

2. Materials

2.1 Analytical standard – tribufos

Identity	Tribufos technical
Chemical name (IUPAC)	S,S,S-tributyl phosphorothioate
Structure	$\begin{array}{c} \text{O} \\ \parallel \\ \text{CH}_3(\text{CH}_2)_3\text{S}-\text{P}-\text{S}(\text{CH}_2)_3\text{CH}_3 \\ \diagup \quad \diagdown \\ \text{S}(\text{CH}_2)_3\text{CH}_3 \quad \text{S}(\text{CH}_2)_3\text{CH}_3 \end{array}$
Storage conditions	Ambient
Batch number	NX81AX0015
CAS number	78-48-8
Purity	98.2%
Supplier	Sponsor
Re-test date	30 September 2012

Certificate of Analysis is presented in Appendix 1.

2.2 Control matrices

The ground water was obtained from Anglian Water Denton Lodge Borehole 1 and the surface water was obtained from Diss Mere. Upon receipt the water samples were allocated a unique Huntingdon Life Sciences, Environmental Analysis Department identification number. The water was characterised in separate studies and the characterisation data is presented in the following table:

Parameter	Found value (surface water)	Found value (ground water)
pH	8.07	7.62
Dissolved Oxygen (analysed on the date of sampling)	7.32 mgO ₂ /L	10.57 mgO ₂ /L
Conductivity	376 µS/cm	397 µS/cm
Alkalinity	198 mg/l as CaCO ₃	568 mg/l as CaCO ₃
Total Hardness	221 mg/l as CaCO ₃	228 mg/l as CaCO ₃
Total Organic Carbon	16.198 mgC/L	1.8 mgC/L
Dissolved Organic Carbon	13.814 mgC/L	3.1 mgC/L

3. Methods

3.1 Validation

Sub-samples of each of the two water types were fortified with known concentrations of the analyte and analysed according to the following regime:

- 2 sub-samples of untreated sample water
- 5 sub-samples of untreated sample water fortified at the LOQ (0.1 µg/L)
- 5 sub-samples of untreated sample water fortified at 1 µg/L

These samples were then analysed using the analytical methodology, with each sample injected onto the chromatographic system once.

3.2 Final extract stability

An experiment was set up to demonstrate the stability of the analyte under the typical storage conditions of the final extracts (prepared in acetonitrile) if they are not quantified immediately after preparation. Processed control extracts, fortified with tribufos were stored at approximately -20°C in the dark (i.e. in a freezer).

Aliquots of each of the control sample extracts were fortified with tribufos at a concentration of 2 ng analyte/mL of final extract. The concentration of analyte in the stored extracts was determined at day 0 and after 7 days. The concentration of the analyte in freshly fortified control extracts was also determined at that time.

3.3 Matrix effects

Any possible sample matrix effects were investigated by the comparison of the instrument response to the analyte in the fortified final extract samples with the response of the analyte in solvent based calibration standard solutions prepared at the same time.

3.4 Analytical method

Samples of water (5 mL) were extracted and cleaned up using solid phase extraction (SPE) cartridges, eluting with acetonitrile (4.5 mL) which was subsequently diluted to volume (5 mL). Quantitation was performed using liquid chromatography with tandem mass spectrometric detection (LC-MS/MS) monitoring ion transitions m/z 315>169 (quantitation) and m/z 315>57 (confirmation).

The analytical method used in the laboratory is presented in Appendix 3.

3.5 Fortification/calibration solutions

A stock standard solution (1 mg/mL) of tribufos was prepared by dissolving an accurately weighed amount in a suitable volume of acetonitrile, correcting for purity as appropriate. This stock solution was further diluted with acetonitrile to produce fortification solutions at 10 µg/mL, 1 µg/mL, 0.1 µg/mL and 0.01 µg/mL concentrations.

The instrument calibration solutions, over the concentration range 0.025 ng/mL to 2 ng/mL, were prepared by serial dilution of the fortification solution in acetonitrile, as detailed below:

Standard solution used (ng/mL)	Volume taken (mL)	Final volume (mL)	Nominal concentration (ng/mL)
100	0.2	10	2
100	0.15	10	1.5
100	0.1	10	1
100	0.08	10	0.8
100	0.04	10	0.4
1	2	10	0.2
1	1	10	0.1
1	0.5	10	0.05
1	0.25	10	0.025

The standard solutions used in this study were also used in other GLP studies being performed for the same Sponsor. The use of these standard solutions is fully traceable to the other studies and copies of the standard solution preparation are included in the raw data package for this study.

3.6 Calculation of results for validation samples

Test samples were quantified using the following equation:

$$\text{Residue found } (\mu\text{g/L}) = x \times \frac{1}{M} \times D$$

Where x (residue concentration in final solution) was calculated using the linear regression

$$y = m x + c \quad \text{where } x \text{ (concentration in ng/mL)} = \frac{y - c}{m}$$

c	=	intercept
m	=	slope
y	=	peak area of sample
M	=	matrix concentration (mL/mL)
D	=	dilution factor

Example calculation of tribufos detected in surface water soil fortified at 0.1 $\mu\text{g/L}$ (analytical identification 11/00/11826 F0.1 A, analysis batch 1). The primary data for this sample is presented in Table 8, Appendix 2.

Linear regression $y = m x + c$

$$2.08614e4 = 239087x + 2675.91$$

where $y = 2.08614e4$
 $m = 239087$
 $c = 2675.91$

$$\text{Therefore, concentration of tribufos } (x) = \frac{2.08614e4 - 2675.91}{239087} = 0.0761 \text{ ng/mL}$$

Matrix concentration = 1 mL matrix/mL final extract

Dilution factor = 1

$$\text{Tribufos detected } (\mu\text{g/L}) = \frac{0.0761 \text{ ng/mL} \times 1}{1 \text{ mL/mL}} = 0.0761 \text{ ng/mL} = 0.076 \mu\text{g/L}$$

$$\text{Recovery } (\%) = \frac{0.076 \mu\text{g/L} \times 100}{0.1 \mu\text{g/L}} = 76\%$$

Appendix 3 Analytical Method

DETERMINATION OF TRIBUFOS IN WATER

1. General principle

Samples are extracted and cleaned up using solid phase extraction (SPE) cartridges. Quantitation is performed using liquid chromatography with tandem mass spectrometric detection (LC-MS/MS).

2. Apparatus, glassware etc

Balances (various ranges)
Volumetric flasks (various sizes)
Syringes (various sizes)
Volumetric pipettes (various sizes)
Polypropylene tubes (15 mL)
Measuring cylinders (various sizes)
Solid phase extraction vacuum manifold

3. Materials

Acetonitrile
Ammonium formate
Methanol
Formic acid
Water
Oasis HLB cartridges (60 mg, 3 mL)

Typical Grade (or equivalent)

HPLC
AR
HPLC
AR
HPLC

4. Preparation of reagents

Preparation of acetonitrile:water (30:70 v:v) - acetonitrile (30 mL) is mixed thoroughly with water (70 mL).

Preparation of water:methanol:formic acid (90:10:0.1 v:v:v) containing 0.01M ammonium formate - methanol (100 ml), ammonium formate (0.6 g) and formic acid (1 ml) is added to HPLC water (900 ml) and mixed thoroughly prior to use.

Preparation of methanol:formic acid (100:0.1 v:v) - methanol (1000 mL) is mixed thoroughly with formic acid (1 mL).

Note: variable quantities of the above may be prepared by adjusting the constituent quantities accordingly.

5. Analytical standard solutions

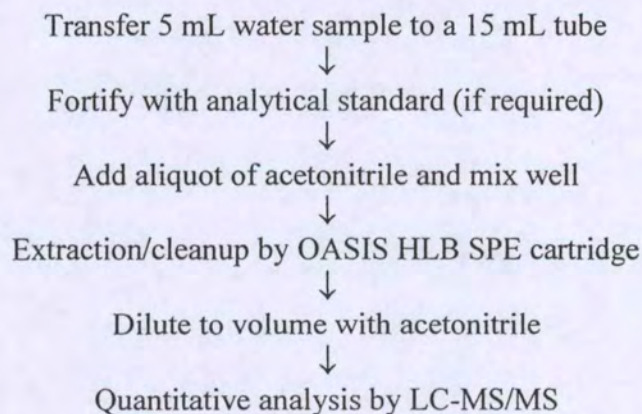
An appropriate amount of the test substance (corrected for purity) is accurately weighed and dissolved in acetonitrile to give the individual stock standard solution. Appropriate dilutions of the stock standard solution are made with acetonitrile to give mixed fortification standard solutions.

The mixed fortification solutions are progressively diluted with acetonitrile to produce a series of instrument calibration solutions in the range 0.025 to 2 ng/mL.

6. Procedure

- 6.1 Transfer an aliquot of sample water (5 mL) to a 15 mL polypropylene tube.
- 6.2 Add fortification solution at this stage if required.
- 6.3 Add an aliquot (2 mL) of acetonitrile and mix well.
- 6.4 Condition the Oasis HLB SPE cartridge with acetonitrile (3 mL) and water (3 mL), discarding the eluate.
- 6.5 Load the extract from step 6.3 onto the SPE cartridge, discarding the eluate.
- 6.6 Wash the cartridge with an aliquot (3 mL) of acetonitrile:water (30:70 v:v), discarding the eluate, allowing the cartridge to have air pumped through for approximately 30 seconds to remove excess solvent.
- 6.7 Elute the SPE cartridge with an aliquot (4.5 mL) of acetonitrile, collecting in a 15 mL polypropylene tube.
- 6.8 Dilute the final extract to volume (5 mL) with acetonitrile. Final matrix concentration \equiv 1 mL sample water / mL final extract.
- 6.9 Perform any further dilutions using acetonitrile, as required.
- 6.10 Quantify the samples by the use of LC-MS/MS.

7. Flow chart of analytical procedure



8. LC-MS/MS conditions

Instrument:	AB Sciex API 4000		
Mode:	Ionspray positive		
Ion monitoring details:	<i>m/z</i> 315>169 <i>m/z</i> 315>57 (confirmatory)		
Column:	Acquity UPLC [®] BEH C ₁₈ (2.1 mm x 50 mm, 1.7 μm), or equivalent, column temperature 45°C		
Mobile phase A:	Water:methanol (90:10 v:v) + 0.01M ammonium formate + 0.1% formic acid		
Mobile phase B:	Methanol:formic acid (100:0.1 v:v)		
Gradient:	Time	%A	%B
	0	30	70
	0.2	30	70
	2.0	5	95
	2.5	5	95
	3	30	70
	4	30	70
Cycle time:	4 min		
Injection volume:	10 μL		
Flow rate:	0.5 mL/min		
Retention time:	approximately 1.5 minutes		
LOQ:	0.1 μg/L		
LOD:	0.025 ng/mL (≡ 0.025 μg/L in sample matrix)		

NOTE – alternative instruments may also be used, operated under conditions that are considered to be equivalent to those described above. However, some differences may be observed in the resulting data, such as slight differences in analyte retention times, or the observed sensitivity of the ion transitions monitored.