occur in the interstitial water of any sediment with  
 28 measurable AVS (Berry et al., 1996).

29 When used together, the overall metal mixture ESB is met if either the SEM-AVS or  
 30 interstitial water ESBs are met. Further, both guidelines are no-effect guidelines, in that they  
 31 predict the absence of toxicity if they are met. Though the probability of toxicity is higher if  
 32 both components of the ESB are exceeded, and this probability increases as the magnitude of

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1 exceedance increases, failing to meet the ESB does not imply that toxicity will necessarily occur.  
2 This is because neither the SEM-AVS or interstitial water components address all factors that  
3 may reduce the bioavailability and toxicity of metals in sediment. Site-specific toxicity testing  
4 may be advisable to refine assessments for specific sites.  
5

#### 6 **4.5.10.3. *Enhancements to the Metals ESB***

7 The AVS guideline is a “no effects” guideline; that is to say that if the guideline is not  
8 exceeded, the sediments should not be toxic due to the metals included in the guideline, but an  
9 exceedance of the guideline does not necessarily mean that the sediments will be toxic due to the  
10 presence of those metals. One way to reduce the uncertainty of a prediction of toxicity is to  
11 normalize for the fraction of organic carbon in the sediment and use  $\sum \text{SEM-AVS} / f_{\text{OC}}$  for the  
12 solid-phase guideline. The use of this guideline is described in the draft metals ESB (EPA  
13 2002f).

14 A refinement of this organic carbon correction has been proposed by Di Toro et al. (2004,  
15 submitted). In this analysis, organic carbon partition coefficients are calculated for each metal  
16 and used to estimate combined AVS-organic carbon partitioning. Although this approach has  
17 not been widely applied as yet, it has a strong theoretical basis, and initial applications suggest  
18 that it can strengthen the ability of EqP to predict metal toxicity (rather than just non-toxicity) in  
19 sediments.

20 The metals ESB currently applies to only six metals: cadmium, copper, lead, nickel,  
21 silver, and zinc. However, AVS can also be used to predict the lack of toxicity in sediments due  
22 to chromium because the presence of AVS in sediments is indicative of reduced sediments.  
23 Chromium is present primarily in its reduced form (CrIII) in reduced sediments, and in this form  
24 it is much less toxic and bioavailable than the oxidized form of chromium (CrVI) (Berry et al., in  
25 prep.). The use of AVS in the prediction of the lack of chromium toxicity in sediments will be  
26 described in an addendum to the metals ESB.  
27

#### 28 **4.5.10.4. *Endpoints of Concern***

29 The metals ESB should be viewed as an approach, as opposed to a specific number. The  
30 ESB calculates a concentration in sediment that will correspond to a given toxicological  
31 endpoint using partitioning theory. However, the correspondence of the benchmark is variable,  
32 depending on which toxicological endpoints are selected. The FCVs from the metals WQC are

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1 used in the ESB as examples because they are familiar and provide a widely accepted level of  
2 protection. Other toxicological endpoints could be chosen, however, if a different level of  
3 protection was desired. For example, the water-only, chronic no-effect concentration for a  
4 threatened or endangered species might be used as the toxicological endpoint. EqP theory could  
5 then be used to calculate a sediment concentration that would be protective of that species.

#### 6 7 **4.5.10.5. Limitations**

8 It should be emphasized that the metals ESBs are intended to protect benthic organisms  
9 from the direct effects of these *six metals* in sediments that are permanently inundated with  
10 water, intertidal, or inundated periodically for durations sufficient to permit development of  
11 benthic assemblages. They do not apply to occasionally inundated soils containing terrestrial  
12 organisms. These benchmarks do not address the possibility of bioaccumulation and transfer to  
13 upper-trophic-level organisms or the synergistic, additive, or antagonistic effects of other  
14 substances.

15 The EqP approach to metal assessment has been questioned on several technical issues,  
16 such as whether it applies to organisms that live in oxygenated burrows, whether it adequately  
17 accounts for ingestion of sediment, and whether it is appropriate to use bulk sediment chemistry  
18 to represent responses to microenvironments that exist in bedded sediment. Many of these issues  
19 are difficult to address in a comprehensive manner, and more study will be required to fully  
20 evaluate them. In the interim, it should be noted that there is a considerable body of  
21 experimental data supporting the EqP approach to metals (Berry et al. 1996; Hansen et al. 1996;  
22 Di Toro et al. 2004; EPA 2002f), and these data include tests with organisms with varying life  
23 histories, that use irrigated burrows, and that ingest sediment. While this does not prove that the  
24 criticisms above are not legitimate, it provides support for continuing to apply the EqP approach  
25 as a reasonable representation of metal toxicity in sediment, at least until a superior approach is  
26 developed.

27 Because AVS is a product of microbial activity, AVS can vary seasonally with changes  
28 in microbial activity. The degree to which such cycling can affect the potential risk from metals  
29 in sediments is unclear but should be considered in sampling programs designed to assess SEM-  
30 AVS.

31 Because metals bound to sulfide do not appear to be sufficiently bioavailable to cause  
32 toxicity in sediment toxicity tests, one would not expect bioaccumulation of metals in sediments

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1 with more AVS than SEM. However, several studies have found bioaccumulation of metals  
2 even when excess AVS is present (Ankley et al., 1996). This has caused considerable debate  
3 about the appropriateness of SEM-AVS for assessing metal toxicity in sediments (e.g., Lee et al.,  
4 2000), because it suggests that metal bioavailability may not be effectively represented by SEM-  
5 AVS analysis. However, there is a large number of studies indicating that toxic effects of metals  
6 are absent in sediments when SEM is less than AVSs, even when bioaccumulation is observed.  
7 This suggests that the bioaccumulated metals may not be toxicologically available or of  
8 sufficient concentration in the organism to cause effects. In addition, these metals do not  
9 biomagnify to higher trophic levels in aquatic ecosystems (Suedel et al., 1994). Therefore, an  
10 ESB based on the difference between the concentrations of SEM and AVS still appears  
11 appropriate for protecting benthic organisms from the direct effects of sediment-associated  
12 metals, but not for protecting against metal bioaccumulation.

#### 14 **4.5.10.6. Use and Implementation**

15 In practice, the sediment benchmarks for these six metals are not exceeded, and benthic  
16 organisms are sufficiently protected (defined in this case as the level of protection afforded by  
17 the WQC), if the sediment meets either one of the following benchmarks:

- 18 • The solid phase benchmark:  $\sum_i[\text{SEM}_i] < [\text{AVS}]$ ; in other words, if metal, measured  
19 as SEM, does not exceed AVS; or
- 20 • The interstitial water benchmark:  $\sum_i([\text{M}_{i,d}]/\text{FCV}_{i,d}) < 1$ ; in other words, if the sum of  
21 the contributions of all six metals in the interstitial water would not be expected to  
22 cause chronic effects at the level of protection afforded by the WQC.

23 If the AVS or interstitial water ESBs is exceeded, there is reason to believe that the  
24 sediment might be unacceptably contaminated by these metals. Further evaluation and testing  
25 would, therefore, be necessary to assess actual toxicity and its causal relationship to the metals of  
26 concern. If data on the sediment-specific SEM, AVS, and organic carbon concentrations are  
27 available, the uncertainty bounds for  $(\sum \text{SEM-AVS})/f_{\text{OC}}$  described in the metals ESB could be  
28 used to further classify sediments as those in which metals are not likely to cause toxicity, metal  
29 toxicity predictions are uncertain, or metal toxicity is likely. For sediments in which toxicity is  
30  
31  
32

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1 likely or uncertain, acute and chronic tests with species that are sensitive to the metals suspected  
2 to be of concern, acute and chronic sediment toxicity identification evaluations (TIEs), in situ  
3 community assessments, and seasonal and spatial characterizations of the SEM, AVS, and  
4 interstitial water concentrations would be appropriate (Ankley et al., 1994).

#### 6 **4.5.11. Soil Toxicity**

7 Variability among soil toxicity test results is due in part to the influence of soil properties  
8 on bioavailability of metals (e.g., pH, organic matter and CEC). See Sections 3.1, 3.4.6, and 3.2,  
9 covering environmental chemistry bioavailability, terrestrial bioavailability, terrestrial exposure  
10 issues. Additionally, incorporation of sparingly soluble substances, such as many environmental  
11 forms of metals, into the soil matrix is difficult, and acclimation/adaptation of test organisms can  
12 further complicate test results. Use of soluble metal salts with the addition of organism to the  
13 test matrix immediately after mixing is not representative of most environmental situations,  
14 where aging and other physical/chemical processes affect metal speciation and uptake.  
15 Furthermore, testing of soil microbial function is particularly problematic because the test  
16 substance is added to soils with the microbial population already in place and that contain  
17 background amounts of metals.

##### 18 **4.5.11.1. Application**

19 Modifications to standard toxicity bioassays for plants and soil organisms to account for  
20 properties of metals were discussed in an expert workshop and subsequently provided in  
21 Fairbrother et al. (2002). These modifications include directions on type of soil matrices to use,  
22 mixing and aging of metals into the soil, and cautions about acclimation of test organisms.  
23 Future studies conducted specifically for development of toxicity endpoint values for metals can  
24 follow these suggested protocols and circumvent many of the past problems.

25 There is, however, a large body of literature on toxicity of metals to soil organisms that  
26 has already been developed, although often the objectives were to understand processes rather  
27 than to develop defensible toxicity thresholds. The challenge, therefore, lies in how to use these  
28 data, taking into account the test-to-test variability in soil chemistry parameters, and how to  
29 develop a technically defensible means of extrapolating toxicity responses across soil type—in  
30 other words, how to adjust the toxicity threshold values for bioavailability differences in test  
31 conditions. Ideally, an aquatic BLM could be extended to terrestrial setting to account for  
32

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1 differences in bioavailability due to environmental chemistry, particularly for plants and soft-  
2 bodied soil invertebrates. Studies have been initiated to develop what has been referred to as a  
3 *terrestrial BLM*, or tBLM. The conceptual approach to development of a tBLM is very much the  
4 same as for the variations of the BLMs that have been developed for aquatic settings (Allen,  
5 2002). Although not currently developed to the point of being of practical use, experimental  
6 testing and model development programs are fully underway in the hope of providing a tool that  
7 will be of great practical utility in the relatively near future.

8 Another approach to addressing soil variability in soil toxicity tests is to normalize test  
9 results by dividing the LC50 by percent organic matter (van Gestal, 1992). This approach is  
10 based on observed correlations between the LC50 of copper to earthworms and soil organic  
11 matter content. Most recently, CEC has been shown to be the most important factor modifying  
12 zinc bioavailability in soils for both invertebrates and plants, and it will presumably show a  
13 similar relationship with other cationic metals (Smolders, 2003 presentation in DC; based on  
14 work by C. Janssen, Univ. Ghent). Therefore, effect responses in different soil types can be  
15 normalized on the basis of relative CEC. It should be remembered that CEC is a function, at  
16 least in part, of soil pH. Therefore, normalization can be done only among soils of similar pH  
17 ranges. However, comparison of field data with laboratory toxicity response information is best  
18 done through measuring metals in soil pore water from field assessments and comparing such  
19 data to spiked laboratory soils.

20 Several studies of plants and invertebrates have suggested that consideration of aging  
21 may result in significant overestimates of field effects using data laboratory studies (e.g., Smit  
22 and Van Gestel, 1998). Estimates of effects of aging (see Section 4.1.7.2., Environmental  
23 Chemistry for an aging discussion) range from 3 to 8-fold differences between laboratory and  
24 field results.

25 Because most soil properties are correlated to some degree, isolating individual soil  
26 parameters and relating them to soil biota toxicity is difficult. Treating the soil like a black box  
27 and using sequential extraction techniques has been useful in determining what fraction of the  
28 metals are in a labile form (Kabata-Pendias and Pendis, 2000). However, some scientists have  
29 questioned the usefulness of this process (Morgan and Morgan, 1988).

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1 **4.5.11.2. Limitations**

2 Only two methods have been proposed in the literature for normalizing toxicity data  
3 across soils to account for differences in bioavailability: adjusting of endpoint values by percent  
4 organic matter in the soil or as a function of organic matter plus clay content. Both methods fail  
5 to incorporate either pH or cationic exchange capacity, both of which are of critical importance  
6 in determining bioavailability. Aging of metals in soils also is not included in these approaches.  
7 Furthermore, the data sets used to generate the relationships were not sufficiently robust to make  
8 generalizations possible across all soils and all organisms. The development of a tBLM shows  
9 promise as a method that will overcome these limitations; however, it likely will not be  
10 completed for at least 2 years after the publication of this Framework.

11 The use of a tissue residue approach has been suggested as another method to address  
12 soil chemistry and metal toxicity issues, suggesting that a metal concentration must reach a  
13 threshold value in the organism or at the target site before effects begin to occur (McCarthy and  
14 Mackay, 1993; Lanno and McCarty, 1997). For essential elements in plants,  
15 deficiency/sufficiency concentrations in foliage have been developed. However, the relationship  
16 between toxicity and tissue residues is complex and varies depending on tissue type (roots vs.  
17 shoots), plant species, and metal. Little information is available for soil invertebrates, so  
18 relationships between tissue concentration and toxic response cannot yet be developed.  
19 Therefore, this approach, although conceptually sound, requires significant research before  
20 critical tissue levels can be established.

21 For plants, a large proportion of the toxicity literature was developed in support of  
22 understanding potential toxicity and metal uptake from biosolids (e.g., sewage sludge). It is  
23 difficult to determine single-species, single-metal thresholds from this database for several  
24 reasons. First, biosolids tend to contain a mixture of metals, so any response observed cannot be  
25 attributed to a single metal and should account for potential antagonism or synergies that might  
26 occur. Second, biosolids are, by their nature, high in organic matter, which significantly affects  
27 bioavailability of the metals. Until a robust method is developed to adjust toxicity endpoints for  
28 the influence of organic matter (and other bioavailability factors), it will remain difficult to apply  
29 such results to unamended soils. The additional organic matter also provides excess nutrients to  
30 the plants, which further confounds possible metal effects. The guidance for development of  
31 Ecological Soil Screening Levels or EcoSSL (U.S. EPA, 2003c) can be used to judge the  
32 applicability of literature studies to plant or soil invertebrate toxicity threshold determinations.  
33

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#### 1 **4.5.12. Food Chain (Wildlife) Toxicity**

2 Toxicity in wildlife from metals exposures is generally poorly understood and is rarely  
3 quantified in field settings. A few notable exceptions are those mechanisms described in avian  
4 waterfowl exposure to selenium (Adams et al., 2003), exposure of waterfowl to lead-  
5 contaminated sediments (Beyer et al., 1998; Blus et al., 1991; Henny et al., 2000), and white-  
6 tailed ptarmigan exposure to cadmium in vegetation (Larison et.al., 2000). Most metals express  
7 multiorgan toxicity, resulting in a general decrease in overall vigor, as opposed to well-defined  
8 mechanisms of action documented from organic xenobiotics such as pesticides. Typically,  
9 toxicological data used to assess the risk of many metals to wildlife are derived from laboratory  
10 species such as rats or mice or domestic livestock species (e.g., cattle and chickens) exposed to  
11 soluble metal salts. Extrapolating the results of such tests to evaluate toxicity to wildlife is  
12 necessary because of the paucity of data on the toxicity of metals to these receptors. However,  
13 extrapolation of results should be approached with caution due to the large amount of uncertainty  
14 that could be introduced into the risk assessment process (Suter, 1993).

15 Laboratory and domestic species may be more or less sensitive to chemicals than is the  
16 selected receptor. Toxicological responses among species vary because of many physiological  
17 factors that influence the toxicokinetics (absorption, distribution, and elimination) and  
18 toxicodynamics (relative potency) of metals after exposure has occurred. For example,  
19 differences in gut physiology, renal excretion rates, and egg production influence the  
20 toxicokinetics of metals. The ability of some species to more rapidly produce protective proteins  
21 such as metallothionein after exposure to metals are toxicodynamic features leading to  
22 interspecific extrapolation uncertainty. For example, mammal studies should not be extrapolated  
23 to birds, and extrapolation of data from rats (simple, monogastric digestive physiology) to  
24 ruminants introduces more uncertainty than does extrapolation from rats to canids, and so on. In  
25 the case of metals, which some species are able to regulate or store in their tissues without  
26 experiencing toxic effects (i.e., biota-specific detoxification), extrapolations between species  
27 used to assess bioaccumulation and toxicity can be especially problematic.

##### 28 29 **4.5.12.1. Application**

30 Methods for extrapolating metal effects data among species are not unique to metals risk  
31 assessment, with the exception of understanding different requirements for essential elements.  
32 Some of the methods for extrapolating effects data among species include body weight  
33 normalization (Sample and Arenal.,1999), distribution-based approaches (Van Straalen, 2001),

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1 or the use of uncertainty factors (e.g., Calabrese and Baldwin, 1994). All of these approaches  
2 suffer from a lack of an underlying physiological basis quantifying toxicokinetic and  
3 toxicodynamic responses among species. A review of potential extrapolation methodologies can  
4 be found in Kapustka et al. (2004).

5 Currently, the best sources of information on metal toxicity thresholds are NAS/NRC  
6 (1980; 1994), McDowell (2003), and the documentation supporting development of EcoSSLs  
7 values (U.S. EPA, 2003c). The EcoSSL document also includes a general approach for  
8 screening studies for acceptability for use in derivation of toxicity thresholds for risk  
9 assessments that can be used for deriving site-specific TRVs for the most applicable endpoints.  
10 These endpoints should then be extrapolated to species with similar physiology, particularly of  
11 the digestive system, due to the predominance of the dietary exposure pathway (e.g., cow data  
12 can be applied to wild bovids such as bison and possibly to other ruminants such as deer or elk).  
13 Uncertainty factors can be carefully applied if there is concern for extrapolation of data to  
14 species in a different taxonomic category (e.g., genus, family or class). General summaries for  
15 some metals are available in Beyer et al. (1996) and Fairbrother et al. (1996).

#### 16 17 **4.5.12.2. Limitations**

18 Information on toxicity of metals to wildlife under field conditions is severely limited,  
19 focusing on only a few species and a few metals. Deriving TRVs for metals in wildlife is  
20 problematic because the administered form of metal is typically not found in most applied  
21 settings. For example, lead has one of the largest databases for laboratory exposures but has  
22 been limited by the form of the metal studied. Until recently, almost all field studies were  
23 conducted in support of toxic effects of lead shot (i.e., pure elemental lead), whereas almost all  
24 laboratory studies have administered lead to test subjects as lead acetate. These extreme forms  
25 of lead relative bioavailability make extrapolations to dietary exposures difficult for this  
26 substance. The best approach in this case is the use of critical tissue residues, because liver lead  
27 levels indicative of lead poisoning are well established (Beyer et al., 1996). Selenium is another  
28 example where tissue-based toxicity thresholds (in this case, in the avian egg) may be most  
29 appropriate (e.g., Adams et al., 2003).

30 Cross-species extrapolations should be conducted with some knowledge of animal  
31 physiology and specific responses to metals. Digestive physiology is the most important  
32 distinction, because most metal exposures in wildlife are by the dietary route. However, other  
33 specific organismal responses should be understood as well. For example, sheep are much more

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1 sensitive to copper than are other ruminants because of the particular nature of their gut flora  
2 (NAS/NRC, 1980). Therefore, extrapolation of sheep data to other ruminants would be highly  
3 overconservative. On the other hand, pigs are extremely tolerant to copper, possibly due to low  
4 gut uptake rates (NAS/NRC, 1980), so extrapolations of such data to other monogastric animals  
5 would not be protective.

6 Interactions of metals (see Section 3.3. Human Health) also should be taken into account  
7 when analyzing metal toxicity data for wildlife. As noted above, values that may be protective  
8 for a particular metal within a certain animal may not be so if other metals are present or  
9 deficient (or vice versa). Therefore, dietary studies should be examined to understand the  
10 presence of other metals and to ascertain the sufficiency of essential elements. Application of  
11 single-metal thresholds to field situations, whether in a site-specific context or on a national  
12 scale, should make provision for adjustment to account for interactive effects.  
13

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