

Advances in the development of tissue chip technologies for toxicity testing

STATE OF THE SCIENCE ON DEVELOPMENT AND USE OF NAMS FOR CHEMICAL SAFETY TESTING

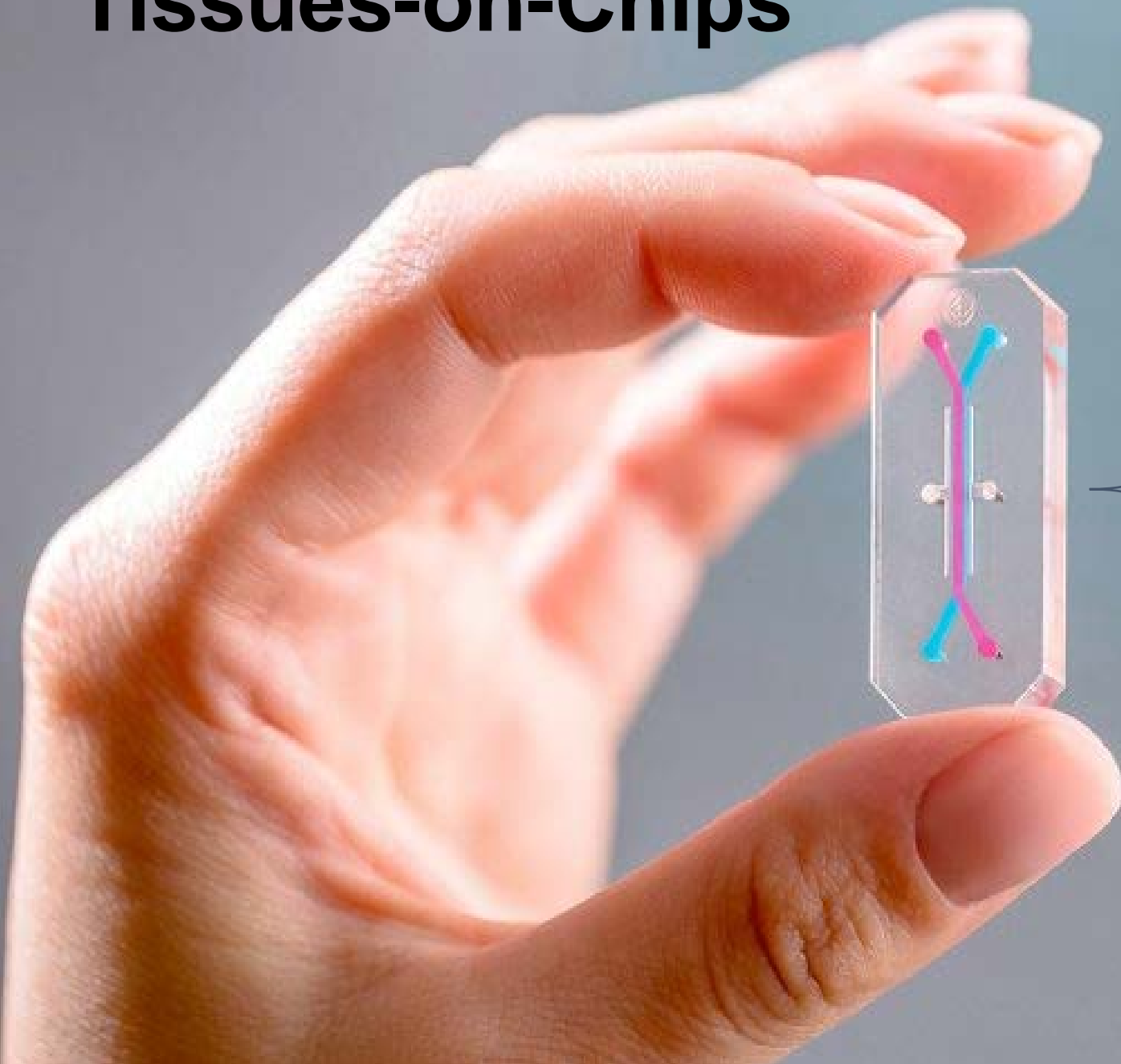
December 17, 2019

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Tissues-on-Chips



Scaffold

Cells

Structure

**Spatial and
Temporal
Patterning**

Perfusion

Bioreactor

Innervation

Host Response

**Functional
Readout**

**Computational
Design**

- Microfluidic cell culture devices
- Created with microchip manufacturing methods
- Contains continuously perfused chambers
- Seeded by human-derived cells
- Cytoarchitecture mimics tissue- and organ-level physiology
- High-resolution, real-time imaging and in vitro analysis of biochemical, genetic and metabolic activities



National Center
for Advancing
Translational Sciences

Tissue Chips 1.0 to Predict Drug Safety (2012-2017)

- James A. Thomson; Morgridge Institute for Research at the University of Wisconsin-Madison
Human induced pluripotent stem cell and embryonic stem cell-based models for predictive neural toxicity and teratogenicity

- John P. Wikswa; Vanderbilt University
Neurovascular unit on a chip: Chemical communication, drug and toxin responses

- Steven C. George; University of California, Irvine
An integrated in vitro model of perfused tumor and cardiac tissue

- D. Lansing Taylor; University of Pittsburgh
A 3-D biomimetic liver sinusoid construct for predicting physiology and toxicity

- James M. Wells; Cincinnati Children's Hospital Medical Center
Generating human intestinal organoids with an enteric nervous system

- John P. Lynch; University of Pennsylvania
Modeling oxidative stress and DNA damage using a gastrointestinal organotypic culture system

- George A. Truskey; Duke University
Circulatory system and integrated muscle tissue for drug and tissue toxicity

- Rocky S. Tuan; University of Pittsburgh
Three-dimensional osteochondral micro-tissue to model pathogenesis of osteoarthritis

- Linda Griffith; Massachusetts Institute of Technology
All-human microphysical model of metastasis and therapy

- Thomas Hartung; Johns Hopkins University
A 3-D model of human brain development for studying gene/environment interactions

- Kevin K. Parker; Harvard University
Human cardio-pulmonary system on a chip

- Joan E. Nichols; The University of Texas Medical Branch at Galveston
Three-dimensional human lung model to study lung disease and formation of fibrosis

- Mark Donowitz; Johns Hopkins University, Baltimore
Human intestinal organoids: Pre-clinical models of non-inflammatory diarrhea

- **Teresa Woodruff; Northwestern University**
Ex Vivo Female Reproductive Tract Integration in a 3-D Microphysiological

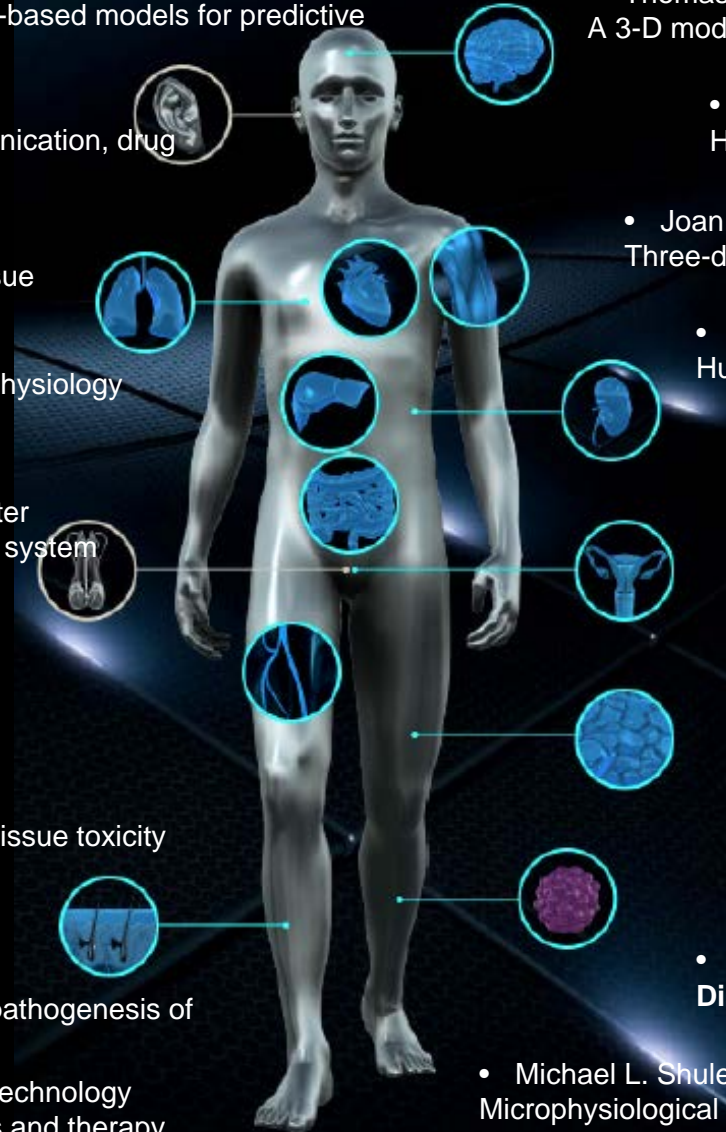
- Jonathan Himmelfarb; University of Washington, Seattle
A tissue-engineered human kidney microphysiological system

- Gordana Vunjak-Novakovic; Columbia University Health Sciences
Integrated Heart-Liver-Vascular Systems for Drug Testing in Human Health and Disease

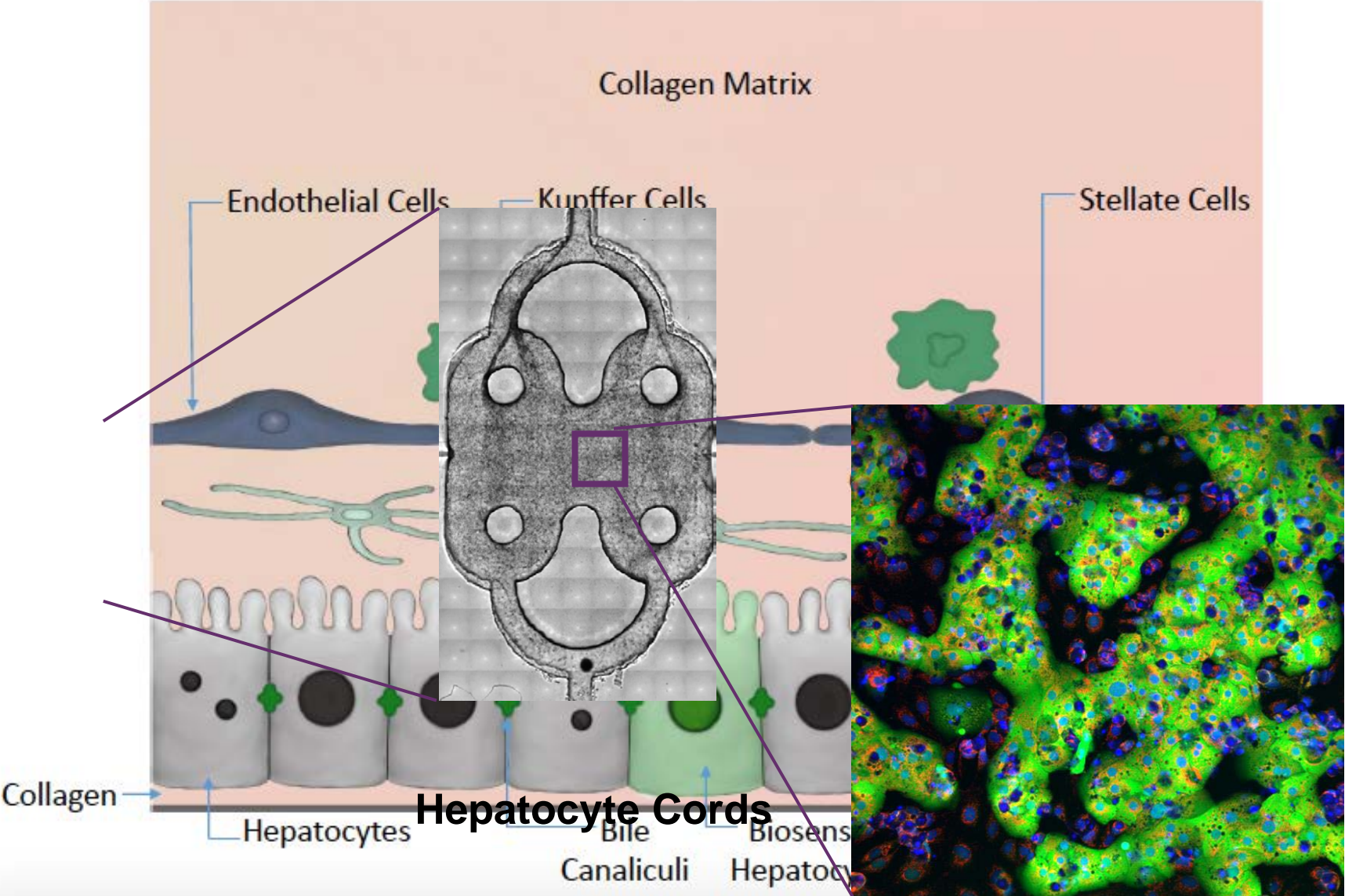
- Angela Christiano; Columbia University Health Sciences
Modeling complex disease using induced pluripotent stem cell-derived skin constructs

- **Kevin E. Healy; University of California, Berkeley**
Disease-specific integrated microphysiological human tissue models

- Michael L. Shuler; Cornell University
Microphysiological systems and low cost microfluidic platform with analytics



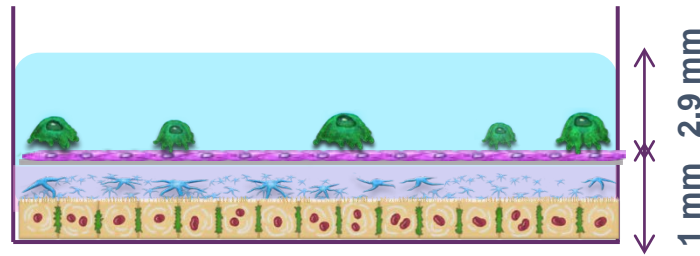
Liver on Chip: Self-assembly of Hepatocytes and NPC



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4 Week Co-culture of Human Hepatocytes and Non-Parenchymal Cells

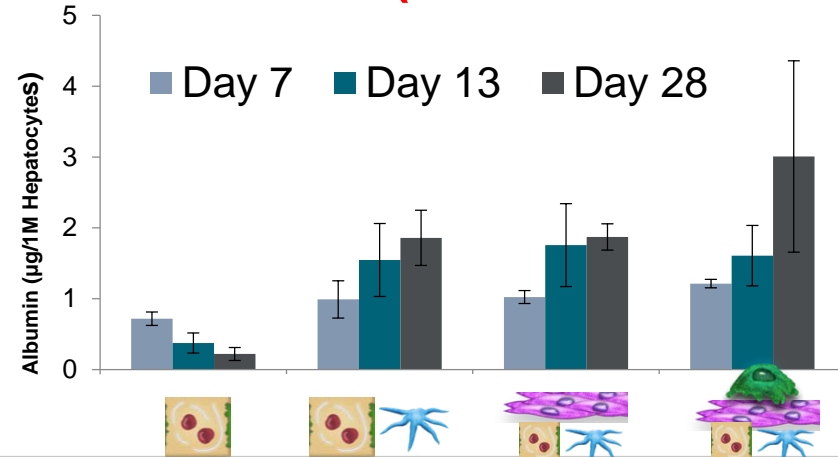
Transwell Culture Format



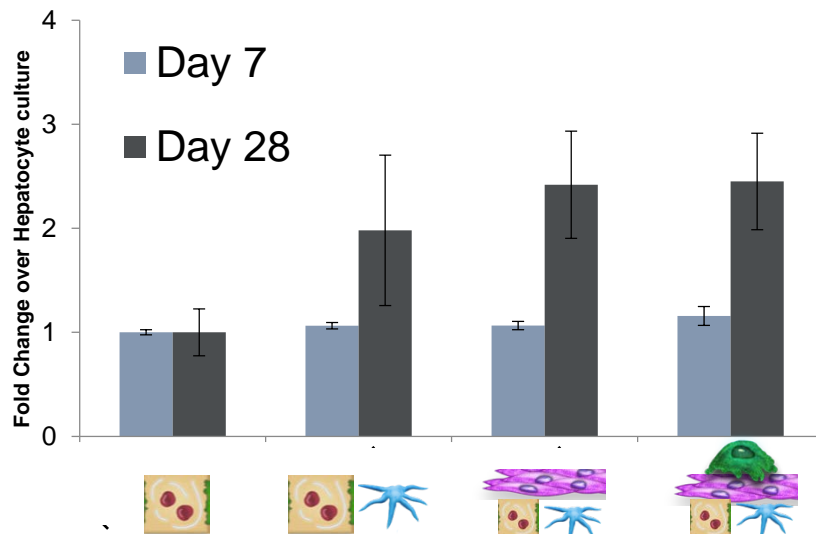
Volume/Cell (nL/cell) = 3

Distance btw cells (mm) = 1

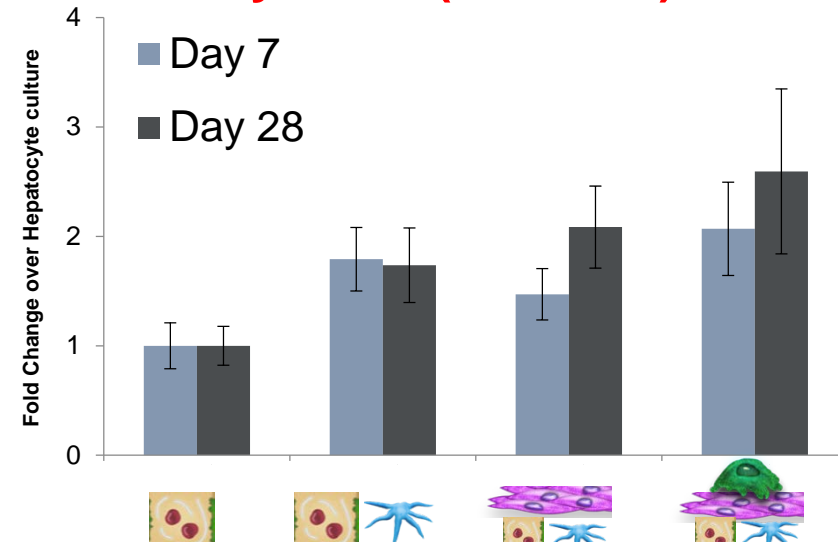
Stable Function (Albumin Secretion)



LDH in cell Lysates (Viable Cultures)

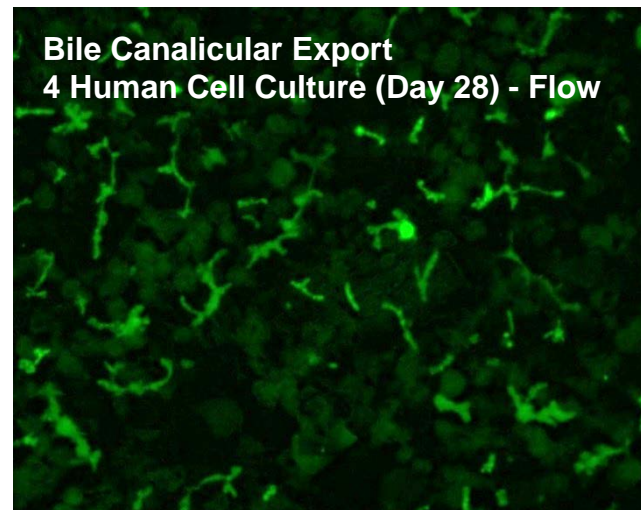
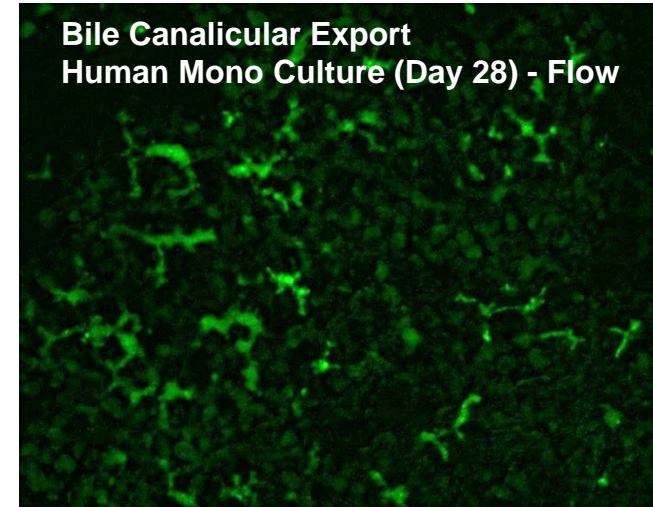
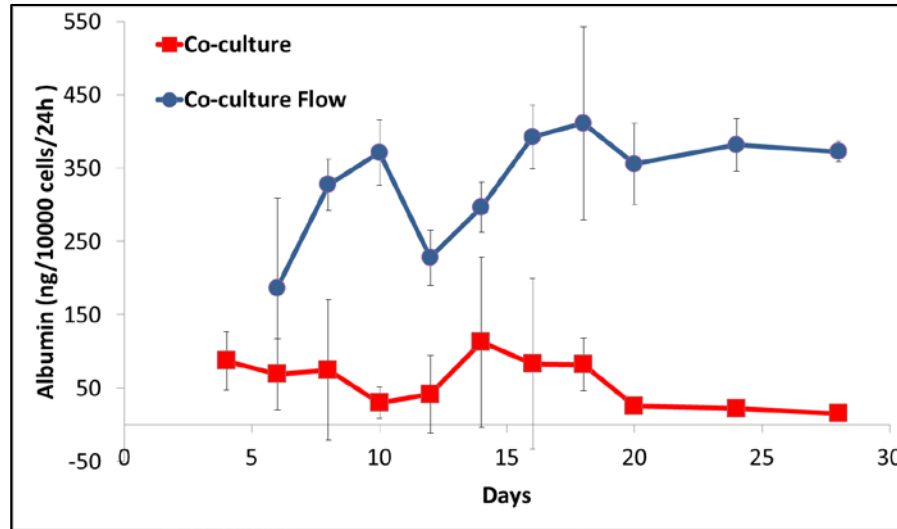


Stable Enzymatic (CYP3A4) Activity

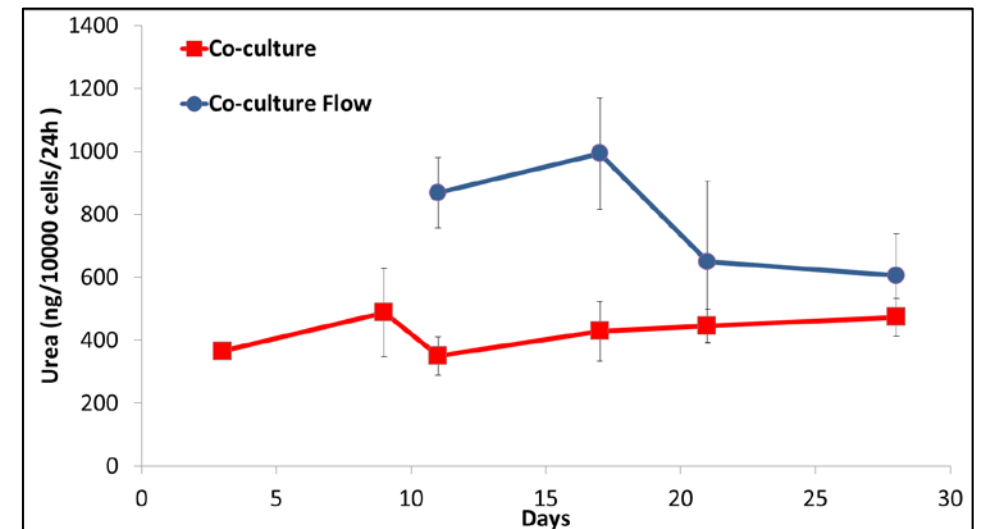


4 Week Culture: Static vs. Flow

Albumin Secretion in Flow vs Static Culture



Urea Secretion in Flow vs Static Culture

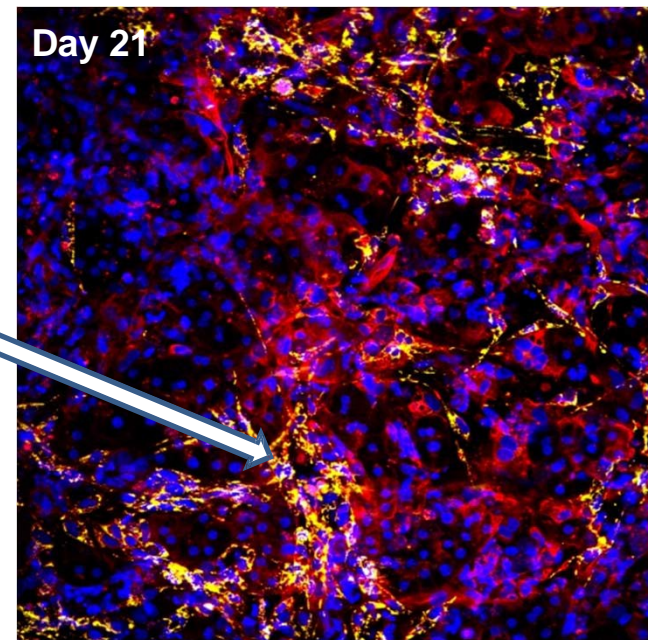


Vernetti et. al. 2016. A human liver microphysiology platform for investigating physiology, drug safety and disease models. *Expt Biol Med* 241: 101

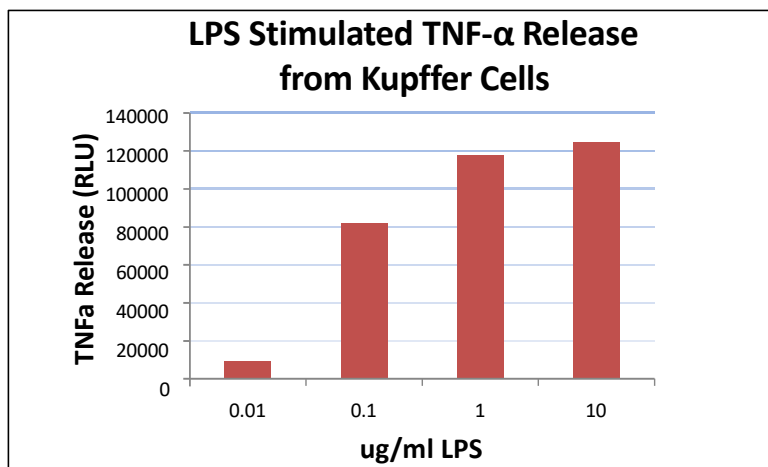
Methotrexate induced fibrosis and LPS Immune-mediated hepatotoxicity

Fibrotic response after stellate cell activation in response to methotrexate

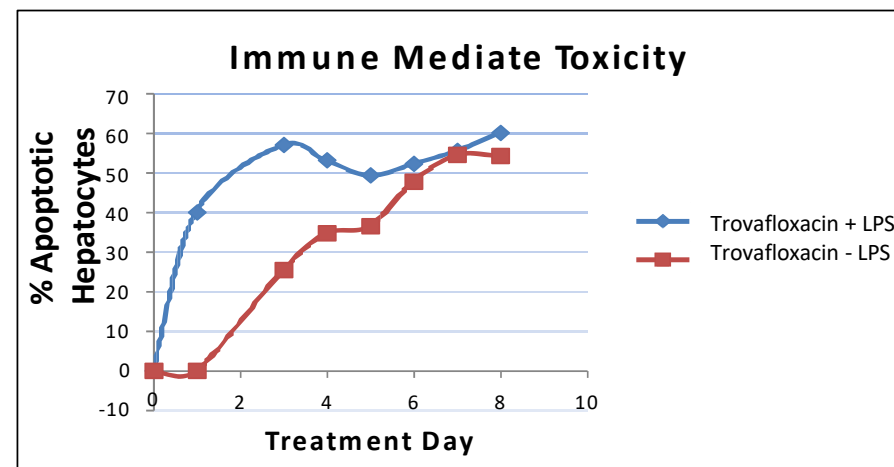
Stellate cells Express Collagen 1A2



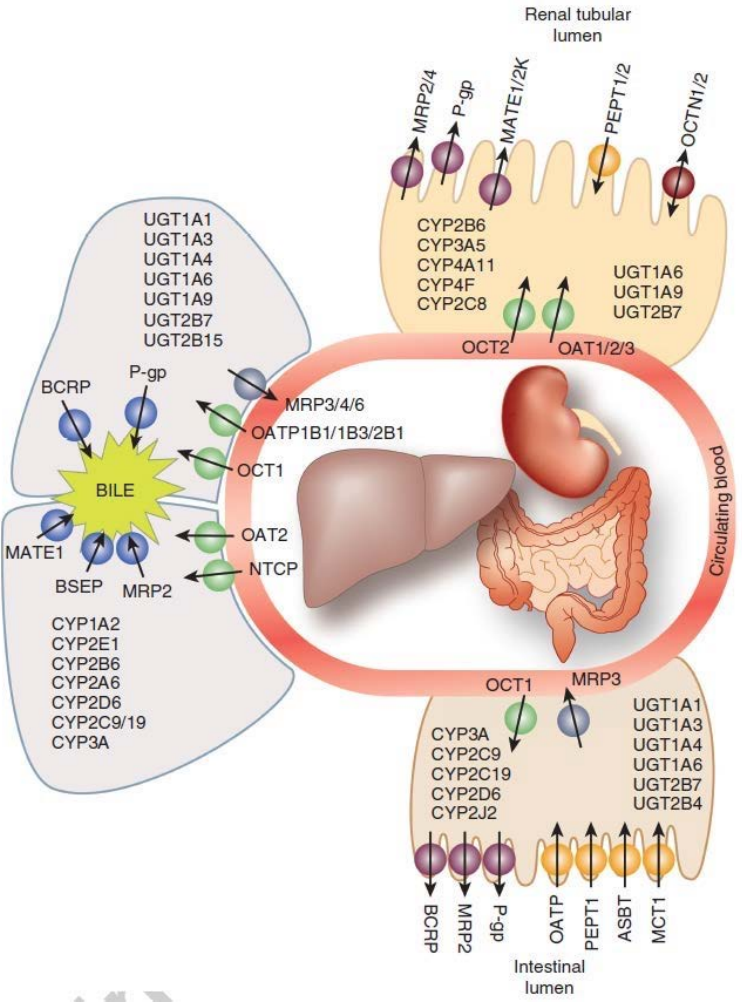
Immune-Mediated Hepatotoxicity by LPS



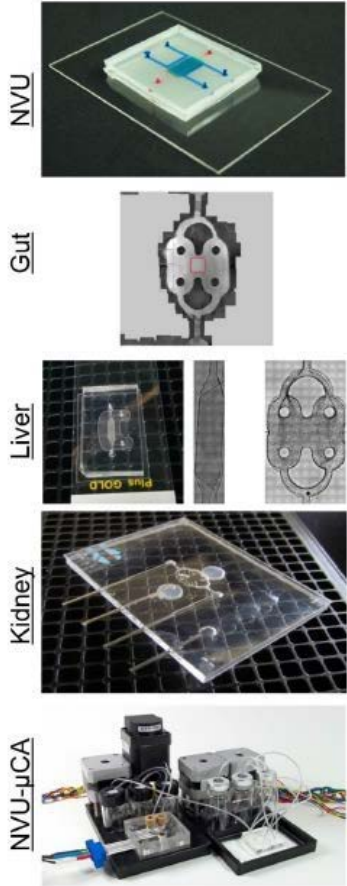
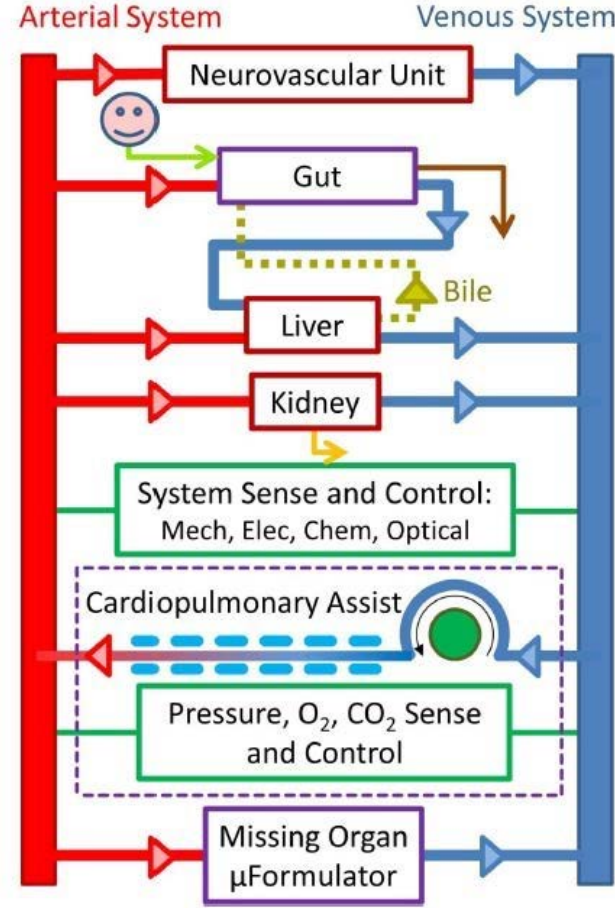
LPS + Trovafloxacin induces apoptosis



Integration of Multiple Tissue Chips



Yeung et al, Kidney International

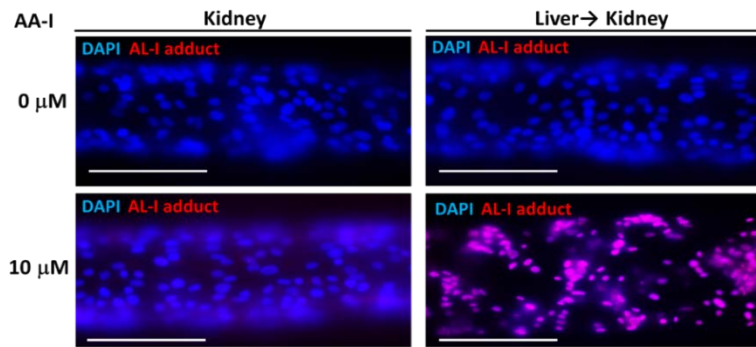


Verneti et al, Scientific Reports 2017

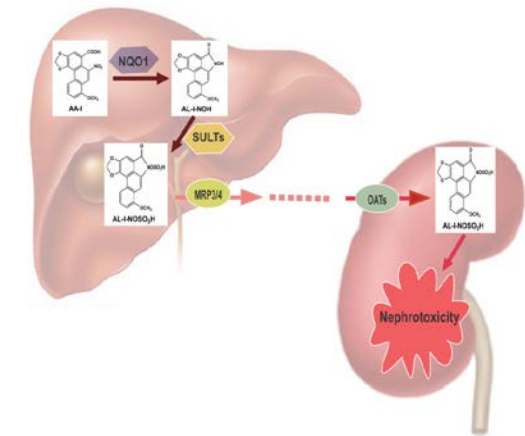
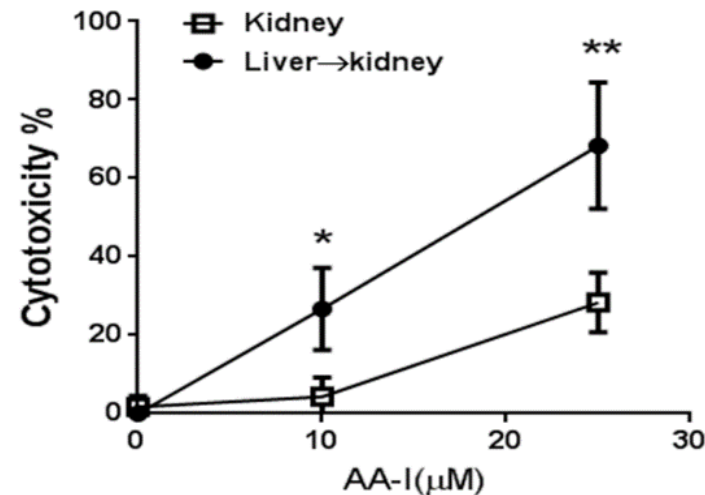
Human Mechanism of Aristolochic Acid (AA) Nephropathy Elucidated by Coupled Liver-Kidney MPS

Vernetti et al, Scientific Reports 2017

- AA is highly nephrotoxic and a Class I carcinogen for the lower urinary tract
- Identified as etiological agent for Chinese Herbs Nephropathy and Balkan Endemic Nephropathy
- Mode of action via adducting to DNA and proteins
- Pathways of human bioactivation not fully elucidated

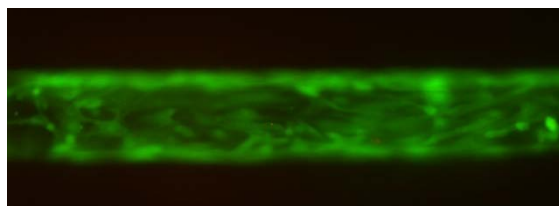


Hepatocyte AA bioactivation required

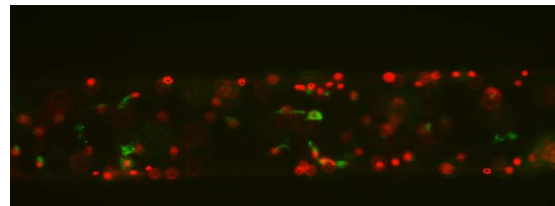


Multistep model

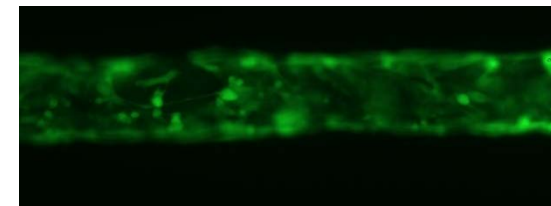
Vehicle control



AL-I-NOSO3 25 uM

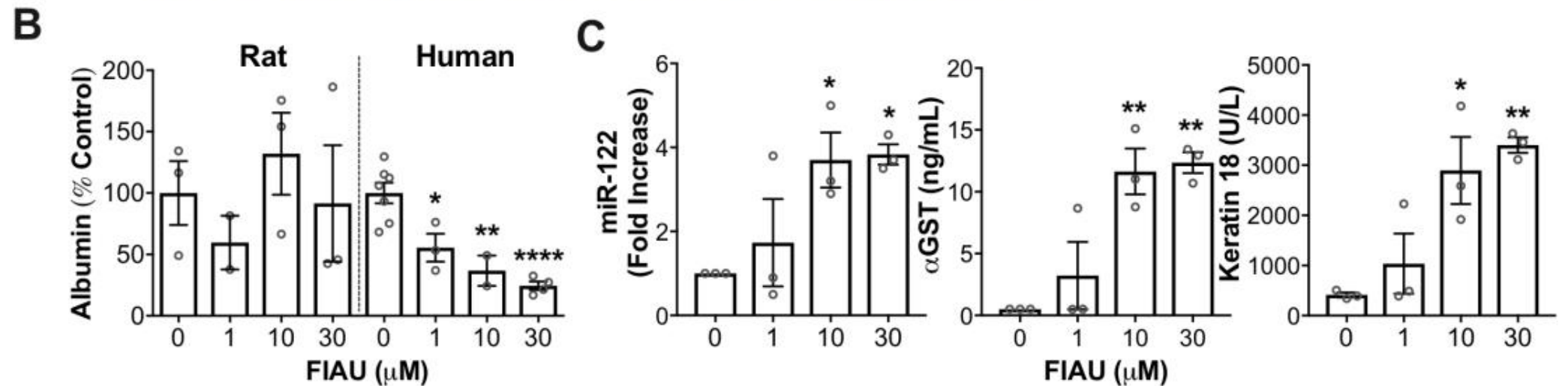
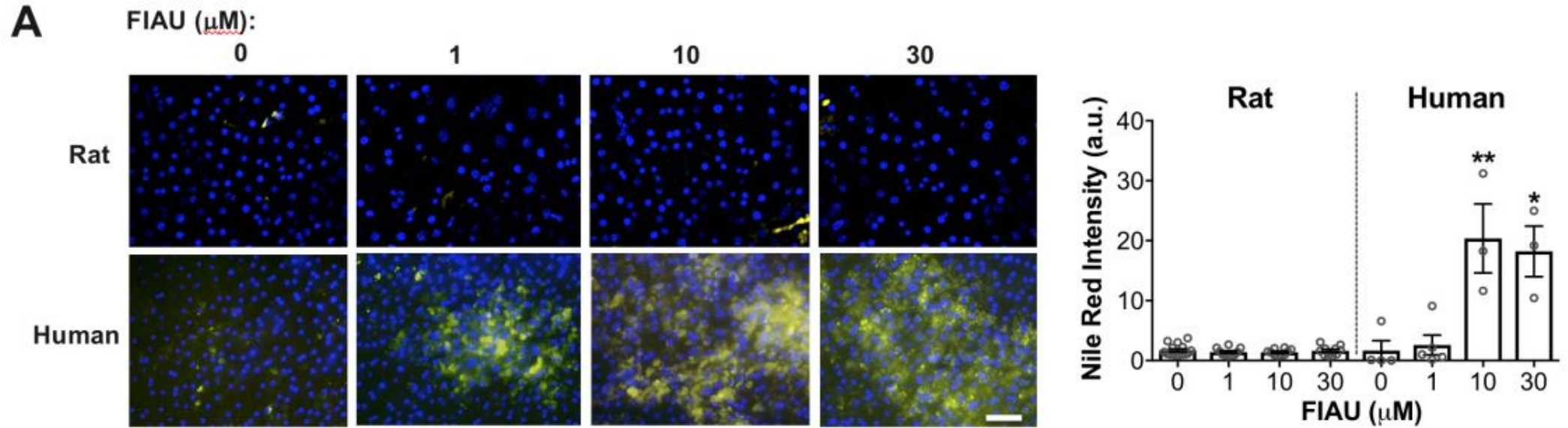


AL-I-NOSO3 25 uM + Pro 2mM

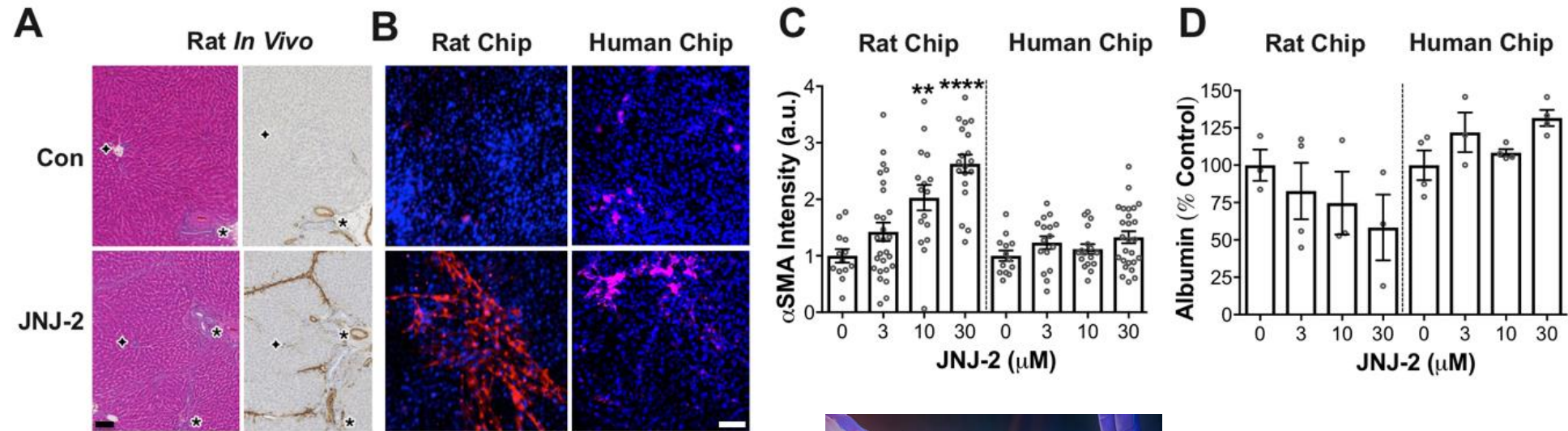


(AL-I-NOSO3 as causative agent: Live/Dead staining following direct kidney 24 h exposure)

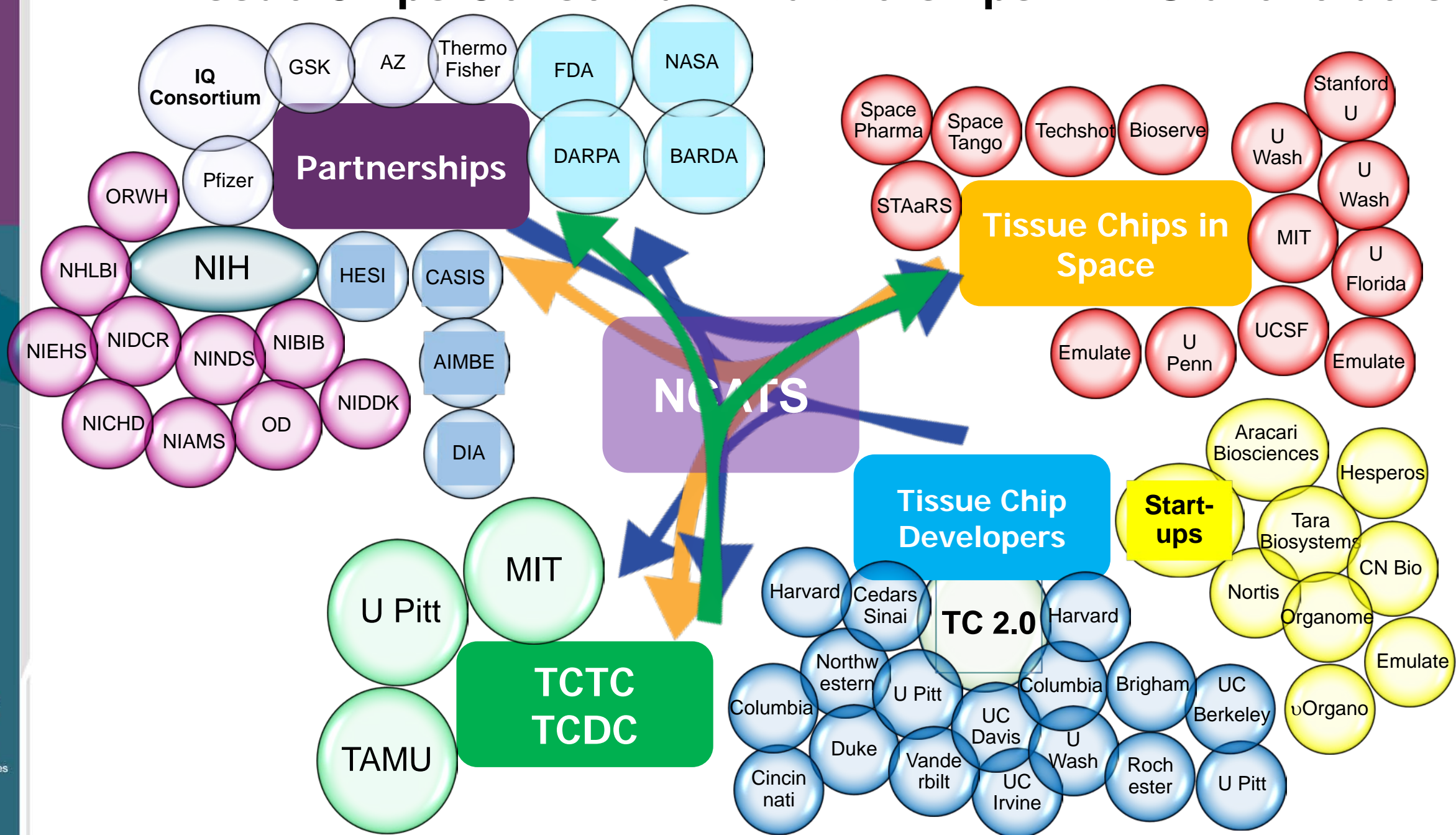
Species Differences in Steatosis using Rat and Human Liver-Chips following Fialuridine (FIAU) Treatment



Species Differences in Fibrosis using Rat and Human Liver-Chips



NIH Tissue Chips Consortium- Partnerships with Stakeholders



NIH Tissue Chips 2.0 for Disease Modeling and Efficacy Testing (2018 to 2022)

Kam Leong, Columbia U

Proteus Syndrome and DiGeorge Syndrome

Danielle Benoit, Lisa Delouise, Catherine Ovitt, U Rochester

Radiation-induced xerostomia

Kevin Kit Parker, William Pu, Harvard U

Barth syndrome, catecholaminergic polymorphic ventricular tachycardia, arrhythmogenic cardiomyopathy

Steven George, David Curiel, Stacey Rentschler, UC Davis and WashU

atrial fibrillation

Joseph Vincent Bonventre, Luke Lee, Brigham and Women's

autosomal dominant/recessive models of polycystic kidney disease, Focal segmental glomerulosclerosis

Christopher Hughes, UC Irvine

Hereditary hemorrhagic telangiectasia, Port Wine stain, Sturge-Weber syndrome

Rocky Tuan, U Pittsburgh

Osteoarthritis, inflammatory arthritis, adipose-mediated diabetic joint complications

Clive Svendsen, Cedars-Sinai

ALS; Parkinson's Disease

Aaron Bowman, Kevin Ess, John Wikswo, Vanderbilt U

tuberous sclerosis complex (TSC) epilepsy, DEPDC5-associated epilepsy, & associated cardiac dysfunction

Gordana Vunjak-Novakovic, Columbia U

Dox induced cardiomyopathy; multi-system pathologies involving heart, liver, skin, bone and vasculature

Donald Ingber, Harvard U

influenza infection, COPD

Jonathan Himmelfarb, U Washington

apolipoprotein L1 mediated kidney disease, drug induced and host-pathogen interaction induced renal thrombotic microangiopathies

Teresa Woodruff, Northwestern U

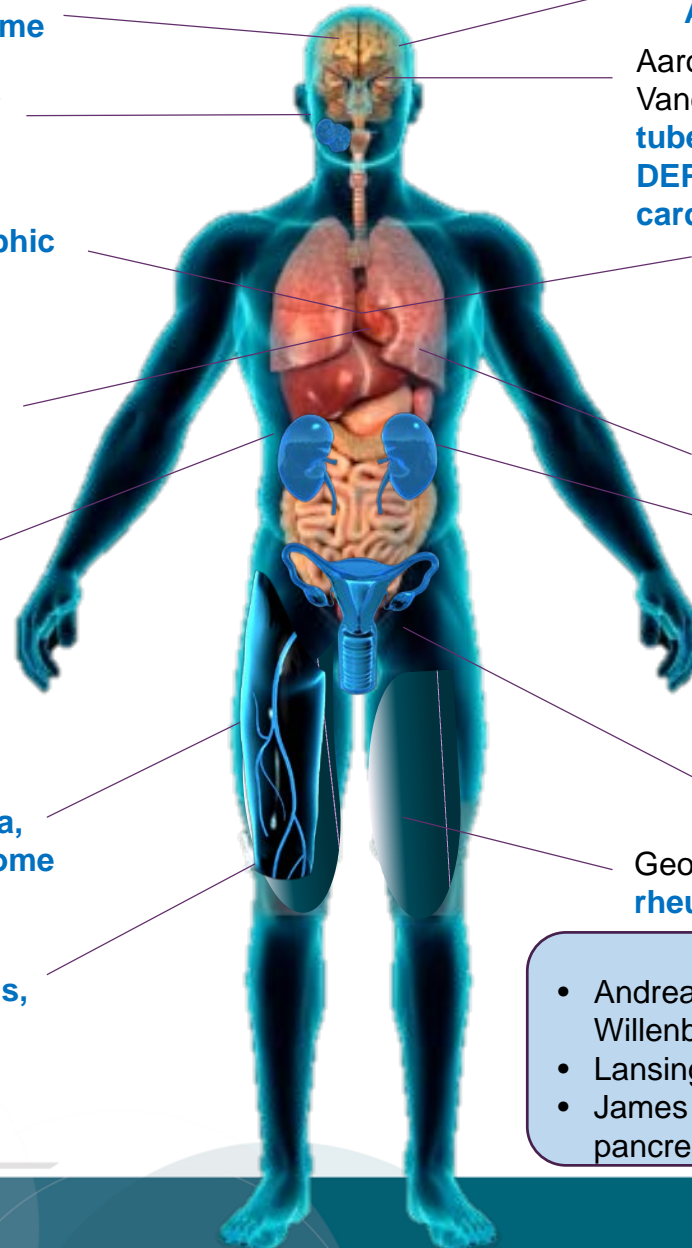
Polycystic Ovarian Syndrome

George Truskey, Duke U

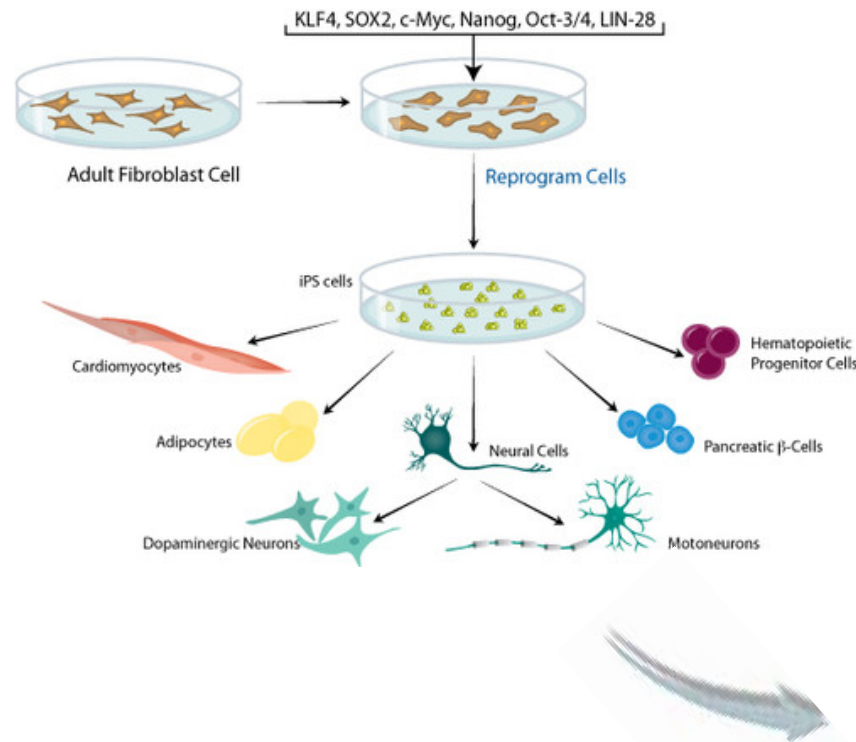
rheumatoid arthritis, atherosclerosis

Type-2 Diabetes Mellitus

- Andreas Stahl, Kevin Healy, Matthias Hebrok, Edward Hsiao, Holger Willenbring, UC Berkeley - Pancreatic islet, liver, adipose
- Lansing Taylor, U Pittsburgh – Vascularized liver and pancreatic islets
- James Wells, Moo-Yeal Lee, Cincinnati Children's Hospital - Liver, pancreatic islet and intestine



Current NIH Initiatives for Tissue Chips



Human body on Chip

- Co-culture of many differentiated iPSC-derived cell types per tissue architecture and composition
- Integration of different tissue chips to form human body on chip
- Genome editing to introduce various polymorphisms on isogenic iPSC lines
- Developmental/pediatric response to drugs/toxins
- Rare diseases

Just awarded:

- **Nociception-on-chip** **RFA-TR-19-003** (5 awards)
- **Immune system-on chip** **PAR-19-138** (3 awards)
- **ADRD on chip** **RFA-NS-19-027** (1 award)

To be reviewed:

- **“Clinical Trials-on-chips” for Precision Medicine (You-on-chip)** **RFA-TR-19-014**

New or ongoing Initiatives:

- **Nervous system MPS** **PAR-16-398** (Standard dates until January 8, 2020)
- **Cancer Biomimetics** **PAR-19-113** (Standard dates until January 8, 2022)
- **BBB interface on chip** **RFA-HL-20-21** (December 2, 2019; October 19, 2020)
- **Biomimetics for Infectious diseases** **RFA-AI-20-009** (to be released)



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Tissue Chips Consortium – Lead: Danilo A. Tagle

Program Managers: Lucie Low, Ph.D. and Passley Hargrove-Grimes

Trans-NIH Microphysiological Systems Working Group

- Nathan M. Appel (NIDA)
- Guillermo Arreaza-Rubin (NIDDK)
- David Balshaw (NIEHS)
- Steven Becker (NEI)
- Lisa Begg (OD)
- Bonnie Burgess-Beusse (NIDDK)
- Warren Casey (NIEHS)
- Preethi Chander (NIDCR)
- Ricardo Cibotti (NIAMS)
- Ki-Cha Flash (NCATS)
- Nancy Freeman (NIDCD)
- Daniel Gossett (NIDDK)
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- Anthony Kirilusha (NIAMS)
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- David Panchision (NIMH)
- Aaron Pawlyk (NIDDK)
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- Bradley Wise (NIA)
- Da-Yu Wu (NIDA)
- Nastaran Zahir (NCI)

• FDA

- Khaled Bouri, Ph.D., M.P.H., **OC**
- Paul Brown, Ph.D., **CDER**
- Tracy Chen, Ph.D., DABT, **OC**
- Karen Davis-Bruno, Ph.D., **CDER**
- Suzanne Fitzpatrick, Ph.D., **CFSAN**
- Timothy McGovern, Ph.D., **CDER**
- Donna Mendrick, Ph.D., **NCTR**
- Thomas Papoian, Ph.D., DABT, **CDER**
- Alexandre Ribeiro, Ph.D., **CDER**
- James Weaver, Ph.D., **CDER**

• ISS-NL (CASIS)

- Michael Roberts, Ph.D.
- Marc Giulianotti, Ph.D.
- Bill McLamb, Ph.D.
- Melissa Rhodes, Ph.D.

Liz Warren, Ph.D.
Patrick O'Neill

• IQ MPS Affiliate

- IQ MPS Executive Committee (EC): IQ MPS Chair (**Will Proctor, Genentech**), Vice Chair (**Monica Otieno, Janssen**) and Vice Chair-Elect (**Terry van Vleet, AbbVie**); IQ-NCATS engagement workstream POC (**Jason Ekert GSK**)
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- Ananthsrinivas Chakilam, Ph.D., **Vertex**
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- David Daignan, Ph.D., **AbbVie**
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- Rahda Sura, Ph.D., **AbbVie**
- Matthew Wagoner, Ph.D., **Takeda**
- David Watson, Ph.D., **Eli Lilly**
- Yvonne Will, Ph.D., **Pfizer**



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Thank you!

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