#### 2. INTRODUCTION

An independent laboratory validation for study RES-00061 is required to meet residue regulatory requirements.

#### 3. OBJECTIVE

The study objective was to independently validate the method reported RES-00061 for the fenpyroximate in s oil according t o t he E C guidance doc ument determination of SANCO/825/00 rev. 8.1, ENV/JM/MONO(2007)17 and EPA OCSPP 850.6100 (2012).

#### 4. **TEST ITEMS**

The certificate of analyses for fenpyroximate is presented in Appendix 1.

#### 4.1. Fenpyroximate

Product Name: Fenpyroximate Standard Common Name: Fenpyroximate 5AA0023P Lot Number: Chemical Name: *tert*-Butyl (*E*)- $\alpha$ -(1,3-dimethyl-5-phenoxypyrazol-4ylmethyleneamino-oxy)-p-toluate Structural Formula: 0 CH<sub>3</sub> CH3  $CH_3$ CH<sub>2</sub> Molecular Formula: C24H27N3O4 Molecular Weight: 421.50 CAS-Registry-No.: 134098-61-6 Purity: 99.4%

Storage Conditions: Expiry Date:

Refrigerated in the dark 06 October 2021

#### 5. TEST SYSTEMS

Sandy loam and clayey loam soil samples were taken from Battelle validation control stock samples. See Appendix 3 and Appendix 4 for the characterisation reports. The soil moisture for each soil type was determined prior to analysis.

### 6. METHOD VALIDATION

The de termination of fenpyroximate residues in soil w as performed using t he m ethod described in RES-00061 (Ref 1).

The control samples were fortified as described in the following table:

Matrix	Analyte	Reagent Blank Replicates	Untreated Control Replicates	Replicates at LOQ Fortification Level	Replicates at LOQ × 10 Fortification Level
Sandy Loam	Fenpyroximate	1	2	5 at 0.01 mg/kg	5 at 0.10 mg/kg
Clayey Loam	ey Loam Fenpyroximate		2	5 at 0.01 mg/kg	5 at 0.10 mg/kg

LOQ = Limit of Quantification

The m ethod w as va lidated i n t erms of 1 inearity, s electivity, a ccuracy a nd pr ecision, monitoring two ion mass transitions.

Matrix effects were investigated at the LOQ and LOQ×10 levels by comparing peak areas of solvent standard solutions to peak areas of matrix-matched standard solutions. Experiments assessed w hether o r n ot m atrix effects w ere s ignificant (i.e. >20% en hancement o r suppression).

# 7. EXPERIMENTAL

### 7.1. Principle of the Method

Residues of fenpyroximate were extracted by subsequent extractions with methanol, acetone, methanol/water and m ethanol/hydrochloric a cid. F inal de termination w as b y LC-MS/MS monitoring two ion mass transitions. The limit of quantification (LOQ) was 0.01 mg/kg and the limit o f d etection (LOD) w as 0.00 02 mg/kg (clayey loam) and 0.0004 mg/kg ( sandy loam).

The analytical flow chart is presented in Figure 1.

### 7.2. Equipment, Consumables and Reagents

Full details of all equipment, consumables and reagents are presented within the analytical methods presented in Appendix 2.

### 7.3. Standards and Fortifications

### 7.3.1. Stock Solutions

Duplicate s tock s olutions were p repared b y dissolving a know n w eight (mg) of fenpyroximate, c orrecting f or pur ity and di ssolving i n acetonitrile to pr oduce a f inal concentration of 1000  $\mu$ g/mL. Full actual stock solution preparation and final concentrations are detailed in the following table:

Analyte	Battelle (BUKL) Stock ID	Purity (%)	Actual Amount Weighed (mg)	Actual Volume Added (mL)	Concentration (µg/mL)
Fenpyroximate	BAT-6752	99.4	10.28	10.218	1000
	BAT-6753	99.4	10.48	10.417	1000

One was used for calibration standard preparation and the other was used for recovery fortification preparation.

### 7.3.2. Fortification Solutions

Two fortification solutions, containing fenpyroximate at concentrations of 1.0 and 10  $\mu$ g/mL, were prepared by diluting appropriate amounts of the stock solution with acetonitrile.

Recovery efficiency samples were fortified according to the following table:

Matrix	Sample Weight* (g)	Fortification Standard Concentration (µg/mL)	Fortification Volume (µL)	Fortification Level (mg/kg)
Sandy and Clayey	25	1.0	250	0.01
Loam Soil	25	10	250	0.10

\* Dry weight of soil

## 7.3.3. Calibration Solutions

An intermediate solution c ontaining fenpyroximate at a concentration of 1000 n g/mL was prepared b y di luting an appropriate amount of the s tock s olution w ith a cetonitrile: w ater (1:1 v : v). Further i ntermediate s olutions containing Fenpyroximate w ere p repared at concentrations of 250, 200, 100 , 50, 20, 10, 3.0 a nd 2.5 ng /mL by d iluting a ppropriate amounts of the 1000 ng /mL i ntermediate s olution w ith a cetonitrile: w ater (1:1, v: v). The intermediate solutions were then used to prepare matrix matched calibration solutions.

All standard solutions were stored in the refrigerator when not in use.

### 7.4. Soil Moisture Determination

Approximately 60 g (3 x 20 g aliquots) of each soil were weighed and left to dry in an oven at 105 °C. The samples were initially weighed after approximately 3 hours of drying. The samples were returned to the oven and re-weighed again after 17 hours. After comparison of the two weights, the difference was deemed negligible and the soil samples were determined to be dry. The final dry weight for each aliquot was used to calculate the moisture of the initial sample

The moisture determination in each soil sample was calculated according to the following equation:

Moisture Content [%] =  $\frac{\text{Initial wet weight (g)} - \text{Final dry weight (g)}}{\text{Initial wet weight (g)}} \times 100$ 

The mean moisture content for each soil type was calculated by averaging the three individual moistures for each soil type.

## 7.5. Extraction Procedure

Aliquots of 25 g (dry weight) of soil were placed into 125 mL extraction bottles. For recovery efficiency tests, the control matrices were fortified with the appropriate spiking solutions at the LOQ and 10xLOQ.

For all samples, 50 mL of methanol was added to each bottle and mechanically shaken for 20 minutes. A fter c entrifugation (1500 r pm f or 2 minutes), t he s ample w as f iltered t hrough cotton wool. The extraction process was repeated 5 more times with the following solvents;

- 1. 50 mL methanol
- 2. 50 mL methanol
- 3. 50 mL acetone
- 4. 50 mL methanol: water (1:1 v:v)
- 5. 50 mL methanol: 0.1 M hydrochloric acid (aq, 1:1 v:v)

After combining all extracts, the final volume was a djusted to 500 mL with water and an aliquot t ransferred t o a glass H PLC v ial. R esidue l evels w ere t hen d etermined b y LC-MS/MS.

The extraction method is presented in the form of a flow chart in Figure 1.

# 7.6. LC-MS/MS Analysis

All s amples w ere a nalysed b y l iquid c hromatography coupled with a t andem m ass spectrometer (LC-MS/MS), monitoring two ion mass transitions.

A summary of these conditions are presented below:

Mass Spectrometer and General Instrument Conditions						
Instruments	<u>API 5500</u> Triple Quadrupole Mass Spectrometer fitted with Turbo ion spray ion source					
Ion Source	Positive Electros	Positive Electrospray (ESI+)				
Run Time	3.5 minutes (Ap)	proximate retention ti	me of 2.5 minutes	5)		
Analyte	Dwell Time (msec)	Transition (m/z)	Collision Energy	Cell Exit Potential		
Econovimete	100	422/366	10	10		
renpyroximate	100	422/135	45	10		
Curtain Gas	40					
CAD Gas	-2 (Medium)	(Medium)				
Gas 1	45					
Gas 2	40					
Spray Voltage	5500 V					
Source Temperature	550°C					
Declustering Potential	g Potential 21					
Entrance Potential	ential 10					

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Mass Spectrometer and General Instrument Conditions					
Instruments	<u>API 6500</u> Triple Quadrupole Mass Spectrometer fitted with Turbo ion spray ion source				
Ion Source	Positive Electrospray (ESI+)				
Run Time	3.5 minutes (App	proximate retention t	ime of 2.5 minutes	5)	
Analyte	Dwell Time (msec)Transition (m/z)Collision Energy		Cell Exit Potential		
Fennyrovimate	100	422/366	17	16	
Tenpyroximate	100	422/135	43	8	
Curtain Gas	40				
CAD Gas	-2 (Medium)				
Gas 1	45				
Gas 2	40				
Spray Voltage 5500 V					
Source Temperature	Source Temperature 550°C				
Declustering Potential	1				
Entrance Potential	10				

HPLC Conditions				
Columns	Sigma, Ascentis Express C18, 50 x 2.1 mm, 2.7 µm			
Column Oven Temperature	45 °C			
Mobile Phase A	Water + 0.1% Formic acid			
Mobile Phase B	Acetonitrile + 0.1% Formic acid			
Method	Time	%A	%B	
	0	40	60	
	3.5	40	60	
Flow Rate	0.5 mL/min			
Injection Volume	10 µL			

# 8. CALIBRATION AND CALCULATIONS

A multi-point calibration curve was obtained from injections of calibration solutions by plotting peak areas of fenpyroximate versus the concentration in ng/mL. The curves were calculated by the method of least squares linear regression. 1/x weighting factor was applied to the curves to improve the accuracy.

The quantification of fenpyroximate in the samples was made by comparison to the calibration curve of the form y = mx + c. The amount of analyte in a given sample was calculated as follows:

Compound  $[mg/kg] = (A-c) \times F$ m x W x 1000

Where:

А	=	Area of analyte peak
m	=	slope of the calibration curve
c	=	intercept of the calibration curve
F	=	final volume (mL)
W	=	initial sample weight (g)

The recovery efficiency in the fortified samples was calculated as follows:

Recovery efficiency  $[\%] = \frac{\text{Amount found (mg/kg)}}{\text{Amount spiked (mg/kg)}} \times 100$ 

Example LC-MS/MS chromatograms of calibration solutions, control samples and fortified samples are presented in Figure 5 to Figure 18.

Examples of calibration curves are presented in Figure 3 to Figure 4 and response factors are presented in Figure 19 to Figure 20.

The response factor was calculated as follows:

Response Factor = <u>Response area (counts)</u> \* Prior to any concentration corrections.

# 15. FIGURES

# Figure 1: Analytical Flow Chart

EXTRA	CTION	
• • • •	Weigh 25 g o Fortify if nece Add 50 mL of Centrifuge sa Filter sample Repeat the e 1. 50 r 2. 50 r 3. 50 r 4. 50 r	f dry soil into a 125 mL extraction bottle ssary Methanol and shake on the reciprocating shaker for 20 minutes mple at 1500 rpm for 2 minutes through cotton wool into appropriate receptacle «traction (shake and filter, combining all extracts) with the following solvents: nL Methanol nL Methanol nL Acetone nL Methanol: Water 1:1 v:v
•	Adjust the fin	al volume of the sample to 500 mL using water
ANAL	YSIS	
•	Transfer an a Analyse by L Matrix standa	liquot of the sample into a clean clear vial using screw caps C-MS/MS rds used

The LOQ of the validated method was: 0.01 mg/kg