

# Using Biochemical Methane Potentials & Anaerobic Toxicity Assays

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# To Be Discussed:

- Description of BMPs and ATAs
- Utilization of BMPs and ATAs in planning process
- Assay process
- Interpreting and utilizing results

BMP - Biochemical Methane Potential

ATA - Anaerobic Toxicity Assay

# Controlled Anaerobic Digestion

- Careful planning and accurate design are necessary to optimize cost recovery
- Multiple data and information sources are available for initial planning
  - Using “book values” can lead to over or under-estimated performance
  - BMPs and ATAs can provide valuable information

# Biochemical Methane Potential Assays (BMPs)

- Bench-scale test, generally 30 day duration
- Developed to determine anaerobic biodegradability of substrate
- Substrate is tested in a laboratory environment under optimal conditions



# BMP Uses

1. Determine concentration of organics in a wastewater that can be anaerobically converted to  $\text{CH}_4$
2. Evaluate potential efficiency of anaerobic process for a specific waste
3. Measure residual organic material amenable to further anaerobic treatment

# Anaerobic Toxicity Assays (ATAs)

- Bench-scale test, 3-5 day duration
- Developed to determine the inhibition of  $\text{CH}_4$  production by a given material
- Material is tested in a laboratory environment under optimal conditions



# ATA Uses

- Predicts likely effect of potential toxicant on biogas and CH<sub>4</sub> production
- Standard ATA does not show microorganism acclimation to toxicant or effect of toxicant build-up in the biomass
  - Continuous bench scale tests may be needed to study acclimation and toxicity due to compound build-up in the biomass

# BMP and ATA References

- BMP

- Owen et al., 1979; Water Research 13:485-492
- Speece, 2008; Anaerobic Biotechnology & Odor/Corrosion Control
- ASTM E2170-01
- ISO 11734

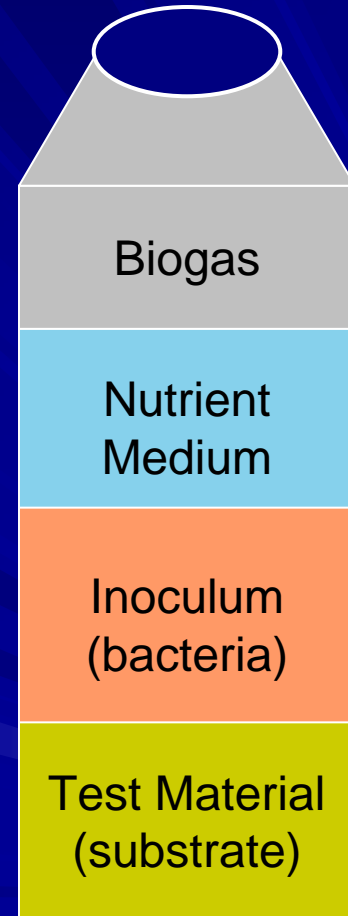
- ATA

- Owen et al., 1979; Water Research 13:485-492
- ISO 13641-1



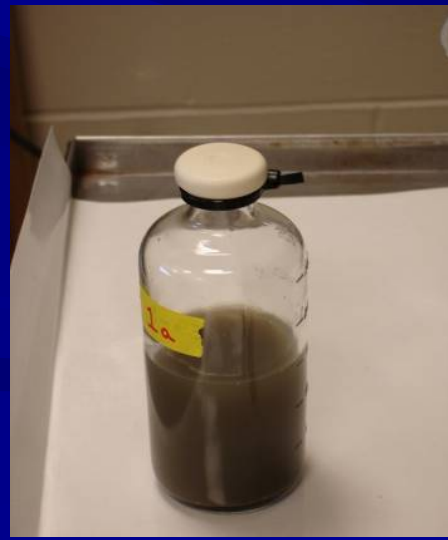
# BMP General Principles

- Characterize substrate/test material:
  - pH, COD, TS, VS
- Place aliquot of test material in a serum bottle with inoculum & nutrient medium
- Prepare blank with inoculum & nutrient medium
- Prepare each assay in triplicate



# BMP General Principles

- Purge bottles with 30% CO<sub>2</sub> / 70% N<sub>2</sub> gas, seal, & place on shaker at 35°C for 30 days



# BMP General Principles

- Measure biogas production & CH<sub>4</sub> content (daily or as needed depending on production)



# BMP General Principles

- Determine normalized biogas & CH<sub>4</sub> production (mL CH<sub>4</sub> / g substrate VS) and the extent of substrate anaerobic biodegradation (%)

# Inoculum Bacteria

- Some labs get inoculum from local anaerobic treatment system and others maintain their own
  - Maintaining consistent source of inoculum removes some variability (Owen et al., 1979)
- Provide consistent food source to bacteria before assay



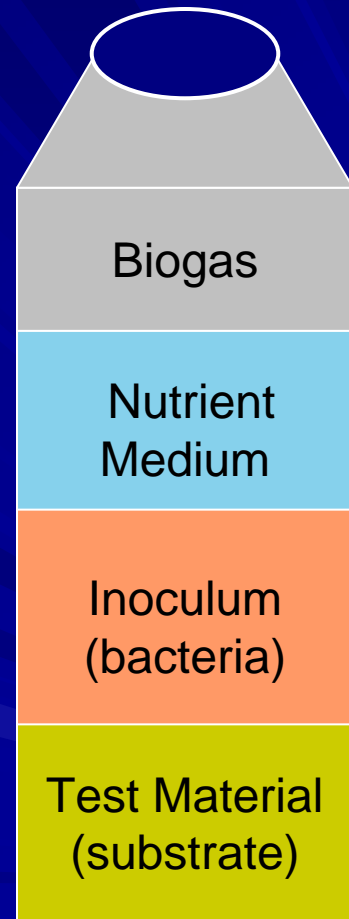
# Quantity of Substrate and Inoculum to Add to BMP Bottle

1. Obtain measurable (but not excessive)  $CH_4$  in BMP
  - Aim for at least 100-150 mL  $CH_4$  / bottle, or 165-250 mL biogas @ 60%  $CH_4$
2. Use substrate [COD] to determine BMP substrate volume
  - 1 g COD reduction = 395 mL  $CH_4$

mg COD required for 125 mL  $CH_4$ :

$$125\text{mL}CH_4 \times \frac{1\text{mg}COD}{0.395\text{mL}CH_4} \times 70\% = 221.5\text{mg}COD$$

\* Assumes 70% COD conversion efficiency



# Quantity of Substrate and Inoculum to Add to BMP Bottle

3. Calculate mass VS in BMP assay based on substrate volume

$$221.5 \text{ mg COD} \times \frac{1000 \text{ mL}}{X \text{ mg COD}} = Y \text{ mL substrate}$$

$$Y \text{ mL substrate} \times \frac{Z \text{ mg VS}}{1000 \text{ mL}} = A \text{ mg VS}$$

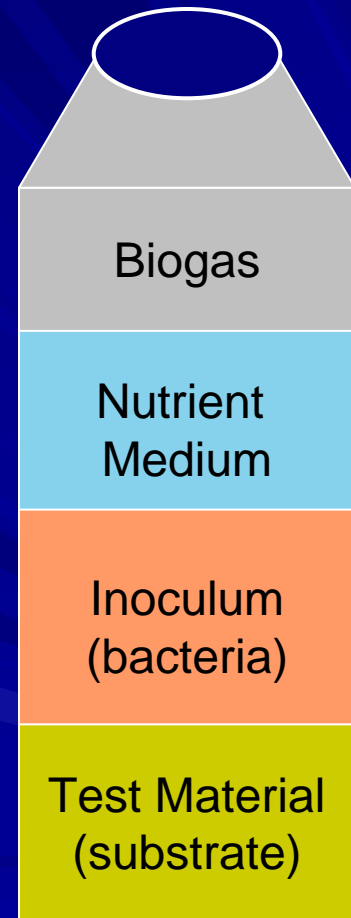
- Where:

X = COD concentration of substrate

Y = mL substrate to obtain necessary CH<sub>4</sub> production

Z = VS concentration of substrate

A = mg substrate to obtain necessary CH<sub>4</sub> production



# Quantity of Substrate and Inoculum to Add to BMP Bottle

4. Calculate volume of inoculum to add to BMP bottle
  - 1 g VS substrate / 1 g VS inoculum

$$\text{inoculum (mL)} = \frac{Y \text{ mL substrate} * Z \text{ mg VS / L}}{B \text{ mg VS / L}} = C \text{ mL inoculum}$$

$$C \text{ mL innoc} \times \frac{B \text{ mg VS}}{1000 \text{ mL}} = D \text{ mg VS inoculum}$$

Where:

Y = mL substrate to obtain necessary CH<sub>4</sub> production

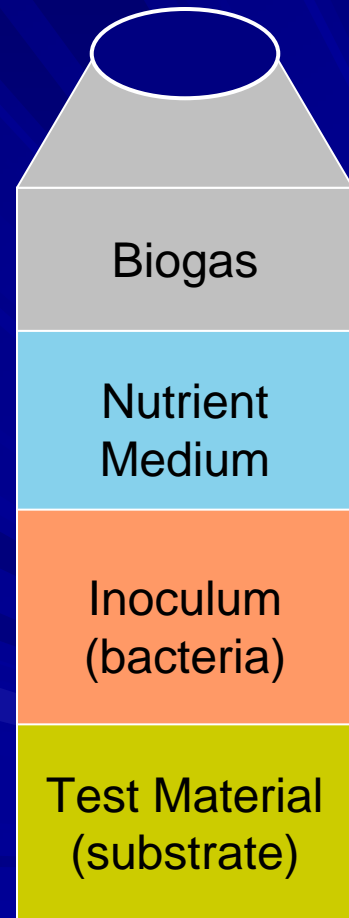
Z = VS concentration of substrate

B = VS concentration of inoculum

C = mL inoculum for 1:1 ratio

D = mass of inoculum for 1:1 ratio

5. Fill remainder of bottle with Nutrient Medium





# BMP Results

- Calculate total  $\text{CH}_4$  production using daily biogas volume and daily  $\text{CH}_4$  content
  - Use blank to account for daily biogas and methane contributed by the inoculum
- Cumulative biogas and methane production between substrates can not be directly compared due to differences in assay loading rates for each material

# BMP Results

- Normalize biogas and methane production to perform comparisons between substrates
- Normalize to mL / g substrate VS in assay

$$\text{Normalized CH}_4 \text{ production} = \frac{\text{ml CH}_4}{\text{g VS}} = \frac{\text{ml CH}_4 \text{ produced}}{\frac{\text{g VS substrate}}{\text{ml}} \times \text{ml substrate in bottle}}$$

Sample Type	TS (%)	VS (%)	COD (mg/L or mg/Kg)	Normalized CH <sub>4</sub> Yield (mL CH <sub>4</sub> /g VS)	BMP Std. Dev.	Methane Yield (m <sup>3</sup> CH <sub>4</sub> /mtonne)
Potato Chips	99.8	93.2	729,000	582	60	542.4
Food Grease	42.3	41.5	1,652,000	811	75.6	336.6
Dairy Manure	15.1	7.2	56,000	264	15.1	19.0

# How do BMPs Compare to Full Scale Digesters?



**vs.**



# How do BMPs Compare to Full Scale Digesters?

- ISU worked with Cornell University to compare BMP assay results to biogas and  $\text{CH}_4$  production from full scale digesters
  - Cornell identified and obtained feedstock for 5 dairy digesters with biogas and  $\text{CH}_4$  monitoring systems

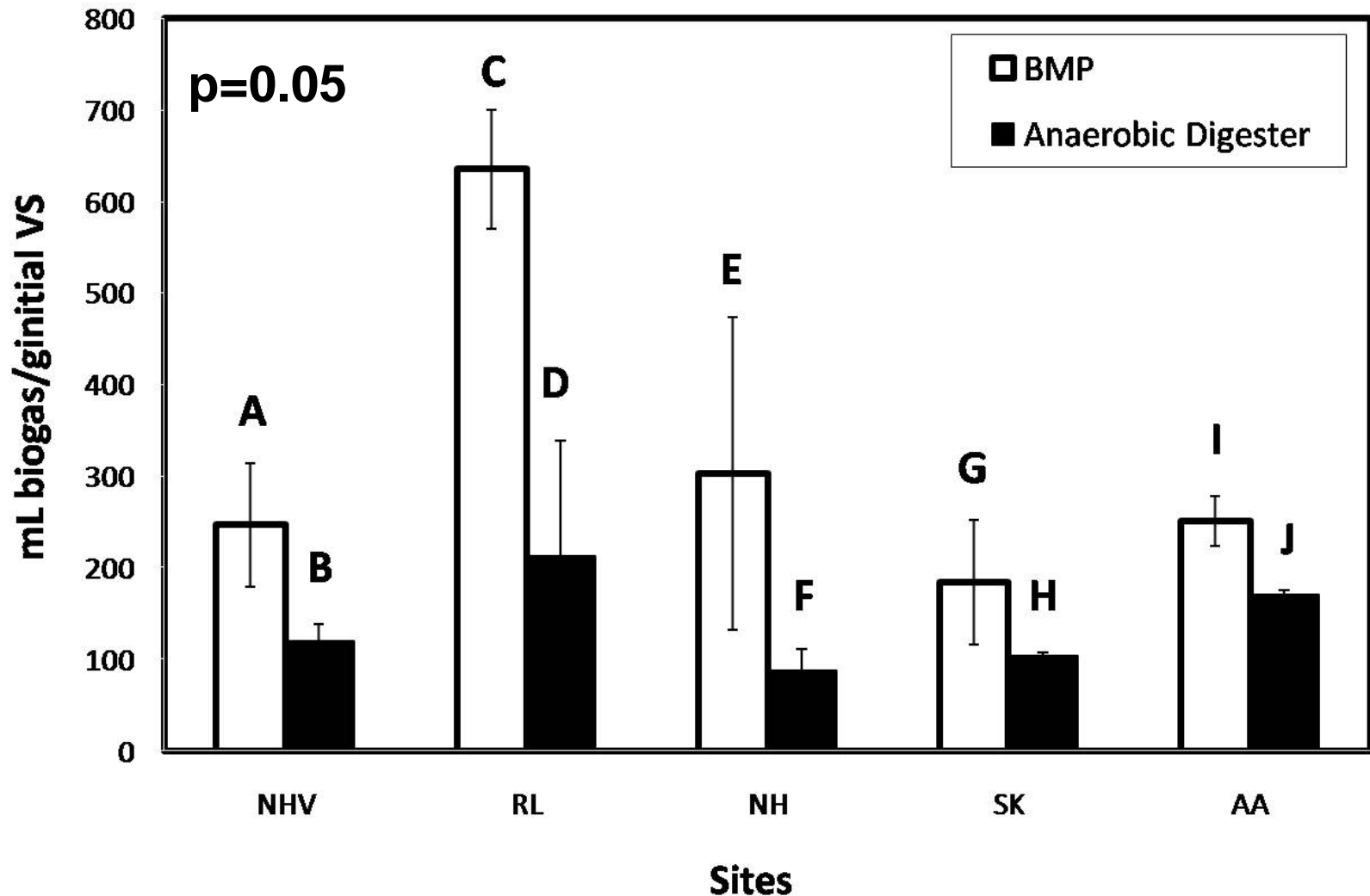


# How do BMPs Compare to Full Scale Digesters?

- ISU characterized the feedstocks and performed BMP assays on each
- Cornell simultaneously collected farm biogas production and CH<sub>4</sub> content data
- Data from the BMPs and full scale systems were normalized and compared

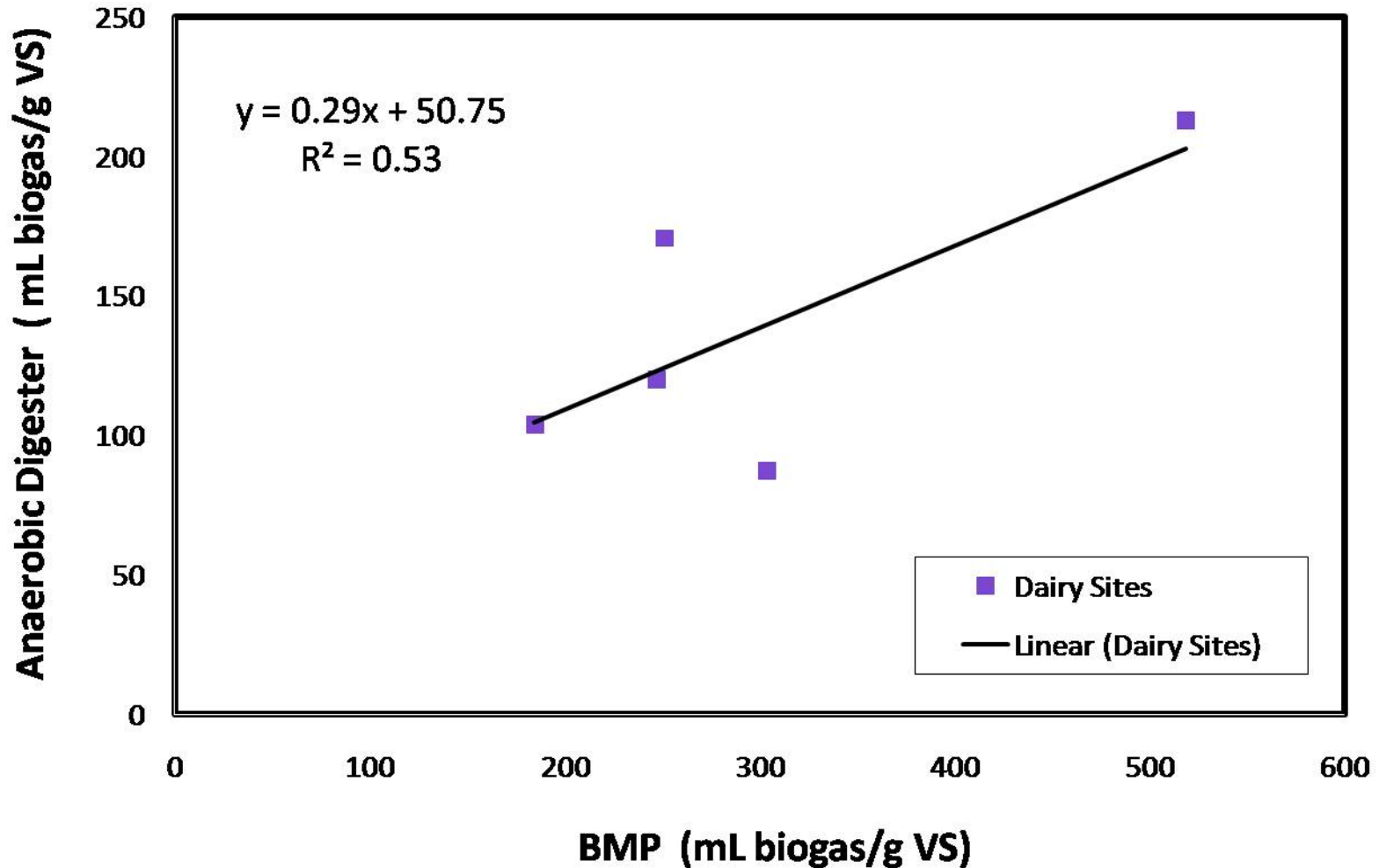
*Bishop et al. 2009. Evaluation of Laboratory Biochemical Methane Potentials as a Predictor of Anaerobic Dairy Manure Digester Biogas and Methane Production. Proceedings of the 2009 ASABE International Meeting. June 21-24, 2009. Reno, NV*

# Biogas Production Comparison



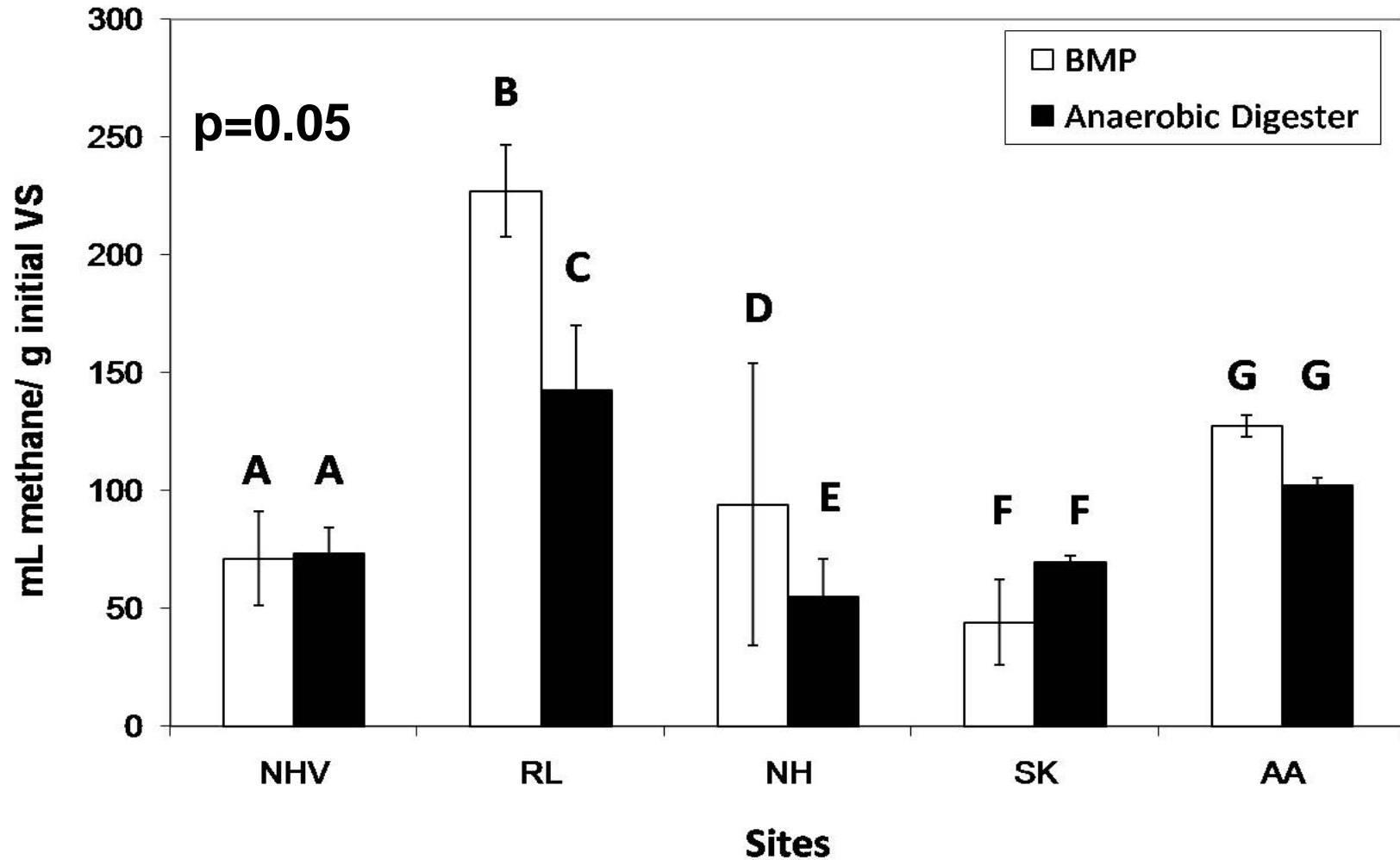
BMPs don't seem to be a good predictor of biogas production in full scale digesters.

# Regression of Biogas Production



The  $R^2$  value does not indicate a strong comparison between BMP and full scale biogas production.

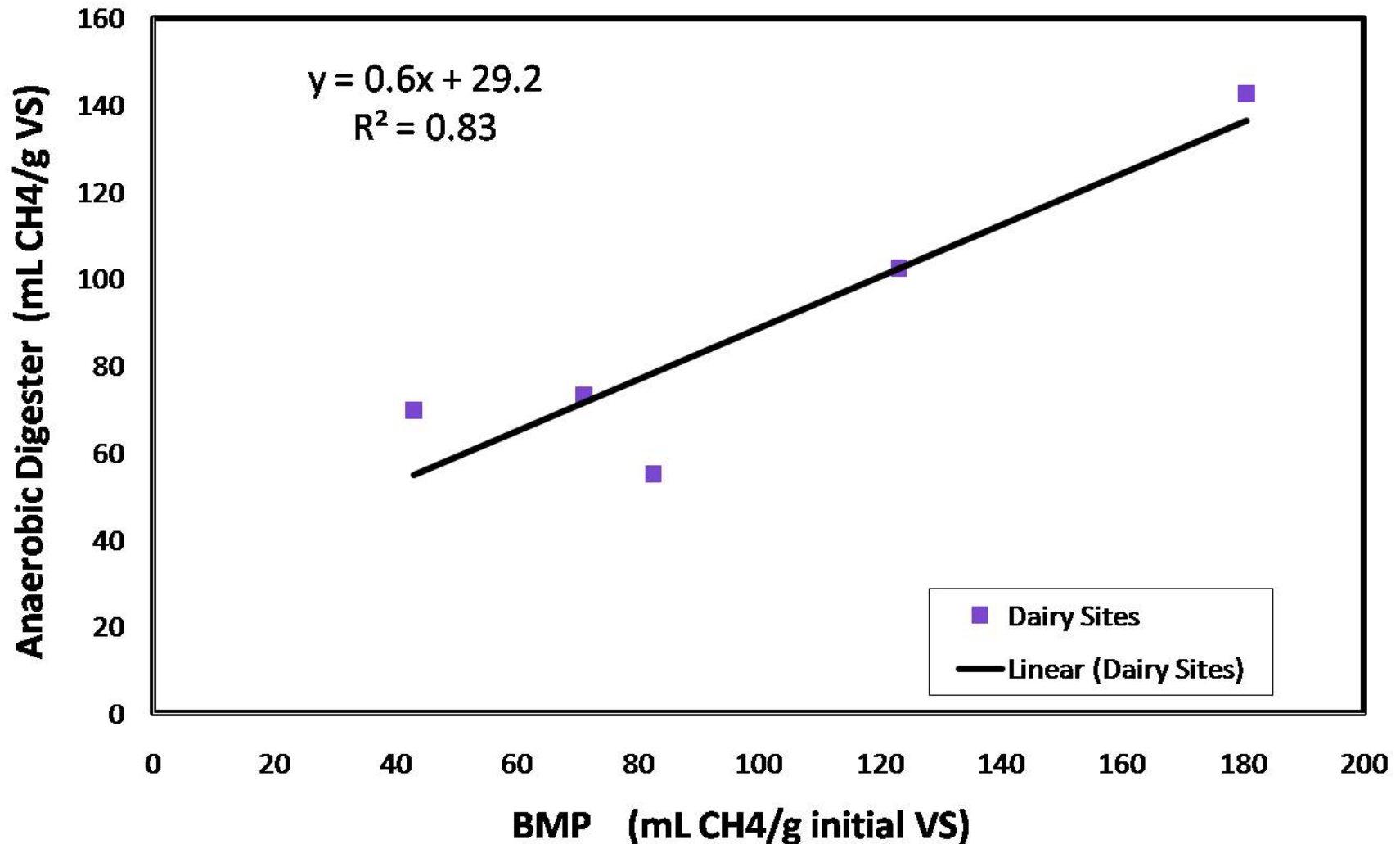
# Methane Production Comparison



BMPs provide a better prediction of methane production than biogas production in full scale digesters.



# Regression of Methane Production



The  $R^2$  value indicates a good correlation between BMP and full scale methane production.

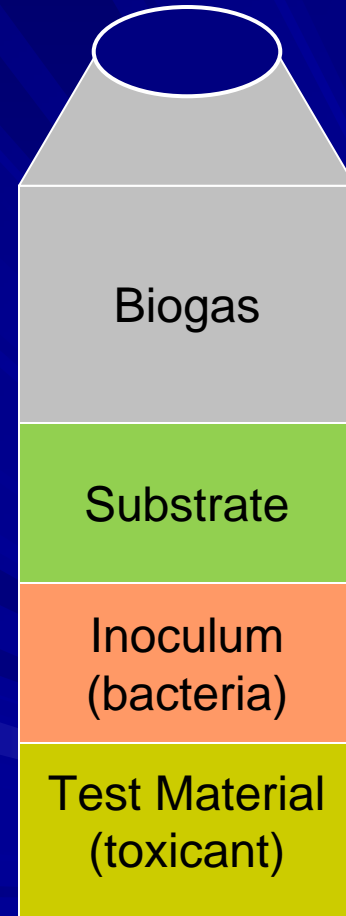


# Summary

- Biogas and methane production were over-predicted by BMPs
  - Average biogas production over predicted by 51.4%
  - Average methane production over predicted by 1.2%

# ATA General Principles

- Characterize the test material/toxicant:
  - pH, COD, TS, VS
- Place aliquot of test material in serum bottle with substrate & inoculum
  - Prepare assays across range of test material concentrations
- Prepare control with substrate & inoculum
- Prepare each assay in triplicate



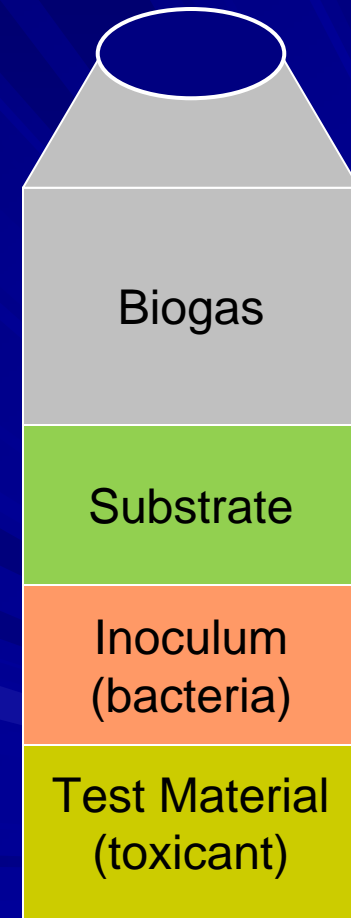
# ATA General Principles

- Purge bottles with 30% CO<sub>2</sub> / 70% N<sub>2</sub> gas, seal, & place on shaker at 35°C for 3 - 5 days
- Measure daily biogas production & CH<sub>4</sub> content
- Develop inhibition curves, calculate percent inhibition, and determine the EC<sub>50</sub>

EC<sub>50</sub> - half maximal effective concentration – concentration of toxicant which induces a response halfway between the baseline and maximum response after a specified exposure time

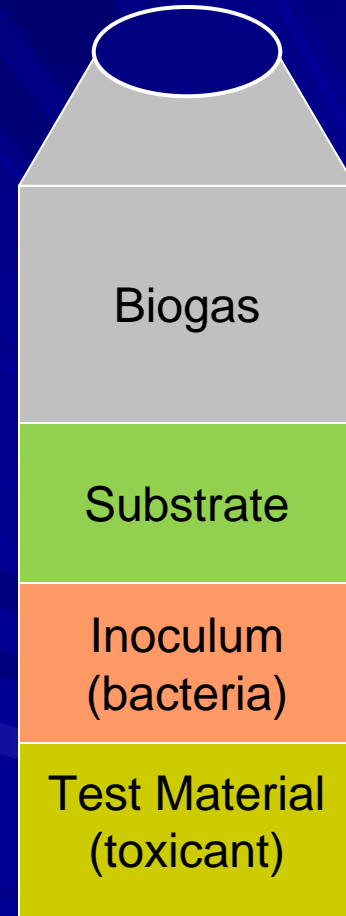
# Quantity of Test Material, Substrate and Inoculum to Add to ATA Bottle

- Develop range of test material concentrations for the assay
  - Use known toxicity range for similar compounds
  - Perform preliminary range finding tests
  - Use range around expected loading rate
- Add test material to assay and use “make-up” water as necessary to achieve equal volumes



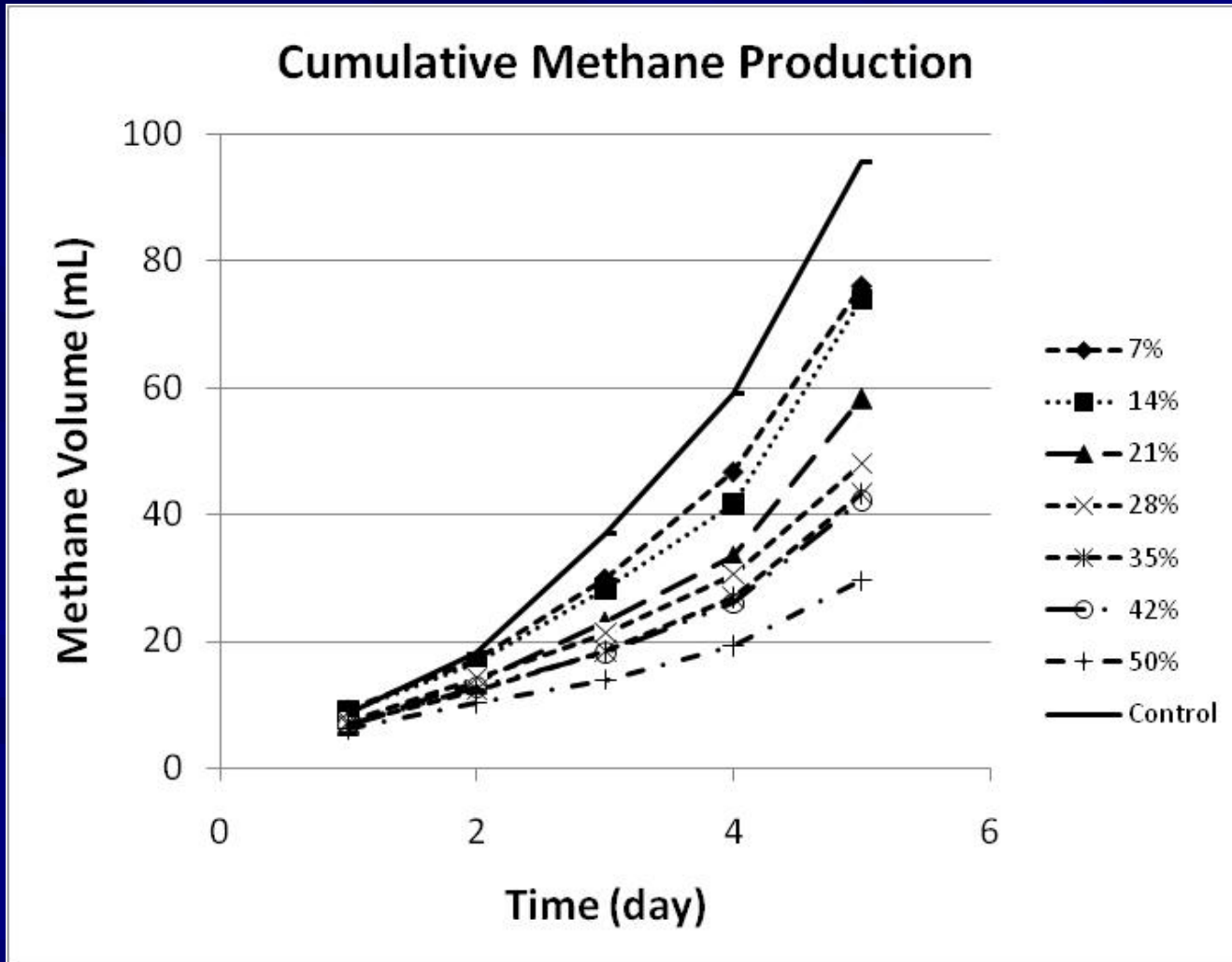
# Quantity of Test Material, Substrate and Inoculum to Add to ATA Bottle

- Add 2 – 4 g of inoculum solids
  - 100 mL of solids with a concentration of 20-40 g/L
- Add 2 ml of substrate
  - Standard mix of nutrient broth, yeast extract, and D-glucose
- Prepare a control of substrate and inoculum for use in determining inhibition



# ATA Results

- Graph cumulative CH<sub>4</sub> production against time for control and test material ranges



Data is from the ISU lab; this graph is the intermediate step to developing an inhibition curve.

# ATA Results

- Select time on linear part of curve (usually 48hrs) and calculate percent inhibition

$$I = \left(1 - \frac{CH_4_{test}}{CH_4_{control}}\right) * 100$$

Where:

I = percent inhibition

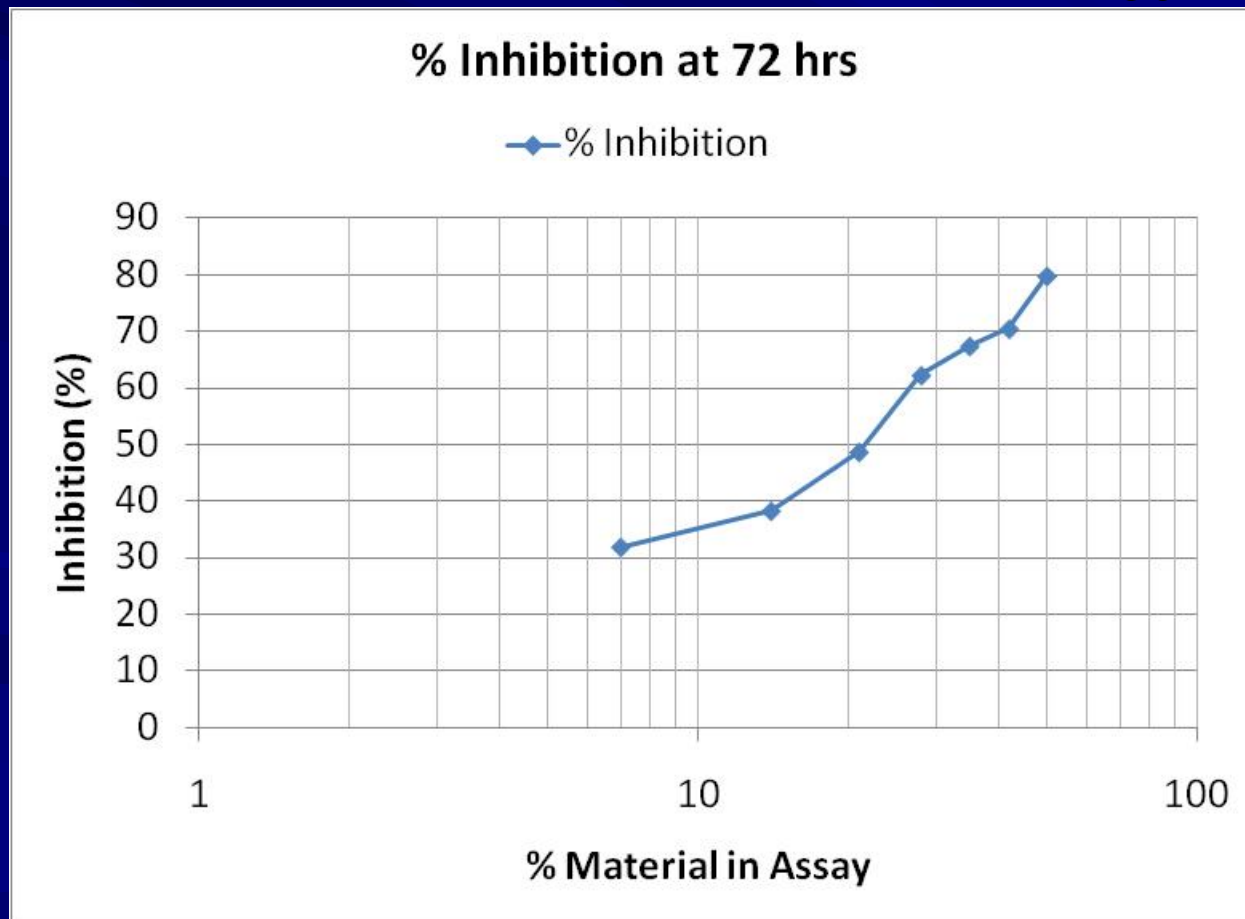
CH<sub>4</sub> test = volume CH<sub>4</sub> in test at a give time

CH<sub>4</sub> control = volume CH<sub>4</sub> in control at a given time



# ATA Results

- Plot Percent Inhibition against the logarithm of the mass concentration of test material and determine  $EC_{50}$



Data is from the ISU lab and represents a potential co-substrate for a manure digester that was determined to be toxic.

# Use Caution When Applying Results

- BMP assays are diluted and can mask substrate toxicity
- BMPs are a batch loaded feed limited process; they are not continuously loaded processes and may not be representative of actual continuously fed digesters
- ATAs don't necessarily account for potential bacteria acclimation to a toxicant or build up of the material in the sludge to toxic levels

# Digester Design



Lab Scale



Bench Scale



Full Scale

- Can't design a digester with a BMP
- Bench & pilot digesters allow better understanding of how selected full scale process will operate
- BMPs & ATAs provide a “first-cut” evaluation when considering co-substrates

The following university labs provide BMP & ATA testing services; these labs are listed for information purposes and ISU makes no endorsement of any lab:

- Iowa State University (**Service Ended 4/15/2010**)

Agricultural Waste Management Lab

Director: Robert Burns, [rburns@iastate.edu](mailto:rburns@iastate.edu)

515-294-4203

- Marquette University

Water Quality Center

Director: Dr. Dan Zitomer, [daniel.zitomer@marquette.edu](mailto:daniel.zitomer@marquette.edu)

(414) 288- 5733

- Michigan State University

Anaerobic Digestion Research & Education Center

Manager: Dana Kirk, [kirkdana@msu.edu](mailto:kirkdana@msu.edu)

517-432-6530

# Questions?

