
2. Materials

2.1 Analytical standard – terbufos

Identity	Terbufos
Chemical name (IUPAC)	S-tert-butylthiomethyl O,O-diethyl phosphorodithioate
Structure	$(\text{C}_2\text{H}_5\text{O})_2\text{—}\overset{\text{S}}{\parallel}\text{P—SCH}_2\text{SC}(\text{CH}_3)_3$
Storage conditions	Ambient
Batch number	0351H01HC
CAS number	13071-79-9
Purity	89.3%
Supplier	Sponsor
Re-test date	17 Dec 2012

2.2 Analytical standard – terbufos sulfoxide

Identity	Terbufos sulfoxide
Chemical name (IUPAC)	S-[(tert-butylsulfinyl)methyl] O,O-diethyl phosphorodithioate
Structure	$(C_2H_5O)_2 - \overset{\overset{S}{ }}{P} - SCH_2SOC(CH_3)_3$
Storage conditions	Refrigerator (approx 4°C)
Batch number	AC11957-97B
CAS number	10548-10-4
Purity	95.8%
Supplier	Sponsor
Expiry date	1 January 2013

2.3 Analytical standard – terbufos sulfone

Identity	Terbufos sulfone
Chemical name (IUPAC)	S-[(tert-butylsulfonyl)methyl] O,O-diethyl phosphorodithioate
Structure	$(C_2H_5O)_2 - \overset{\overset{S}{ }}{P} - SCH_2SO_2C(CH_3)_3$
Storage conditions	Refrigerator (approx 4°C)
Batch number	L67-206
CAS number	56070-16-7
Purity	99.5%
Supplier	Sponsor
Re-test date	1 April 2012

Certificates of Analysis are presented in Appendix 1.

2.4 Control matrices

The ground water was obtained from Anglian Water Denton Lodge Borehole 1 and the surface water was obtained from Diss Mere. Upon receipt the water samples were allocated a unique Huntingdon Life Sciences, Environmental Analysis Department identification number. The water was characterised in separate studies and the characterisation data is presented in the following table:

Parameter	Found value (surface water)	Found value (ground water)
pH	8.07	7.62
Dissolved Oxygen (analysed on the date of sampling)	7.32 mgO ₂ /L	10.57 mgO ₂ /L
Conductivity	376 µS/cm	397 µS/cm
Alkalinity	198 mg/l as CaCO ₃	568 mg/l as CaCO ₃
Total Hardness	221 mg/l as CaCO ₃	228 mg/l as CaCO ₃
Total Organic Carbon	16.198 mgC/L	1.8 mgC/L
Dissolved Organic Carbon	13.814 mgC/L	3.1 mgC/L

3. Methods

3.1 Validation

Sub-samples of each of the two water types were fortified with known concentrations of the analytes simultaneously and analysed according to the following regime:

- 2 sub-samples of untreated sample water
- 5 sub-samples of untreated sample water fortified at the LOQ (0.1 µg/L)
- 5 sub-samples of untreated sample water fortified at 1 µg/L

These samples were then analysed using the analytical methodology, with each sample injected onto the chromatographic system once.

3.2 Final extract stability

An experiment was set up to demonstrate the stability of the analytes under the typical storage conditions of the final extracts if they are not quantified immediately after preparation. Processed control extracts, fortified with the three analytes simultaneously were stored at approximately -20°C in the dark (i.e. in a freezer).

Aliquots of each of the control sample extracts were fortified with the three analytes at a concentration of 2.5 ng analyte/mL of final extract. The concentration of analytes in the stored extracts was determined at day 0 and after 7 days. The concentration of the analytes in freshly fortified control extracts was also determined at the same time that samples stored for 7 days were analyzed.

3.3 Matrix effects

Any possible sample matrix effects were investigated by the comparison of the instrument response to the analytes in the fortified final extract samples with the response of the analytes in solvent based calibration standard solutions prepared at the same time.

3.4 Analytical method

Samples of water (25 mL) were extracted and cleaned up using solid phase extraction (SPE) cartridges, eluting with acetonitrile (3 mL) which was subsequently diluted to volume (5 mL) with HPLC water. Quantitation was performed using liquid chromatography with tandem mass spectrometric detection (LC-MS/MS). The ion transitions monitored for quantitation were m/z 289>103, m/z 305>187 and m/z 321>171 for terbufos, terbufos sulfoxide and terbufos sulfone respectively. To demonstrate possible confirmation of residues, alternative ion transitions, m/z 289>233, m/z 305>243 and m/z 321>265 were also monitored, respectively.

The analytical method used in the laboratory is presented in Appendix 3.

3.5 Fortification/calibration solutions

Individual stock standard solutions (1 mg/mL) of the three analytes were prepared by dissolving an accurately weighed amount in a suitable volume of acetonitrile, correcting for purity as appropriate. These stock solutions were further diluted with acetonitrile to produce mixed fortification solutions at 10 µg/mL, 1 µg/mL and 0.1 µg/mL concentrations.

The instrument calibration solutions, over the concentration range 0.1 ng/mL to 10 ng/mL, were prepared by serial dilution of the fortification solution in acetonitrile:water (60:40 v:v), as detailed below:

Standard solution used (ng/mL)	Volume taken (mL)	Final volume (mL)	Nominal concentration (ng/mL)
1000	0.1	10	10
1000	0.05	10	5
1000	0.025	10	2.5
10	1	10	1
10	0.75	10	0.75
10	0.5	10	0.5
10	0.25	10	0.25
1	1	10	0.1

The standard solutions used in this study were also used in other GLP studies being performed for the same Sponsor. The use of these standard solutions is fully traceable to the other studies and copies of the standard solution preparation are included in the raw data package for this study.

3.6 Calculation of results for validation samples

Test samples were quantified using the following equation:

$$\text{Residue found } (\mu\text{g/L}) = x \times \frac{1}{M} \times D$$

Where x (residue concentration in final solution) was calculated using the linear regression

$$y = mx + c \quad \text{where } x \text{ (concentration in ng/mL)} = \frac{y - c}{m}$$

c	=	intercept
m	=	slope
y	=	peak area of sample
M	=	matrix concentration (mL/mL)
D	=	dilution factor

Example calculation of terbufos detected in surface water fortified at 1 $\mu\text{g/L}$ (analytical identification 11/00/11826 F1 A, analysis batch 1). The primary data for this sample is presented in Table 20, Appendix 2.

Linear regression $y = mx + c$

$$2.39784\text{e}5 = 62293.3x + 1074.18$$

where

$$y = 2.39784\text{e}5$$

$$m = 62293.3$$

$$c = 1074.18$$

Therefore, concentration of terbufos (x) = $\frac{2.39784\text{e}5 - 1074.18}{62293.3} = 3.832 \text{ ng/mL}$

Matrix concentration = 5 mL matrix/mL final extract
Dilution factor = 1

$$\text{Terbufos detected } (\mu\text{g/L}) = \frac{3.832\text{ng/mL} \times 1}{5 \text{ mL/mL}} = 0.766 \mu\text{g/L}$$

$$\text{Recovery } (\%) = \frac{0.766 \mu\text{g/L} \times 100}{1 \mu\text{g/L}} = 77\%$$

Appendix 3 Analytical Method

DETERMINATION OF TERBUFOS, TERBUFOS SULFOXIDE AND TERBUFOS SULFONE IN WATER

1. General principle

Samples are extracted and cleaned up using solid phase extraction (SPE) cartridges. Quantitation is performed using liquid chromatography with tandem mass spectrometric detection (LC-MS/MS).

2. Apparatus, glassware etc

Balances (various ranges)
Volumetric flasks (various sizes)
Syringes (various sizes)
Volumetric pipettes (various sizes)
Polypropylene tubes (15 and 50 mL)
Measuring cylinders (various sizes)

3. Materials

Acetonitrile
Ammonium formate
Methanol
Formic acid
Water
Oasis HLB cartridges (60 mg, 3 mL)

Typical Grade (or equivalent)

HPLC
AR
HPLC
AR
HPLC

4. Preparation of reagents

Preparation of acetonitrile:water (60:40 v:v) - acetonitrile (600 mL) is mixed thoroughly with water (400 mL).

Preparation of acetonitrile:water (20:80 v:v) - acetonitrile (200 mL) is mixed thoroughly with water (800 mL).

Preparation of water:methanol:formic acid (90:10:0.1 v:v:v) containing 0.01M ammonium formate - methanol (100 ml), ammonium formate (0.6 g) and formic acid (1 ml) is added to HPLC water (900 ml) and mixed thoroughly prior to use.

Preparation of methanol:formic acid (100:0.1 v:v) - methanol (1000 mL) is mixed thoroughly with formic acid (1 mL).

Note: variable quantities of the above may be prepared by adjusting the constituent quantities accordingly.

5. Analytical standard solutions

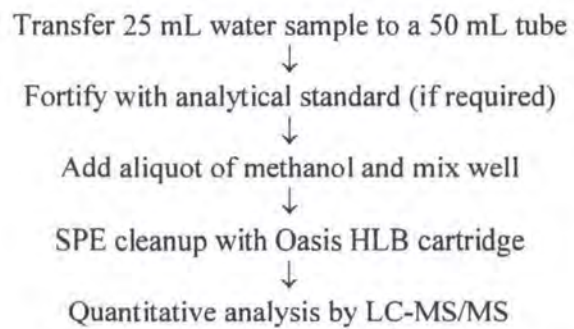
An appropriate amount of the test substances (corrected for purity) are accurately weighed and dissolved in acetonitrile to give the individual stock standard solutions (typically 1 mg/mL concentration). Appropriate dilutions of the stock standard solutions are made with acetonitrile to give mixed fortification standard solutions (typically 10 µg/mL, 1 µg/mL and 0.1 µg/mL).

The mixed fortification solutions are progressively diluted with acetonitrile:water (60:40 v:v) to produce a series of instrument calibration solutions in the range 0.1 to 10 ng/mL.

6. Procedure

- 6.1 Transfer an aliquot of sample water (25 mL) to a 50 mL polypropylene tube.
- 6.2 Add fortification solution at this stage if required.
- 6.3 Add an aliquot (10 mL) of methanol and mix well.
- 6.4 Condition the Oasis HLB SPE cartridge with acetonitrile (3 mL) and water (3 mL), discarding the eluate.
- 6.5 Load the extract from 6.3 onto the SPE cartridge, discarding the eluate.
- 6.6 Wash the cartridge with an aliquot (5 mL) of acetonitrile:water (20:80 v:v), discarding the eluate, allowing the cartridge to have air pumped through for approximately 30 seconds to remove excess solvent.
- 6.7 Elute the SPE cartridge with an aliquot (3 mL) of acetonitrile, collecting in a 15 mL polypropylene tube.
- 6.8 Dilute the final extract to volume (5 mL) with water. Final matrix concentration \equiv 5 mL sample matrix / mL final extract.
- 6.9 Perform any further dilutions using acetonitrile:water (60:40 v:v), as required.
- 6.10 Quantify the samples by the use of LC-MS/MS.

7. Flow chart of analytical procedure



8. LC-MS/MS conditions

Instrument:	AB Sciex API 4000																					
Mode:	Ionspray positive																					
Ion monitoring details:	Terbufos: m/z 289>103 m/z 289>233 (confirmatory) Terbufos sulfoxide: m/z 305>187 m/z 305>243 (confirmatory) Terbufos sulfone: m/z 321>171 m/z 321>265 (confirmatory)																					
Column:	Acquity UPLC [®] BEH C ₁₈ (2.1 mm x 50 mm, 1.7 µm), or equivalent, column temperature 45°C																					
Mobile phase A:	Water:methanol (90:10 v:v) + 0.01M ammonium formate + 0.1% formic acid																					
Mobile phase B:	Methanol:formic acid (100:0.1 v:v)																					
Gradient:	<table> <thead> <tr> <th>Time</th> <th>%A</th> <th>%B</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>50</td> <td>50</td> </tr> <tr> <td>0.2</td> <td>50</td> <td>50</td> </tr> <tr> <td>2.0</td> <td>5</td> <td>95</td> </tr> <tr> <td>2.5</td> <td>5</td> <td>95</td> </tr> <tr> <td>3</td> <td>50</td> <td>50</td> </tr> <tr> <td>4</td> <td>50</td> <td>50</td> </tr> </tbody> </table>	Time	%A	%B	0	50	50	0.2	50	50	2.0	5	95	2.5	5	95	3	50	50	4	50	50
Time	%A	%B																				
0	50	50																				
0.2	50	50																				
2.0	5	95																				
2.5	5	95																				
3	50	50																				
4	50	50																				
Cycle time:	4 min																					
Injection volume:	10 µL																					
Flow rate:	0.5 mL/min																					
Retention times:	Terbufos: approx. 1.8 minutes Terbufos sulfoxide: approx. 1 minute Terbufos sulfone: approx. 1 minute																					
LOQ:	0.1 µg/L																					
LOD:	0.1 ng/mL (=0.02 µg/L in sample matrix)																					

NOTE – alternative instruments may also be used, operated under conditions that are considered to be equivalent to those described above. However, some differences may be observed in the resulting data, such as slight differences in analyte retention times, or the observed sensitivity of the ion transitions monitored.