FINAL REPORT

TITLE

Independent Laboratory Validation (ILV) of the BAYER Method TD-002-W13-01: The Determination of Thiodicarb and its Metabolite Methomyl in Water using LC/MS/MS

TEST GUIDELINES

US EPA Test Guidelines: OCSPP 850.6100, OPPTS 850.7100 and OCSPP 860.1340(c) (6)





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2.0 **INTRODUCTION**

This report describes the independent laboratory validation (ILV) of Bayer Analytical Method TD-002-W13-01, "An Analytical Method for the Determination of Residues of Thiodicarb and its Metabolite Methomyl in Water Using LC/MS/MS" performed by ADPEN Laboratories, Inc. The analytical method is presented in Appendix 1.

This study was designed to satisfy harmonized guideline requirements described in US EPA Test Guidelines OCSPP 850.6100: Environmental Chemistry Methods and Associated Independent Laboratory Validation (Reference 2), OPPTS 850.7100 (Reference 3), and OPPTS (OCSPP) 860.1340(c) (6) (Reference 4). This study was conducted in compliance with EPA FIFRA Good Laboratory Practice Standards, 40 CFR Part 160 (Reference 5).

3.0 **MATERIALS AND METHODS**

3.1 **Reference Substances**

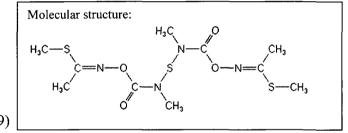
The following reference substances were obtained from Bayer CropScience:

Standard name: Thiodicarb

dimethyl-*N*,*N*-[thiobis](methylimino)carbonyloxy]]bis[ethanimidothioate]

59669-26-0 **Empirical formula:** $C_{10}H_{18}N_4O_4S_3$ Molecular Weight: 354.47 g/mol K-1470 Ref. Substance Lot: 0428200301 99.8% 10-Feb-2019 Storage conditions: Refrigerator (E-109)

Methomyl



CH₂

Standard name:

Standard Number:

IUPAC name:

CAS number:

GLP purity:

Expiration date:

IUPAC name: methyl-N-[[(methylamino)carbonyl]oxy]ethanimidothioate 16752-77-5 (E: 19929-35-9; Z: 19928-37-1) CAS number: Empirical formula: $C_5H_{10}N_2O_2S$ Molecular structure: Molecular Weight: 162.21 g/mol Standard Number: K-2059 Ref. Substance Lot: 1218200306 GLP purity: 99.5% H₃C Expiration date: 11-Feb-2018 CH₃ Storage conditions: Refrigerator (E-109)



3.2 Test System

The test system for this study was an in-house bulk control surface water sample, which was

The sample was logged into the study through the Laboratory Information Management System (LIMS) and assigned a unique laboratory code, which is cross-referenced to the sample number on raw data and detailed residue reports. During the course of this study, the sample was stored in freezer E-16, which operates at a temperature of less than -20 °C.

Sample extracts from the analytical set for thiodicarb (WO-13052401) were injected immediately after extraction. Sample extracts from the analytical set for methomyl (WO-13052404) were stored for one day in freezer E-51 while awaiting LC/MS/MS analysis. The temperature for freezer E-51 at this time was approximately -20 °C.

3.3 Analytical Procedures

Analytical Method TD-002-W13-01 was independently validated as written. The apparatus and reagents were used as outlined in the analytical method, see Appendix 1, with equivalent apparatus or reagents substituted as necessary.

3.3.1 Fortifications

Untreated control water samples were fortified using the appropriate fortification standard at LOQ (0.1 ppb) and $10 \times \text{LOQ}$ (1.0 ppb) concentrations as per the method. Fortifications used in this method validation are below.

Matrix	Fortification Volume (mL)	Fortification Concentration (ng/mL)	Sample Wt. (mL)	Final Concentration (ppb)	Replicates
Water	0.1	0.01	10.0	0.1	5
	0.1	0.10	10.0	1.0	5

3.3.2 Extraction Procedure

- 1. A 10-mL aliquot of water was transferred in to a 50-mL disposable polypropylene centrifuge tube.
- 2. SPE manifold with PTFE needles was prepared and Waters Oasis HLB 1cc Vac RC 60 mg cartridge was attached.
- 3. The cartridges were conditioned with 2-mL of methanol (MeOH) followed by of HPLC-grade Water (2 mL). The sample was eluted by gravity.
- 4. Recovery samples were fortified using individual standard solutions.

- 5. Samples were capped and briefly shaken to mix and centrifuged at 4000 RPM for 10 minutes.
- 6. A 5-mL aliquot from Step 5 was loaded on HLB cartridge and was eluted by gravity.
- 7. Approximately 20" Hg of vacuum was applied for 15 minutes and eluate was collected.
- 8. A 15-mL centrifuge tube was placed under each cartridge.
- 9. Sample was eluted by gravity with MeOH (2 mL) and the column was dried by applying approximately 5" Hg vacuum for 1 minute followed by MeOH/DCM solution (3:1, 1 mL v/v) eluting by gravity.
- 10. Step 9 was repeated.
- 11. Samples were evaporated to dryness in N-Evap at 45 °C, approximately 20 psi.
- 12. Samples were reconstituted with 2.5 mL L-Cysteine HCl in MeOH and vortexed for 20 seconds.
- 13. Samples were transferred to auto-sample vials for LC/MS/MS analysis.

3.3.3 Modifications

Other than instrument parameter optimizations, the following modification was made to the analytical procedure.

1. After fortification, recovery samples were centrifuged at the speed of 4,000 rpm instead of 12,500 g as stated in the method.

3.3.4 Suggestions

- 1. Due to criticality of the evaporation step in the procedure, one must consider stopping the evaporation in nitrogen evaporator a little earlier and use the technique of rolling the concentration tube and allowing the final volume of solvent to evaporate on its own.
- 2. In section 4.1 of Bayer Method TD-002-W13-01, specify the time for which the primary stock solutions of thiodicarb and methomyl are stable.
- 3. In section 4.3 of Bayer Method TD-002-W13-01, clarification is required that two untreated water samples will be fortified with each analyte separately.

HPLC Conditions					
HPLC Instrument:	Agilent 1200 SL				
HPLC Column:	Ascentis® Expr	scentis® Express C18, 2.1 mm × 100 mm, 2.7 µm			
Column Temp:	45 °C				
Injection Volume:	5 μL				
Mobile Phase A:	90/10 Water/Methanol with 10mM ammonium formate and 120µL/L formic acid				
Mobile Phase B:	10/90 Water/Methanol with 10mM ammonium formate and 120µL/L formic acid				
Gradient:	Time	Flow	В		
	(min)	(mL/min)	(%)		
	0	0.5	5		
	0.5	0.5	5		
	0.51	0.5	60		
	1.8	0.5	100		
	2.5	0.5	100		
	2.51	0.5	5		
	5	0.5	5		

3.4 Instrument Operating Parameters

MS/MS Conditions					
Mass Spectrometer	Agilent 6490 Series QQQ				
Ion Source	Electro Spray Ionization (ESI)				
Polarity	Positive				
Scan Type	MRM				
Gas Flow	14 L/min				
Temperature (TEM)	150 °C				
Nebulizer	45 psi				
Sheath Gas Heater	300 °C				
Sheath Gas Flow	12 L/min				
Capillary (V)	3000				
V Charging	1500				
MRM Conditions	Thiodicarb (Quantitation)	Thiodicarb (Confirmatory)	Methomyl (Quantitation)	Methomyl (Confirmatory)	
Q1 m/z	355.06	355.06	163.06	163.06	
Q3 m/z	88	108	88	105.9	
Dwell Time (msec)	100	100	500	300	
Frag (V)	380	380	380	380	
Collision Energy (CE)	6	10	2	6	

3.5 Data Acquisition

Peak integration and peak area count quantitation were performed by MassHunter Quantitative Analysis (version B.04.01) data handling software. A best-fit, linear regression equation was derived and used in conjunction with the analyte response in each sample to calculate the concentration of the analyte. The square of correlation coefficients (\mathbb{R}^2) for the calibration curves for each analytical set was greater than 0.99.

Statistical treatment of the data including the calculation of percent recovery, means, and standard deviations were calculated within LIMS and/or Microsoft® Office Excel spread sheets. Example calculations are presented in Appendix 4.

4.0 **RESULTS AND DISCUSSION**

4.1 Method Establishment/Pre-Validation Evaluation

Prior to conducting the ILV, method control was established. Instrument parameters for the mass spectrometer detector were optimized by infusing standard solutions of the target analytes. The optimized instrument parameters, analyte retention times, instrument detection limits, and linearity were established by injecting a series of calibration standards

4.5 **Protocol Changes**

Protocol change number 1 was issued to update the experimental design section to instruct the fortification of control water samples should be performed separately in two validation sets due to the rapid degradation of thiodicarb to methomyl.

5.0 CONCLUSIONS

In summary, ADPEN Laboratories successfully and independently validated Bayer Analytical Method TD-002-W13-01 on the second trial.

The method was demonstrated to be suitable for the determination of thiodicarb and methomyl in water at an LOQ of 0.1 ppb and at $10 \times LOQ$. The method is well-written and contains sufficient information to guide the analyst through the procedure for the first time.

Transfer 10mL of sample water to a 50-mL conical centrifuge tube

TABLE 1.Flow Diagram of the Analytical Procedure

Prepare SPE manifold with PTFE needles, and attach labeled SPE cartridges Condition cartridges with 2mL of MeOH by gravity flow Condition cartridges with 2mL of HPLC Water by gravity flow Fortify recovery samples as required, using individual standard solutions (Cap and briefly shake to mix) Centrifuge samples @ 4,000 rpm for 10 minute Run 5-mL aliquot of sample through cartridge by gravity flow Dry cartridge by applying vacuum at ~20" Hg for 15 minutes (discard all eluates) J. Place a 15-mL centrifuge tube under each cartridge Elute with 2mL MeOH by gravity flow Dry cartridge by applying vacuum at \sim 5" Hg for 1 minute Elute with 1mL MeOH/DCM solution (3/1) and dry cartridge by vacuum (~5Hg for 1 minute) 1 Repeat above step T Dry samples in N-Evap @ 45°C, ~30psi (for approximately 30 minutes) t Reconstitute samples in 2.5mL L-Cysteine HCl in MeOH and vortex for 20 seconds T Transfer samples to LC vials, cap vials and samples are ready for analysis (Store samples in freezer $@ < -10 \degree$ C if not analyzed immediately)

Lab Code	Fortification Level (ppb)	Number of Samples			
Analytical set name: WO-13052401 (thiodicarb)					
13052401-MB-1	Method Blank	1			
130513004-001C, 130513004-001D	Control	2			
13052401-Recovery1-1, 13052401-Recovery1-2, 13052401-Recovery1-3, 13052401-Recovery1-4, 13052401-Recovery1-5	0.1	5			
13052401-Recovery2-1, 13052401-Recovery2-2, 13052401-Recovery2-3, 13052401-Recovery2-4, 13052401-Recovery2-5	1.0	5			
Analytical set name: WO-13052404 (methomyl)					
13052404-MB-1	Method Blank	1			
130513004-001E, 130513004-001F	Control	2			
13052404-Recovery1-1, 13052404-Recovery1-2, 13052404-Recovery1-3, 13052404-Recovery1-4, 13052404-Recovery1-5	0.1	5			
13052404-Recovery2-1, 13052404-Recovery2-2, 13052404-Recovery2-3, 13052404-Recovery2-4, 13052404-Recovery2-5	1.0	5			

TABLE 2. Water Fortification Levels