

ORGANOPHOSPHORUS PESTICIDES

5600

Formula: Table 1

MW: Table 1

CAS: Table 1

RTECS: Table 1

METHOD: 5600, Issue 1

EVALUATION: FULL

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OSHA : Table 2
NIOSH: Table 2
ACGIH: Table 2

PROPERTIES: Table 3

SYNONYMS: Table 4

SAMPLING	MEASUREMENT
<p>SAMPLER: FILTER/SOLID SORBENT TUBE (OVS-2 tube: 13-mm quartz filter; XAD-2, 270 mg/140 mg)</p> <p>FLOW RATE: 0.2 to 1 L/min</p> <p>VOL-MIN: 12 L -MAX: 240 L; 60 L (Malathion, Ronnel)</p> <p>SHIPMENT: cap both ends of tube</p> <p>SAMPLE STABILITY: at least 10 days at 25 °C at least 30 days at 0 °C</p> <p>BLANKS: 2 to 10 field blanks per set</p>	<p>TECHNIQUE: GC, FLAME PHOTOMETRIC DETECTION (FPD)</p> <p>ANALYTE: organophosphorus pesticides, Table 1</p> <p>EXTRACTION: 2-mL 90% toluene/10% acetone solution</p> <p>INJECTION VOLUME: 1-2 µL</p> <p>TEMPERATURE -INJECTION: 240 °C -DETECTOR: 180 °C to 215 °C (follow manufacturer's recommendation) -COLUMN: Table 6</p> <p>CARRIER GAS: He at 15 psi (104 kPa)</p> <p>COLUMN: fused silica capillary column; Table 6</p> <p>DETECTOR: FPD (phosphorus mode)</p> <p>CALIBRATION: standard solutions of organophosphorus compounds in toluene</p> <p>RANGE: Table 8, Column C</p> <p>ESTIMATED LOD: Table 8, Column F</p> <p>PRECISION (\bar{S}_p): Table 5, Column E</p>
ACCURACY	
<p>RANGE STUDIED: Table 5, Column A</p> <p>ACCURACY: Table 5, Column B</p> <p>BIAS: Table 5, Column C</p> <p>OVERALL PRECISION (\hat{S}_{τ}): Table 5, Column D</p>	
<p>APPLICABILITY: The working ranges are listed in Table 5. They cover a range of 1/10 to 2 times the OSHA PELs. This method also is applicable to STEL measurements using 12-L samples. This method may be applicable to the determination of other organophosphorus compounds after evaluation for desorption efficiency, sample capacity, sample stability, and precision and accuracy.</p>	
<p>OTHER METHODS: This method may be used to replace previous organophosphorus pesticide methods. See Table 10 for partial listing. The OVS-2 tube is similar in concept to the device of Hill and Arnold [11], but offers greater convenience and lower flow resistance.</p>	
<p>INTERFERENCES: Several organophosphates may co-elute with either target analyte or internal standard causing integration errors. These include other pesticides (see Table 7), and the following: tributyl phosphate (plasticizer), tris-(2-butoxy ethyl) phosphate (plasticizer used in some rubber stoppers), tricresyl phosphate (petroleum oil additive, hydraulic fluid, plasticizer, flame-retardant, and solvent), and triphenyl phosphate (plasticizer and flame-retardant in plastics, lacquers, and roofing paper).</p>	

REAGENTS:

1. Organophosphorus analytes listed in Table 1. and (optional) triphenyl phosphate, analytical standard grade.*
2. Toluene, pesticide analytical grade.*
3. Acetone, ACS reagent grade or better.*
4. Desorbing solution. Add 50 mL acetone to a 500-mL volumetric flask. Dilute to volume with toluene.
NOTE: For optional internal standard, add 1 mL of a 5 mg/mL solution of triphenyl phosphate in toluene to 500 mL desorbing solution.
5. Organophosphorus stock solutions, 10 mg/mL. Prepare individual standard stock solutions of each pesticide of interest in 90/10 toluene/acetone (V/V). All pesticides in Table 1 were found to be soluble to at least 10 mg/mL.
6. Spiking solutions for calibration (step 9) and media fortification (steps 10, 11).
NOTE: Spiking solutions may contain more than one analyte.
 - a. Spiking solution SS-1: Dilute the volume of stock solution indicated in column F of Table 11 to 10 mL with toluene or 90/10 toluene/acetone.
 - b. Spiking solution SS-2: Dilute 1 mL of SS-1 solution with toluene in a 10-mL volumetric flask.
7. Purified gases: Helium, hydrogen, nitrogen, dry air, and oxygen, (if required by detector).

* See Special Precautions

SPECIAL PRECAUTIONS: Organophosphorus compounds are highly toxic. Special care must be taken to avoid inhalation or skin contact through the wearing of gloves and suitable clothing when handling pure material [13-17].

Toluene is flammable and toxic. Acetone is highly flammable. Prepare all samples in a well ventilated hood.

EQUIPMENT:

1. Sampler: glass tube, 11-mm ID x 13-mm OD x 50 mm long, with the outlet end drawn to a 6-mm o.d. x 25 mm long tube. The enlarged part of the tube contains a 270-mg front section of 20/60 mesh XAD-2 sorbent or equivalent held in place by a 9 to 10-mm o.d. quartz fiber filter and polytetrafluoroethylene (PTFE) retaining ring. The front section is separated from the back section of 140 mg XAD-2 sorbent or equivalent with a short plug of polyurethane foam. The back section is held in place by a long plug of polyurethane foam. The tube is available commercially as the OVS-2 sampler. See Figure 2.
NOTE: Some OVS-2 tubes contain glass fiber filters, as specified in the OSHA methods (see Table 10). These tubes, however, did not perform as well for the more polar analytes (amides, phosphoramides, and sulfoxides; see Table 9). Low or erratic recoveries for Malathion may be encountered with glass fiber filters.
2. Personal sampling pump, 0.2 to 1 L/min. with flexible connecting tubing, preferably silicon, polyethylene, or PTFE tubing.
3. Vials, 4-mL with PTFE-lined cap; 2-mL GC autosampler vials with PTFE-lined crimp caps.
4. Gas chromatograph, flame photometric detector with 525-nm bandpass filter for phosphorus mode, integrator, and column (Table 6).
5. Syringes, 5-mL and 100-, 50-, and 10-mL for making standard solutions and GC injections.
6. Volumetric flasks, 500-, 10-, and 2-mL.
7. Tweezers.
8. GC vial crimper.
9. Small ultrasonic cleaning bath.

SAMPLING:

1. Calibrate each personal sampling pump with a representative sampler in line.
 2. Connect the sampler to personal sampling pump with flexible tubing. The sampler should be placed vertically with the large end down, in the worker's breathing zone in such a manner that it does not impede work performance. [4, 12]
3. Sample at an accurately known flowrate between 0.2 and 1 L/min for a total sample size of 12 to 240 L.

4. Cap both ends of the sampler with plastic caps and pack securely for shipment.

SAMPLE PREPARATION:

5. Remove cap from large end and remove PTFE retainer ring; transfer filter and front XAD-2 section to a 4-mL vial. Transfer the short polyurethane foam plug along with back-up XAD-2 section to a second 4-mL vial.
6. Add 2 mL of desorbing solvent to each vial using a 5-mL syringe or 2-mL pipette. Cap each vial.
7. Allow to stand 30 minutes, immerse vials approximately 15 mm in an ultrasonic bath for 30 minutes. Alternatively, place the vials in a shaker or tumbler for 1 hour.
8. Transfer 1 to 1.5 mL from each 4-mL vial to a clean 2-mL GC vial, cap and label.

CALIBRATION AND QUALITY CONTROL:

9. Calibrate daily with at least six working standards covering the analytical range of the method for individual analytes.
 - a. Add known amounts of calibration spiking solution (SS-1 or SS-2 according to schedule in Table 11) to desorbing solution in 2-mL volumetric flasks and dilute to the mark.
NOTE: If an internal standard is included in the desorbing solution, then exactly 2 mL of desorbing solution in a volumetric flask must be concentrated slightly under a gentle stream of nitrogen in order to accommodate the specified volume of the spiking solutions. After adding the spiking solutions to the slightly concentrated desorbing solution, dilute to the 2-mL mark with toluene or 90/10 toluene/acetone.
 - b. Include a calibration blank of unspiked desorbing solution.
 - c. Analyze together with field samples, field blanks, and laboratory control samples (steps 12 and 13).
 - d. Prepare calibration graph (peak area vs. μg analyte), or if internal standard (IS) is used (peak area of analyte/peak area of IS vs. μg analyte).
10. Prepare Laboratory Control Samples (LCS) with each sample set, in duplicate.
 - a. Remove cap from large end of sampler tube. Apply 30 μL of spiking solution SS-1 to face of quartz fiber filter. Cap and allow to stand for a minimum of 1 hour. Preferably, these should be prepared as soon as samples arrive and should be stored with the field samples until analyzed.
 - b. Include an unspiked sampler as a media blank.
 - c. Analyze along with field samples and blanks, and liquid calibration standards (steps 12 through 16).
11. When extending application of this method to other organophosphorus compounds, the following minimal desorption efficiency (DE) test may be performed as follows:
 - a. Determine the NIOSH REL, OSHA PEL, or ACGIH TLV in mg/m^3 .
 - b. Prepare spiking solution SS-1 (refer to Table 11, or use the following formulae, which are specific for the calculation of the weight of analyte to add to 10 mL toluene/acetone 90:10).
For $\text{REL} > 1 \text{ mg}/\text{m}^3$ (assuming 12-L collection vol.), let $W = \text{REL} \times 4 \text{ m}^3$
For $\text{REL} \leq 1 \text{ mg}/\text{m}^3$ (assuming 120-L collection vol.), let $W = \text{REL} \times 40 \text{ m}^3$
where $W =$ weight (mg) of analyte to dissolve into 10 mL of desorbing solvent.
Let $[\text{SS-1}] = W/10 \text{ mL}$ where $[\text{SS-1}] =$ concentration of spiking solution SS-1 in mg/mL .
Let $[\text{SS-2}] = [\text{SS-1}] \times 0.1$ where $[\text{SS-2}] =$ concentration of spiking solution SS-2.
 - d. Prepare three tubes at each of five levels plus three media blanks. Concentration at each level may be calculated using formulae in entry 20, part II of Table 11.
 - i. Remove plastic cap from large end of sampler, apply appropriate volume of spiking solution to face of quartz fiber filter following schedule in part I of Table 11.
 - ii. Cap and allow sampler to stand overnight.
 - e. Prepare tubes for analysis (Steps 5 through 8).
 - f. Analyze with liquid standards (Steps 12 and 13).

- g. Prepare a graph of desorption efficiency (DE) vs. µg of analyte.
- h. Acceptable desorption criteria for 6 replicates is >75% average recovery with a standard deviation of <±9%.

MEASUREMENT:

12. Set gas chromatograph according to manufacturer's recommendations and to conditions listed in Table 6 and on page 5600-1. Inject sample aliquot manually using solvent flush technique or with autosampler. See Table 7 for retention times of selected analytes.

NOTE: If peak area is greater than the linear range of the working standards, dilute with desorbing solution or with desorbing solution (containing internal standard) and reanalyze. Apply the appropriate dilution factor in calculations.

13. Measure peak area of analyte and of internal standard.

CALCULATIONS:

14. Determine the mass in µg (corrected for DE) of respective analyte found in the sample front (W_f) and back (W_b) sorbent sections, and in the media blank front (B_f) and back (B_b) sorbent sections.

NOTE: The filter is combined with the front section. If $W_b > W_f/10$, report breakthrough and possible sample loss.

15. Calculate concentration, C, of analyte in the air volume sampled, V (L):

$$C = \frac{(W_f + W_b - B_f - B_b)}{V}, \text{ mg/m}^3.$$

CONFIRMATION:

16. Whenever an analyte is detected, and its identity is uncertain, confirmation may be achieved by analysis on a second column of different polarity. If primary analysis was performed using a non-polar or weakly polar column (DB-1 or DB-5), confirmation should be accomplished by reanalysis on a polar column (DB-1701 or DB-210). See Table 7 for approximate retention times for each column type. Fewer analytes co-elute on DB-210 than on DB-1701. Relative retention times are more convenient for the identification of unknown analytes. If Parathion is not used as the retention time reference compound, then another related compound such as tributyl phosphate, Ronnel, or triphenyl phosphate may be substituted.

EVALUATION OF METHOD:

This method was evaluated over the ranges specified in Table 5 at 25 °C using 240-L air samples. Sampler tubes were tested at 15% and 80% relative humidity and at 10 °C and 30 °C. In these tests, test atmospheres were not generated; instead, analytes were fortified on the face of the sampler filters. This was followed by pulling conditioned air at 1 L/min. for 4 hours. No difference in sampler performance was noted at any of these temperature/humidity combinations. Evaluations of sampler precision and stability were conducted at 30 °C and 15% relative humidity. Overall sampling and measurement precisions, bias, accuracy, and average percent recovery after long-term storage are presented in Table 5. No breakthrough was detected after 12 hours of sampling at 1 L/min with a sampler fortified with the equivalent of 4x the NIOSH REL. Malathion and Ronnel were tested at 1/40 x REL, Sulprofos at 1/20 x REL (See Table 5, note 4). All criteria [9] were met.

REFERENCES:

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METHOD WRITTEN BY:

John M. Reynolds and Don C. Wickman, DataChem Laboratories, Salt Lake City, UT.

Table 1. Formula and Registry Numbers

Compound (alphabetically)	MW ⁽¹⁾ (Daltons)	Empirical Formula	Structural Formula # RECS ⁽²⁾
1. Azinphos methyl	317.32	C ₁₀ H ₁₂ N ₃ O ₃ PS ₂	(CH ₃ O) ₂ P(=S)SCH ₂ (C ₇ H ₄ N ₃ O)86-50-0TE1925000
2. Chlorpyrifos	350.58	C ₉ H ₁₁ Cl ₃ NO ₃ PS	(C ₂ H ₅ O) ₂ P(=S)O(C ₅ HN)Cl ₃ 2921-88-2 TF6300000
3. Diazinon	304.34	C ₁₂ H ₂₁ N ₂ O ₃ PS	(C ₂ H ₅ O) ₂ P(=S)O(C ₄ HN ₂)(CH ₃)CH(CH ₃) ₂ 333-41-5 TF3325000
4. Dicrotophos	237.19	C ₈ H ₁₆ NO ₅ P	(CH ₃ O) ₂ P(=O)OC(CH ₃)=CHC(=O)N(CH ₃) ₂ 141-66-2 TC3850000
5. Disulfoton	274.39	C ₈ H ₁₉ O ₂ PS ₃	(C ₂ H ₅ O) ₂ P(=S)S(CH ₂) ₂ SC ₂ H ₅ 298-04-4 TD9275000
6. Ethion	384.46	C ₉ H ₂₂ O ₄ P ₂ S ₄	[(C ₂ H ₅ O) ₂ P(=S)S] ₂ CH ₂ 563-12-2 TE4550000
7. Ethoprop	242.33	C ₈ H ₁₉ O ₂ PS ₂	(C ₃ H ₇ S) ₂ P(=O)OC ₂ H ₅ 13194-48-4 TE4025000
8. Fenamiphos	303.36	C ₁₃ H ₂₂ NO ₃ PS	(CH ₃) ₂ CHNHP(=O)(O[C ₂ H ₅])O(C ₆ H ₃)(CH ₃)SCH ₃ 22224-92-6 TB3675000
9. Fonofos	246.32	C ₁₀ H ₁₅ OPS ₂	C ₂ H ₅ OP(C ₂ H ₅)(=S)S(C ₆ H ₅)944-22-9 TA5950000
10. Malathion	330.35	C ₁₀ H ₁₉ O ₆ PS ₂	(CH ₃ O) ₂ P(=S)SCH[C(=O)OC ₂ H ₅]CH ₂ C(=O)OC ₂ H ₅ 121-75-5 WM8400000
11. Methamidophos	141.12	C ₂ H ₈ NO ₂ PS	CH ₃ OP(=O)(NH ₂)SCH ₃ 10265-92-6 TB4970000
12. Methyl parathion	263.20	C ₈ H ₁₀ NO ₅ P	(CH ₃ O) ₂ P(=S)O(C ₆ H ₄)NO ₂ 298-00-0 TG0175000
13. Mevinphos (E)	224.15	C ₇ H ₁₃ O ₆ P	(CH ₃ O) ₂ P(=O)OC(CH ₃)=CHC(=O)OCH ₃ 298-01-1 ⁽²⁾ GQ5250100
Mevinphos (E & Z)			7786-34-7 ^(3,4) GQ5250000
14. Monocrotophos (Z)	223.17	C ₇ H ₁₄ NO ₅ P	(CH ₃ O) ₂ P(=O)OC(CH ₃)=CHC(=O)NHCH ₃ 919-44-8 ⁽²⁾ TC4981100
Monocrotophos (E)			6923-22-4 ^(3,4) TC4375000
15. Parathion	291.26	C ₁₀ H ₁₄ NO ₅ PS	(C ₂ H ₅ O) ₂ P(=S)O(C ₆ H ₄)NO ₂ 56-38-2 TF4550000
16. Phorate	260.36	C ₇ H ₁₇ O ₂ PS ₃	(C ₂ H ₅ O) ₂ P(=S)SCH ₂ SC ₂ H ₅ 298-02-2 TD9450000
17. Ronnel	321.54	C ₈ H ₈ Cl _x O ₃ PS	(CH ₃ O) ₂ P(=S)O(C ₆ H ₂)Cl ₃ 299-84-3 TG0525000
18. Sulprofos	322.43	C ₁₂ H ₁₉ O ₂ PS ₃	C ₂ H ₅ OP(S[C ₃ H ₇])(=S)O(C ₆ H ₄)SCH ₃ 35400-43-2 TE4165000
19. Terbufos	288.42	C ₉ H ₂₁ O ₂ PS ₃	(C ₂ H ₅ O) ₂ P(=S)SCH ₂ SC(CH ₃) ₃ 13071-79-9 TD7740000

- (1) Molecular weights calculated from the empirical formula using 1979 IUPAC Atomic Weights of the Elements.
- (2) RTECS = NIOSH Registry of Toxic Effects of Chemical Substances [1].
- (3) Merck Index [2].
- (4) Farm Chemicals Handbook [3].

Table 2. Toxicity and Maximum Exposure Limits

Compound STEL (alphabetically)	LD₅₀, mg/Kg⁽¹⁾	OSHA PEL⁽⁴⁾	NIOSH REL⁽⁵⁾		ACGIH TLV
		mg/m³	mg/m³	(ppm)	mg/m³/mg/m³
1. Azinphos methyl	11 f	0.2	0.2	(0.015)	0.2 (skin)
2. Chlorpyrifos	145	0.2	0.2	(0.014)	0.2 (skin)
0.6 ⁽⁵⁾					
3. Diazinon	250 m, 285 f	0.1	0.1	(0.008)	0.1 (skin)
4. Dicrotophos	16 f, 21 m	0.25	0.25	(0.026)	0.25 (skin)
5. Disulfoton	2.3 f, 6.8 m	0.1	0.1	(0.009)	0.1 ⁽³⁾ (skin)
6. Ethion	27 f, 65 m	0.4	0.4	(0.025)	0.4 (skin)
7. Ethoprop	61.5 ⁽²⁾				
8. Fenamiphos	10	0.1	0.1	(0.008)	0.1 (skin)
9. Fonofos	3 f, 13 m ⁽³⁾	0.1	0.1	(0.010)	0.1 (skin)
10. Malathion	1000 f, 1375 m	10	10	(0.740)	10 (skin)
11. Methamidophos	25 m, 27 f				
12. Methyl parathion	14 m, 24 f	0.2	0.2	(0.019)	0.2 (skin)
13. Mevinphos	3.7 f, 6.1 m	0.1	0.1	(0.011)	0.1 (skin)
0.27 ⁽⁶⁾					
14. Monocrotophos (skin)	17 m, 20 f	0.25	0.25	(0.027)	0.25 ⁽³⁾
15. Parathion	3.6 f, 13 m	0.1	0.05	(0.004)	0.1 (skin)
16. Phorate	1.1 f, 2.3 m	0.05	0.05	(0.005)	0.05 (skin)
0.2 ^(5,6)					
17. Ronnel	1250 m, 2630 f	10	10	(0.760)	10 (skin)
18. Sulprofos	227	1	1	(0.076)	1 ⁽³⁾ (skin)
19. Terbufos	1.6-4.5 m, 9.0 f				

- (1) Rat-oral; from Merck Index, unless otherwise noted, f = female, m = male [2].
- (2) Farm Chemicals Handbook [3].
- (3) RTECS [1].
- (4) OSHA Final Rule, 1989 (unenforceable, 1992); only Malathion and Parathion had previous PELs.
- (5) NIOSH Recommendations for Occupational Safety and Health [5].
- (6) ACGIH [18].

Table 3. Physical Properties⁽¹⁾

Compound Solubility in Water (alphabetically) at 20 °C)	Liquid Density (g/mL)	MP (°C)	BP °C (mm Hg)	Vapor Pressure		
				<u>Pascal</u>	<u>mm Hg @ °C</u> (mg/L)	
1. Azinphos methyl 20 ⁽³⁾	1.44	73-74 30 ⁽⁸⁾	unstable >200	0.00018	1.35x10 ⁻⁶	
2. Chlorpyrifos —	none found	41-42	—	0.0025	1.87x10 ⁻⁵	25
3. Diazinon 40	1.116-1.118	liquid	decomp. >120	0.019	1.4x10 ⁻⁴	20
4. Dicrotophos miscible	1.216	liquid	400 (760)		none found	
5. Disulfoton nearly insol. 20 ⁽³⁾	1.144	oil insol. ⁽³⁾	—	0.024 0.0074	1.8x10 ⁻⁴ 5.4x10 ⁻⁵	20
6. Ethion slightly sol.	1.220	-12 to -13	—	0.0002	1.5x10 ⁻⁶	
7. Ethoprop 750	1.094	oil	—	0.047	3.5x10 ⁻⁴	26
8. Fenamiphos 20 ⁽³⁾	none found	49 329	—	0.00012	9x10 ⁻⁷	
9. Fonofos 13 ⁽⁷⁾	1.16	liquid	—		none found	
10. Malathion 30 ⁽⁶⁾	1.23 145	2.9	156 (0.7) ⁽⁶⁾	0.005	4x10 ⁻⁵	
11. Methamidophos readily sol. 20 ⁽³⁾	1.31	54.	—	0.04 0.0023	3x10 ⁻⁴ 1.7x10 ⁻⁵	30
12. Methyl parathion 20 ⁽³⁾	— 1.358 50	37-38	—	0.0002	1.5x10 ⁻⁶	
13. Mevinphos 20 ⁽⁴⁾	1.25 miscible	20.6 ⁽⁴⁾	325 (760) ⁽⁴⁾	0.4 0.29	3x10 ⁻³ 2.2x10 ⁻³	
14. Monocrotophos 20 ⁽⁷⁾	— none found	54-55 ⁽⁵⁾	—	0.0009	7x10 ⁻⁶	20

miscible						
15. Parathion 20 ⁽⁶⁾	1.26 20	6	375 (760)	0.005	3.78x10 ⁻⁵	
				0.00089	6.7x10 ⁻⁶	
20 ⁽³⁾	10 ⁽⁸⁾					
16. Phorate 50	1.156	liquid	118-120 (2.0) ⁽³⁾	0.11	8.4x10 ⁻⁴	20
17. Ronnel 40 @ 25 °C	sp. gr=1.48 ⁽²⁾	41	—	0.1	8.x10 ⁻⁴	25
18. Sulprofos 20 ⁽³⁾	1.20 nearly insol. ⁽³⁾	liquid	210 (0.1) ⁽⁶⁾	<0.0001	<10 ⁻⁶	
19. Terbufos 10-15	1.105	-29.2	—		none found	

(1) From Merck Index, unless otherwise noted [2].
°C for commercial mixture.

(2) NIOSH 2nd Edition Method for Ronnel, S299 [7].
(EPN, Malathion, Parathion) [8].

(3) Farm Chemicals Handbook [3].
specific analyte Method) [4].

(4) NIOSH 3rd Edition Method 2503 for Mevinphos [8].

(5) 54-55 °C for pure material, 25-30

(6) NIOSH 3rd Edition Method 5012

(7) OSHA Stopgap Methods (See

(8) NIOSH Pocket Guide [6].

Table 4. Synonyms

Compound⁽¹⁾ (alphabetically)	Other name⁽²⁾	CAS Name
1. Azinphos methyl S-[(4-oxo-1,2,3-benzotriazin-3(4H)-yl)methyl] ester ⁽³⁾	Guthion*	Phosphorodithioic acid, O,O-dimethyl
2. Chlorpyrifos O-(3,5,6-trichloro-2-pyridinyl) ester ⁽³⁾	Dursban*	Phosphorothioic acid, O,O-diethyl
3. Diazinon O-[6-methyl-2-(1-methylethyl)-4-pyrimidinyl] ester ⁽³⁾	Spectracide*	Phosphorothioic acid, O,O-diethyl
4. Dicrotophos 3-(dimethylamino)-1-methyl-3-oxo-1-propenyl dimethyl ester ⁽³⁾	Bidrin*	Phosphoric acid, Phosphoric acid, dimethyl ester, ester with
cis-3-hydroxy-N,N-dimethylcrotonamide ⁽⁴⁾		
5. Disulfoton S-[2-(ethylthio)ethyl] ester ⁽³⁾	Di-Syston*	Phosphorodithioic acid, O,O-diethyl
6. Ethion O,O,O',O'-tetraethyl ester ⁽³⁾		Phosphorodithioic acid, S,S'-methylene O,O,O',O'-Tetraethyl
S,S'-methylenediphosphorodithioate ⁽⁴⁾		
7. Ethoprop	Prophos*	Phosphorodithioic acid, O-ethyl S,S-dipropyl ester ⁽³⁾
8. Fenamiphos 3-methyl-4-(methylthio)phenyl ester ⁽³⁾	Nemacur*, Phenamiphos ⁽¹⁾	(1-Methylethyl)phosphoramidic acid, ethyl Phosphoramidic acid, isopropyl-,
4-(methylthio)-m-tolyl ethyl ester ⁽⁴⁾		
9. Fonofos ester ⁽³⁾	Dyfonate*	Ethyl phosphonodithioic acid, O-ethyl S-phenyl Phosphonodithioic acid, ethyl-, O-ethyl S-phenyl
ester ⁽⁴⁾		
10. Malathion diethyl ester ⁽³⁾	Cythion*	[(Dimethoxyphosphinothioyl)thio]butanedioic acid Succinic acid, mercapto-, diethyl ester, S-ester with
O,O-dimethylphosphorodithioate ⁽⁴⁾		
11. Methamidophos	Monitor*	Phosphoramidothioic acid, O,S-dimethyl ester ⁽³⁾
12. Methyl parathion O-(4-nitrophenyl) ester ⁽³⁾	Parathion Methyl ⁽¹⁾	Phosphorothioic acid, O,O-dimethyl
13. Mevinphos	Phosdrin*	3-[(Dimethoxyphosphinyl)oxy]-2-butenic acid,

methyl ester ⁽³⁾		Crotonic acid, 3-hydroxy-, methyl ester dimethyl
phosphate ⁽⁴⁾		
14. Monocrotophos	Azodrin*	Phosphoric acid, dimethyl
[1-methyl-3-(methylamino)-3-oxo-1-propenyl] ester ⁽³⁾		Phosphoric acid, dimethyl ester, ester with
(E)-3-hydroxy-N-methylcrotonamide ⁽⁴⁾		
15. Parathion	Ethyl Parathion ⁽¹⁾	Phosphorothioic acid, O,O-diethyl O-(4-nitrophenyl)
ester ⁽³⁾		
16. Phorate	Thimet*	Phosphorodithioic acid, O,O-diethyl
S-[(ethylthio)methyl] ester ⁽³⁾		
17. Ronnel	Fenclorphos ⁽¹⁾	Phosphorothioic acid, O,O-dimethyl
O-(2,4,5-trichlorophenyl) ester ⁽³⁾		
18. Sulprofos	Bolstar*	Phosphorodithioic acid, O-ethyl
O-[4-(methylthio)phenyl] S-propyl ester ⁽⁴⁾		
19. Terbufos	Counter*	Phosphorodithioic acid, O,O-diethyl
S-[[[(1,1-dimethylethyl)thio]methyl] ester ⁽⁴⁾		

- (1) Common name as given in Farm Chemicals Handbook [3].
 (2) *=Trade name (Trademark or Registered Name) as given in Farm Chemicals Handbook [3].
 (3) Merck Index [2].
 (4) RTECS [1] or alternate CAS name in Merck Index [2].

Table 5. Method Evaluation⁽¹⁾

Compound (alphabetically) days	A. Range Studied ⁽²⁾		B. Accuracy	CDEG.	
	mg/m ³	mg/sample		Bias Ave.	Precision Range
25 °C (0 °C)					
1. Azinphos methyl 0.070	0.02-0.4 0.030	0.0048-0.096 97 (105)	± 0.178	-0.038 (-0.120 - +0.028)	
2. Chlorpyrifos 0.068	0.02-0.4 0.018	0.0048-0.096 92 (90)	± 0.163	-0.027 (-0.054 - +0.017)	
3. Diazinon 0.065	0.01-0.2 0.020	0.0024-0.048 94 (93)	± 0.162	-0.032 (-0.057 - -0.005)	
4. Dicrotophos 0.066	0.025-0.5 0.025	0.006-0.120 89 (92)	± 0.169	-0.037 (-0.102 - -0.032)	
5. Disulfoton 0.066	0.01-0.2 0.024	0.0024-0.048 87 (89)	± 0.196	-0.064 (-0.081 - -0.032)	
6. Ethion 0.068	0.04-0.8 0.018	0.0096-0.192 96 (95)	± 0.165	-0.029 (-0.056 - -0.003)	
7. Ethoprop ⁽³⁾ 0.066	0.01-0.2 0.024	0.0024-0.048 97 (93)	± 0.157	-0.025 (-0.058 - +0.025)	
8. Fenamiphos 0.063	0.01-0.2 0.022	0.0024-0.048 94 (96)	± 0.155	-0.029 (-0.066 - +0.002)	
9. Fonofos 0.066	0.01-0.2 0.023	0.0024-0.048 95 (92)	± 0.168	-0.036 (-0.076 - +0.008)	
10. Malathion ⁽⁴⁾ 0.067	0.025-0.5 0.019	0.006-0.120 93 (93)	± 0.172	-0.038 (-0.064 - -0.014)	
11. Methamidophos ⁽⁵⁾ 0.069	0.02-0.4 0.026	0.0048-0.096 88 (95)	± 0.156	-0.018 (-0.046 - +0.011)	
12. Methyl parathion 0.063	0.02-0.4 0.018	0.0048-0.096 95 (95)	± 0.160	-0.034 (-0.082 - +0.016)	
13. Mevinphos 0.067	0.01-0.2 0.028	0.0024-0.048 89 (91)	± 0.176	-0.042 (-0.061 - -0.004)	
14. Monocrotophos 0.071	0.025-0.5 0.026	0.006-0.120 88 (92)	± 0.185	-0.043 (-0.047 - -0.020)	
15. Parathion 0.071	0.005-0.1 0.019	0.0012-0.024 92 (92)	± 0.163	-0.021 (-0.045 - +0.011)	

16. Phorate	0.005-0.1	0.0012-0.024	± 0.202	-0.070 (-0.097 - -0.047)
0.066	0.025	91 (91)		
17. Ronnel ⁽⁴⁾	0.025-0.5	0.006-0.120	± 0.172	-0.040 (-0.076 - +0.021)
0.066	0.018	95 (94)		
18. Sulprofos ⁽⁴⁾	0.01-0.2	0.0024-0.048	± 0.181	-0.047 (-0.054 - -0.031)
0.067	0.017	94 (94)		
19. Terbufos ⁽³⁾	0.01-0.2	0.0024-0.048	± 0.188	-0.054 (-0.091 - -0.024)
0.067	0.022	92 (91)		

- (1) NIOSH Back-up Data Report [9]
- (2) The ranges studied were 1/10 to 2x the NIOSH REL (except as noted) sampling at 1 L/min for 4 hours.
- (3) No NIOSH REL or OSHA PEL available; used 0.1 mg/m³.
- (4) Malathion and Ronnel were studied at 1/400 to 1/20 the NIOSH REL, Sulprofos at 1/200 to 1/10 the NIOSH REL.
- (5) No NIOSH REL or OSHA PEL available; used 0.2 mg/m³.

Table 6. Recommended Gas Chromatographic Columns and Conditions⁽⁶⁾

Parameter	Wide Bore Fused Silica Capillary Column			
Stationary Phase ⁽²⁾	DB-1	DB-5	DB-1701	DB-210
Polarity	non-polar	weakly polar	mod. polar	mod. polar
Length (meters)	30	30	30	30
I.D. (millimeters)	0.32	0.32	0.32	0.32
Film thickness (mm) ⁽³⁾	0.25	1.0	1.0	0.25
Injection (vol., mode) ⁽⁴⁾⁽⁵⁾	1 μ L, SPL	1 μ L, DIR	1 μ L, DIR	1 μ L, SPL
Oven Temperatures				
Initial ($^{\circ}$ C)	100	125	125	100
Final ($^{\circ}$ C)	275	275	275	250
Max. Recommended ($^{\circ}$ C) ⁽⁵⁾	325	325	280	240/260
Program ($^{\circ}$ C/min.)	3.0	4.0	4.0	3.0
Carrier Gas (Helium)				
Head pressure (p.s.i.)	15	15	15	15

- (1) Actual conditions may vary depending on column and analytical objectives. The conditions given above are those corresponding to the RT data in Table 7.
- (2) DB-1, 100% methyl silicone; DB-5, 5% phenyl, 95% methyl silicone; DB-1701, 14% cyanopropylphenyl, 86% methyl silicone; DB-210, 50% trifluoropropyl, 50% methyl silicone. Other phase types may also work well.
- (3) Thinner films give faster separations at lower temperatures promoting analyte stability.
- (4) Use 2-mm i.d. injection port lines for 0.5- μ L injection and 4-mm i.d. injection port liners for 1-to-2- μ L injections with 0.32-mm i.d. capillary columns.
- (5) SPL = splitless mode, initial oven temperature 5-10 $^{\circ}$ C < BP of desorption solvent;
DIR = direct mode, initial oven temperature 5-10 $^{\circ}$ C > BP of desorption solvent.
In both modes, split-vent off time should be 60 sec for 1-2 μ L injections with 4-mm ID injection port liners, and 20-30 sec for 0.5- μ L injections with 2-mm ID injection port liners. Use 2-mm ID injection port liners for 0.5- μ L injection and 4-mm ID injection port liners for 1-2 μ L injections with 0.32-mm ID capillary columns.
- (6) J & W Scientific Catalog, p. 21. [10]

Table 7. Approximate Retention Times of Selected Organophosphorous Compounds⁽¹⁾

Compound (by RT on DB-1)	Capillary Column ⁽²⁾					
	DB-1			DB-5	DB-1701	DB-210
	RT min	RRT ⁽³⁾	Elution T, °C ⁽⁴⁾	RT, min	RT, min	RT, min
1. TEPP	3.71	0.128	111	5.47	7.18 ^(B)	7.88
2. Triethylphosphorothioate	4.37	0.151	113	6.34	7.14 ^(B)	4.93
3. Methamidophos	5.12	0.177	115	7.64	13.61	12.03
4. Dichlorvos	5.81	0.200	117	8.24	10.67	10.54
5. Mevinphos	10.45	0.360	131	12.92	16.69	19.20
6. Ethoprop	17.15	0.592	151	19.09	21.52	20.10
7. Naled	17.61	0.608	153	no data	23.17 ^(C)	21.46 ^(H)
8. Dicrotophos	18.00	0.621	154	19.94	25.84 ^(E)	31.43
9. Monocrotophos	18.27	0.630	155	20.12	28.11	31.60
10. Sulfotepp	19.06	0.658	157	no data	23.09 ^(C)	21.11
11. Phorate	19.18	0.662	158	20.94	23.10 ^(C)	18.92
12. Dimethoate	19.44	0.671	158	21.84	no data	29.33 ^(I)
13. Demeton-S	20.15	0.695	160	21.70	25.06 ^(D)	24.97
14. Dioxathion	21.30	0.735	164	23.04	26.33 ^(F)	23.46
15. Fonofos	22.04	0.760	166	23.57	25.87 ^(E)	22.20
16. Terbufos	22.22	0.767	168	23.80	25.02 ^(D)	21.52 ^(H)
17. Disulfoton	23.09	0.797	169	24.19	26.43 ^(F)	22.78
18. Diazinon	23.37	0.806	170	23.75	25.00 ^(D)	20.99
19. Methyl parathion	25.37	0.875	176	26.48	31.37	33.21
20. Oxydemeton methyl	26. ⁽⁵⁾	0.90	179	no data	no data	no data
21. Ronnel	26.86	0.927	181	27.39	29.30	26.27
22. Pirimiphos methyl	28.13	0.971	184	27.90	29.72	26.77
23. Malathion	28.53	0.984	186	28.33	31.78 ^(G)	33.08 ^(J)
24. Fenthion	28.74	0.992	186	28.93	31.78 ^(G)	29.35 ^(I)
25. Parathion	28.98	1.000	187	29.10 ^(A)	33.28	35.60
26. Chlorpyrifos	29.11	1.004	187	29.10 ^(A)	30.79	27.72
27. Cruformate	29.64	1.023	189	29.54	34.00	35.34
28. Isofenphos	31.91	1.101	196	31.17	33.81	33.02 ^(J)
29. Tetrachlorvinphos	33.26	1.148	200	32.60	35.96	37.01
30. Fenamiphos	34.09	1.176	202	33.03	37.14	38.95
31. Merphos	35.19	1.214	206	no data	30.57	23.89
32. Fensulfothion	36.61	1.263	210	35.78	42.41	46.98
33. Ethion	37.88	1.307	214	36.30	39.30	37.96
34. Sulprofos	38.49	1.328	216	36.96	39.54	37.11
35. Triphenyl phosphate	40.88	1.411	223	39.06	no data	no data
36. EPN	42.64	1.471	228	41.06	47.83	47.13
37. Azinphos methyl	44.16	1.524	232	43.67	no elution	49.24
38. Leptophos	45.12	1.557	235	43.91	47.38	41.68
39. Azinphos ethyl	46.55	1.606	240	46.50	47.43	50.40
40. Coumaphos	49.31	1.702	248	50.10	67.86	60.88

(1) Actual retention times (RT) will vary with individual columns and chromatographic conditions. See Table 9 for chromatographic performance notes. Capillary Column conditions given in Table 6. Data from Backup Data Report [9].

(2) Sets of co-eluting or nearly co-eluting peaks are identified by letters: (A), (B), (C), (D), (E), (F), (G), (H), (I), and (J)

(3) Retention times relative to Parathion.

(4) Elution temperature (°C) for DB-1 column (see Table 6 for column conditions.) Elution temperatures are provided for convenience in selecting approximate GC oven temperatures for isothermal analyses of single or several closely eluting analytes.

(5) Broad, tailing peak.

Table 8. Applicable Working Range and Estimated LOD

F.	G.	A.		B.	C.	D.	E.
		Atmospheric ⁽¹⁾		Sample ⁽⁴⁾	Margin of Instrument ⁽⁶⁾	Instrument ⁽⁴⁾	Atmospheric ⁽⁴⁾
Estimated LOD							
Compound Sensitivity ⁽²⁾ (alphabetically) REL/LOD		mg/m ³	ppm ⁽³⁾	µg/sample	ng on column	ng on column	µg/sample/m ³
1. Azinphos methyl 0.12		0.02-0.6 0.0012	0.0015-0.046 167	2.4 to 72	1.2-36		0.06
2. Chlorpyrifos 0.04		0.02-0.6 0.0004	0.0014-0.042 500	2.4 to 72	1.2-36		0.02
3. Diazinon 0.04		0.01-0.3 0.0004	0.0008-0.024 250	1.2 to 36	0.6-18		0.02
4. Dicrotophos 0.2		0.025-0.75 0.002	0.0026-0.077 125	3.0 to 90	1.5-45		0.1
5. Disulfoton 0.04		0.01-0.3 0.0004	0.0009-0.027 250	1.2 to 36	0.6-18		0.02
6. Ethion 0.04		0.04-1.2 0.0004	0.0025-0.076 1000	4.8 to 144	2.4-72		0.02
7. Ethoprop 0.04		0.01-0.3 0.0004	0.0010-0.030 (7)	1.2 to 36	0.6-18		0.02
8. Fenamiphos 0.14		0.01-0.3 0.0014	0.0008-0.024 71	1.2 to 36	0.6-18		0.07
9. Fonofos 0.04		0.01-0.3 0.0004	0.0010-0.030 250	1.2 to 36	0.6-18		0.02
10. Malathion 0.1 0.001	10,000	1.0-30	0.074-2.2	12. to 360 ⁽⁵⁾	6.-180 ⁽⁵⁾		0.05
11. Methamidophos 0.6 0.005	(7)	0.02-0.6	0.0035-0.10	2.4 to 72	1.2-36		0.3
12. Methyl parathion 0.04		0.02-0.6 0.0004	0.0019-0.056 500	2.4 to 72	1.2-36		0.02
13. Mevinphos 0.12		0.01-0.3 0.0012	0.0011-0.033 83	1.2 to 36	0.6-18		0.06
14. Monocrotophos 0.4 0.004	63	0.025-0.75	0.0027-0.082	3.0 to 90	1.5-45		0.2
15. Parathion 0.04		0.005-0.15 0.0004	0.0004-0.013 125	0.6 to 18	0.3-9		0.02
16. Phorate 0.04		0.005-0.15 0.0004	0.0005-0.014 125	0.6 to 18	0.3-9		0.02

17. Ronnel	1.0-30	0.076-2.3	12. to 360 ⁽⁵⁾	6.-180 ⁽⁵⁾	0.02
0.04	0.0004	25,000			
18. Sulprofos	0.1-3.0	0.0076-0.23	12. to 360	6.-180	0.03
0.06	0.0005	2000			
19. Terbufos	0.01-0.3	0.0008-0.026	1.2 to 36	0.6-18	0.02
0.04	0.0004	(7)			

- (1) To cover range of 1/10 to 3x NIOSH REL.
- (2) REL in mg/m³ (Table 2) ÷ Atmospheric LOD (Column G, Table 8).
- (3) Calculated for 25 °C and 760 mm Hg (NTP).
- (4) Calculated for a collection volume of 120 L (2 h @ 1 L/min, 4 h @ 0.5 L/min, or 10 h @ 0.2 L/min).
- (5) Calculated for a collection volume of 12 L (12 min @ 1 L/min, 24 min @ 0.5 L/min, or 1 h @ 0.2 L/min).
- (6) Desorbing sample in 2.0 mL solvent and injecting 1 µL into gas chromatograph.
- (7) No REL

Table 9. Notes on Analytical Characteristics of Organophosphorous Compounds⁽¹⁾

Compound (Alphabetically)	Analytical Characteristics		
	A Chemical and Physical	B Desorption and Solution	C Gas Chromatography
1. Azinphos methyl (Guthion*)			3,5,6
2. Azinphos ethyl (Guthion ethyl)			5
3. Chlorpyrifos (Dursban*)			
4. Coumaphos (Co-Ral*)			5
5. Crufomate (Ruelene*)	1	1,4	1
6. Demeton (Systox*)	2,6	5	3
7. Diazinon (Spectracide*)			
8. Dichlorvos (DDVP, Vapona*)	7		4
9. Dicrotophos (Bidrin*)			
10. Dimethoate (Cygon*)	1	1,4	1
11. Dioxathion (Delnav*)			
12. Disulfoton (Di-Syston*)	2		2
13. EPN (Santox*)			5
14. Ethion			
15. Ethoprop (Prophos*)			
16. Fenamiphos (Nemacur*)	1	1,4	1
17. Fensulfothion (Dasanit*)	3	4	
18. Fenthion (Baytex*)		5	
19. Fonofos (Dyfonate*)			
20. Isofenphos (Oftanol*)	1	1	1
21. Leptophos (Phosvel*)		5	5
22. Malathion (Cythion*)			
23. Merphos (Folex*)	4	5	2
24. Methamidophos (Monitor*)	1	1,3,4	1,4
25. Methyl parathion (Parathion methyl)			
26. Mevinphos (Phosdrin*)	6,7		3,4
27. Monocrotophos (Azodrin*)	1	1,2,4	1
28. Naled (Dibrom*)	5	5	2
29. Oxydemeton methyl (Metasystox-R)	3	1,5	1,2
30. Parathion (Ethyl parathion)			
31. Phorate (Thimet*)	2,7		2
32. Pirimiphos methyl (Actellic*)			4
33. Ronnel (Fenchlorphos)			
34. Sulfotepp (TEDP)			
35. Sulprofos (Bolstar*)			
36. TEPP	7	5	4
37. Terbufos (Counter*)	2		2
38. Tetrachlorvinphos (Gardona*)			
39. Tributyl phosphate			7
40. Triphenyl phosphate			7

* = Trade name, Registered name, or Trademark (Farm Chemicals Handbook [3]).

(1) Observations made during selection and validation of selected analytes [9]; refer to notes on the following page.

Table 9 (Continued) - Notes on ANALYTICAL CHARACTERISTICS**A. CHEMICAL AND PHYSICAL**

1. Amide or phosphoramidate, some slightly acidic, very polar chemically.
 2. Alkyl thioether, easily oxidized to sulfone and sulfoxide.
 3. Sulfoxides, easily oxidized to sulfone. Also very polar chemically.
 4. Phosphite, easily air oxidized to phosphate (Merphos → DEF).
 5. Vicinal dibromide, easily debrominated (Naled → Dichlorvos).
 6. Two or more isomers commonly exist (e.g. Demeton-O and Demeton-S; cis- and trans- mevinphos).
 7. Relatively volatile, can be lost if media or vials are left uncapped for even a short period of time.
- General: Organophosphorous compounds are easily destroyed at mildly alkaline conditions (pH ≥ 8). Losses can occur for trace levels of compounds on alkaline glass surfaces. Glassware should be neutralized after washing, if alkaline detergent is used.

B. DESORPTION AND SOLUTION

1. Solubility of concentrated solutions in toluene enhanced by the addition of 1% methanol or 10% acetone. Solubility in hexane very unfavorable even for dilute solutions.
 2. Changing from 100% toluene to 90/10 toluene/acetone, desorption from glass fiber filters improved from 62% to 98%, desorption from quartz fiber filters improved from 30% to 101%.
 3. Changing from glass fiber filters to quartz fiber filters, desorption in toluene improved from 16% to 88% and desorption in 90/10 toluene/acetone improved from 70% to 99%.
 4. These compounds are more chemically polar than the other listed organophosphorous compounds; desorption from XAD-2 or from glass or quartz fiber filters in hexane was incomplete or non-existent. Desorption in toluene was adequate except as noted in 2 and 3 above. The use of toluene containing 10% acetone improved recoveries for all analytes to satisfactory levels.
 5. The desorption characteristics of these compounds were not evaluated.
- General:
- a. The presence of acidic hydrogen or double bonded oxygen anywhere in the molecular structure greatly decreases solubility in non-polar solvents and increases the difficulty of desorption from polar surfaces and sorbents.
 - b. While glass fiber filters and toluene desorbant were adequate for most compounds in preliminary tests, the method was given wider application for the more polar compounds by the use of quartz fiber filters and 90/10 toluene/acetone desorbant.
 - c. Solvents with lower boiling points (e.g., methylene chloride, chloroform, methyl-t-butyl ether, and ethyl acetate) had good desorption power nearing equivalent to 90/10 toluene acetone, but rendered less satisfactory gas chromatographic responses for the analytes. This effect may be due to better analyte mass-transfer from the injection port to the capillary column with higher boiling solvents using splitless or direct injection techniques.

C. GAS CHROMATOGRAPHY

1. Poor chromatography may be encountered with dirty or undeactivated columns or injection ports. Clean quartz wool plugs stuffed in the injection port liner are better than silanized glass wool at reducing losses within the injection port.
2. Multiple, shifted, irregular, or severely tailing peaks may be observed in the chromatogram if degradation or oxidation of the analytes occur prior to injection, within the injection port, or during chromatographic separation on-column.
3. Multiple peaks may be observed due to presence of isomers.
4. Short elution time, compound may co-elute with solvent if oven temperature is too high.
5. Long elution time, compound may be lost if run time is too short, column or injection port is too cool, or split-vent-valve opens too soon when injected in splitless or direct injection mode.
6. Azinphos methyl did not elute from DB-1701 even though Azinphos ethyl did elute.
7. Potential internal standards: Triphenyl phosphate is more favorable if multiple analytes are expected because it is less volatile and elutes in an area of the chromatogram having fewer competing analytes.

Table 10. Other Methods of Analysis for Organophosphorous Compounds in Air

Document	Method Number	Organophosphorous Compound(s)																														
Hill & Arnold ⁽¹⁾		Chlorpyrifos, Demeton-O, Demeton-S, Diazinon, Dimethoate, Malathion, Paraoxon, and Parathion																														
NMAM, 2nd ed. ⁽²⁾	v. 1 P&CAM 158	Parathion																														
	v. 5 P&CAM 295	Dichlorvos (DDVP)																														
	v. 6 P&CAM 336	TEPP																														
	v. 3 S 208	Tributyl phosphate																														
	v. 3 S 209	Triorthocresyl phosphate																														
	v. 3 S 210	Triphenyl phosphate																														
	v. 6 S 280	Demeton																														
	v. 3 S 285	EPN																														
	v. 3 S 295	Parathion																														
	v. 6 S 296	Mevinphos																														
	v. 6 S 299	Ronnel																														
	v. 3 S 370	Malathion																														
NMAM, 3rd ed. ⁽³⁾	2503	Mevinphos																														
	2504	TEPP																														
	5012	EPN, Malathion, and Parathion																														
	5514	Demeton																														
OSHA ⁽⁴⁾	62	Chlorpyrifos, Diazinon, Parathion, DDVP, and Malathion																														
OSHA Stopgap ⁽⁵⁾	Each method is separate and unnumbered. Refer to by name.	<table border="0"> <tr> <td>Azinphos ethyl</td> <td>Ethoprop</td> <td>Mevinphos</td> </tr> <tr> <td>Azinphos methyl</td> <td>Fenamiphos</td> <td>Monocrotophos</td> </tr> <tr> <td>Coumaphos</td> <td>Fensulfothion</td> <td>Oxydemeton methyl</td> </tr> <tr> <td>Crufomate</td> <td>Fenthion</td> <td>Phorate</td> </tr> <tr> <td>Demeton</td> <td>Fonofos</td> <td>Pirimiphos methyl</td> </tr> <tr> <td>Dicrotophos</td> <td>Isofenphos</td> <td>Sulprofos</td> </tr> <tr> <td>Dioxathion</td> <td>Leptophos</td> <td>TEDP (sulfotepp)</td> </tr> <tr> <td>Disulfoton</td> <td>Methamidophos</td> <td>TEPP</td> </tr> <tr> <td>EPN</td> <td>Methyl dematon</td> <td>Terbufos</td> </tr> <tr> <td>Ethion</td> <td>Methyl parathion</td> <td></td> </tr> </table>	Azinphos ethyl	Ethoprop	Mevinphos	Azinphos methyl	Fenamiphos	Monocrotophos	Coumaphos	Fensulfothion	Oxydemeton methyl	Crufomate	Fenthion	Phorate	Demeton	Fonofos	Pirimiphos methyl	Dicrotophos	Isofenphos	Sulprofos	Dioxathion	Leptophos	TEDP (sulfotepp)	Disulfoton	Methamidophos	TEPP	EPN	Methyl dematon	Terbufos	Ethion	Methyl parathion	
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Ethion	Methyl parathion																															

(1) Hill and Arnold [11]

(2) NMAM, 2nd ed. [7]

(3) NMAM, 3rd ed. [8]

(4) OSHA Analytical Methods Manual [12]

(5) OSHA Stopgap Methods [4]

Table 11. Preparation of Spiked Media and Liquid Calibration Standards

	Spiked Concentration					Solutions
	A.	B.	C.	D.	E.	
Spiking Levels (Fraction of REL) ⁽¹⁾	1/30x REL	1/10x REL	1/3x REL	1x REL	3x REL	
I. Spiking of Media or Liquid						
1. Spiking Solution to use:	SS-2	SS-2	SS-1	SS-1	SS-1	Dilute this volume (mL) of 10 mg/mL stock solution to prepare 10 mL of solution SS-1
2. Preferred Syringe Size:	50 µL	50 µL	50 µL	50 µL	100 µL	
3. Spiking Volume ⁽²⁾ to use:	10 µL	30 µL	10 µL	30 µL	90 µL	
II. Total µg spiked⁽³⁾						
1. Azinphos methyl	0.8	2.4	8	24	72	0.8
2. Chlorpyrifos	0.8	2.4	8	24	72	0.8
3. Diazinon	0.4	1.2	4	12	36	0.4
4. Dicrotophos	1.0	3	10	30	90	1.0
5. Disulfoton	0.4	1.2	4	12	36	0.4
6. Ethion	1.6	4.8	16	48	144	1.6
7. Ethoprop	0.4	1.2	4	12	36	0.4
8. Fenamiphos	0.4	1.2	4	12	36	0.4
9. Fonofos	0.4	1.2	4	12	36	0.4
10. Malathion ⁽⁵⁾	4	12	40	120	360	4.0
11. Methamidophos	0.8	2.4	8	24	72	0.8
12. Methyl parathion	0.8	2.4	8	24	72	0.8
13. Mevinphos	0.4	1.2	4	12	36	0.4
14. Monocrotophos	1.0	3	10	30	90	1.0
15. Parathion	0.2	0.6	2	6	18	0.2
16. Phorate	0.2	0.6	2	6	18	0.2
17. Ronnel ⁽⁵⁾	4	12	40	120	360	4.0
18. Sulprofos	4	12	40	120	360	4.0
19. Terbufos	0.4	1.2	4	12	36	0.4
20. General (for 120L) ^(4,5)	x/30	x/10	x/3	x	3x	4y

- (1) For a collection volume of 120 L. Range corresponds to values within column C, Table 8.
- (2) For liquid calibration standard preparations, add specified volume to 2 mL desorption solution in 2-mL volumetric flask. For laboratory control samples spiked at the REL, apply volume specified in column D to front section of sampler; in duplicate. For Desorption Efficiency determination, apply specified volume to front section of sampler; do each of five levels in triplicate.
- (3) Total µg per sample, for spiked media, or per 2 mL desorption solution for liquid calibration standards.
- (4) Where x, µg/sample = REL, µg/L x 120 L/sample; and y, mg/mL = REL, mg/m³ x 4 m³/mL.
- (5) For all REL > 1 mg/m³, use 1/10 x REL in the calculations (assumes that collection volume in these cases would be 12 L instead of 120 L).





