

Appendix N

Endocrine Disruption and Invertebrates

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6 Considerations by the EDSTAC Screening and Testing Work Group (STWG) have predominantly
7 dealt with vertebrate animals for several reasons. The first, and perhaps overriding one, is that the
8 charge given to the work group of focusing on estrogen, androgen, and thyroid hormone actions
9 is not especially relevant to important and well-studied hormones of invertebrates. The purported
10 endocrine disruption effects of public concern are almost exclusively human health or vertebrate
11 wildlife related. The expertise in the work group is, also, predominantly with the vertebrate
12 classes. However, invertebrates represent over 95% of all animals, are ubiquitous, and are
13 tremendously important ecologically and economically. Commercial fisheries of shrimp, crab, and
14 oyster and agriculturally important insect pollination are but a few key examples. Because
15 invertebrates are ubiquitous and are easily adapted for laboratory testing, they can serve as
16 sentinels and surrogates for investigating environmental stress. For these reasons, invertebrates
17 should not be ignored from consideration.

18
19 Endocrine disruption has been well studied and well exploited for certain invertebrates, especially
20 the insects. The endocrine systems of insects have been intentionally targeted for insecticidal
21 activity and several insecticides have been developed and used to suppress insect populations by
22 disrupting their normal endocrine functions. Juvenile hormone mimics (e.g., methoprene),
23 antijuvenile hormone analogs (e.g., precocene), chitin synthesis inhibitors (e.g., diflubenzuron),
24 ecdysone analogs (e.g., tebufeno-zide), and molting disruptants (e.g., fenoxycarb) are some
25 examples. These insect growth regulating compounds have also been observed to have adverse
26 effects in related arthropods such as crustaceans, including disrupting normal molting processes,
27 limb regeneration, and reproduction (Christiansen et al., 1977a, b; 1979; Cunningham, 1976;
28 Forward and Costlow, 1978; Landau and Rao, 1980; Nimmo et al., 1980; Touart and Rao, 1987).
29 Other substances like the organotin TBT have caused imposex and intersex conditions in
30 gastropods (Gibbs and Bryan, 1986; Reijnders and Brasseur, 1992) and sewage outfalls have
31 caused intersex conditions in harpacticoid copepods (Moore and Stevenson, 1994), conditions
32 indicative of endocrine disruption.

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34 Although the relevance of estrogen and androgen hormones to invertebrates is unclear,
35 invertebrates may be useful as surrogates for investigating phenomena relevant to these hormones
36 in vertebrates. Estrogens have been reported to play a meaningful role in development and
37 reproduction in echinoderms and molluscs (Takeda, 1979; Brueggemeier et al., 1988; Shirai and
38 Walker, 1988). Daphnids have been used to investigate the effects of xenoestrogens on steroid
39 metabolism (Baldwin et al., 1995; Baldwin et al., 1997) and sex reversal (Shurin and Dodson,
40 1997). Because of their generally shorter life cycles and relative ease of handling many species in
41 the laboratory, invertebrates could be useful for evaluating endocrine disrupting phenomena.
42 However, additional research is needed before this promise is realized.

1 There are, therefore, two aspects to considering endocrine disruption for invertebrates, one is
2 relevance to the health of invertebrate organisms themselves and the other is relevance of
3 invertebrates as surrogates for investigating vertebrate-related phenomena. Conventional risk
4 assessment of toxic chemicals such as outdoor use pesticides and high volume industrial chemicals
5 generally include a crustacean reproduction or life cycle test in the data set used in the assessment.
6 Although specific endocrine system endpoints are not considered, the apical nature of these tests
7 may be adequate to detect the adverse consequences of an endocrine disrupting chemical in
8 crustacean arthropods. Additional information is needed to determine what is most useful beyond
9 these conventional tests for the wider invertebrate taxa. As surrogates, more information on the
10 correlation of endocrine phenomena between invertebrates and vertebrates would be helpful. For
11 instance, to what degree does a substance which disrupts ecdysteroid metabolism in crustacea
12 disrupt sex steroid metabolism in vertebrates? Perhaps good correlations may be found, but more
13 comparative information is needed before recommendations of specific invertebrate tests useful for
14 evaluating potential endocrine disrupting activity relevant to vertebrates can be made.

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16 No invertebrate assays, therefore, have been evaluated for use in T1S for detecting estrogen,
17 androgen, or thyroid hormone disruption. Invertebrate tests have been proposed for T2T. It is
18 recommended that a workshop of invertebrate endocrinologists and toxicologists be convened to
19 address first, the suitability of invertebrate assays for estrogen and androgen (not thyroid) for use
20 in a screening battery, and second, future improvements to the broader consideration of endocrine
21 disruption in the environment and the utility of invertebrates as surrogate test organisms.

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