

MRID: 50105408

STUDY TITLE

Method Development and Validation of an Analytical Method for the Determination of BAS 720 H and its 2 Metabolites Reg. No 4110603 and Reg. No 4110542 in Water (Analytical Method L0209)

EPA Guideline(s)

OPPTS 850.6100

Study Identification Number(s)

BASF Study ID: 391110 SGS Project No.: IF-13/02484111

BASF Registration Document Number

2015/7008009

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3 Information about the Study

3.1 Test System

The specimen of surface water was obtained from a natural surface water course known as Burbach located in Dietzhoelztal, D-35716 in Germany. The specimen of ground water was obtained from a well located in Glashuetten, D-61479 in Germany . The specimens were given the following numbers:

ground water: 130206571 surface water: 130160769

See also Table 1 and 3.

4 Test / Reference Items

4.1 BAS 720 H

Common name Imazamox Reg. No. 4096483

IUPAC-Name (RS)-2-(4-isopropyl-4-methyl-5-oxo-2-imidazolin-2-yl)-5-

methoxymethylnicotinic acid

CAS-No. 114311-32-9 Molecular formula $C_{15}H_{19}N_3O_4$

Structure

Molecular weight 305.3 Lot No. AC12820-7

Appearance solid white (assayed by SGS-IF)

Purity 99.5 %

Storage advice keep in refrigerator or freezer

Expiry Date 01 May 2021

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4.2 Reg. No. 4110542

CL-No. 312622 Reg. No. 4110542

IUPAC-Name 2-(4-isopropyl-4-methyl-5-oxo-2-imidazolin-2-yl)pyridine-3,5-

dicarboxylic acid

CAS-No. 146953-32-4 Molecular formula $C_{14}H_{15}N_3O_5$

Structure

Molecular weight 305.3 Lot No. L82-7

Appearance solid white (assayed by SGS-IF)

Purity 88.4 %

Storage advice keep in refrigerator or freezer, hygroscopic (keep away from

humidity)

Expiry Date November 01, 2013

4.3 Reg. No. 4110603

CL-No. 354825 Reg. No. 4110603

IUPAC-Name 5-hydroxy-6-(4-isopropyl-4-methyl-5-oxo-2-imidazolin-2-yl)

nicotinic acid

CAS-No. -

Molecular formula C₁₃H₁₅N₃O₄

Structure

HO OH

Molecular weight 277.3 Lot No. L67-144

Appearance solid yellow-green (assayed by SGS-IF)

Purity 98.2 %

Storage advice Keep at room temperature (typically + 25°C) or cooler

Expiry Date July 01, 2020

These reference items and the respective data were supplied by the sponsor. Stock solutions and working solutions of the reference items were stored between + 2 °C and + 8 °C in capped vials protected from light.

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5 Purpose of the Study

The scope of this study was to validate an analytical method for the determination of BAS 720 H, Reg. No. 4110603 (CL-No. 354825) and Reg. No. 4110542 (CL-No. 312622) in water according to the guidelines mentioned in chapter 1.1 at a limit of quantification of 0.025 µg/L using LC-MS/MS for quantitation and confirmation.

6 Deviation to Study Plan

The analytical method L0209 was validated for determination of BAS 720 H and its metabolites Reg. No. 4110542 and 4110603 in water instead of the BASF methods M3515 (BASF Doc ID 2002/7006086) and BASF Method D1102.

7 Analytical Phase

7.1 Analytical Method

The used method L0209 was based on the existing BASF methods M 3513, BASF Doc ID 2002/7006086, "LC/MS Determination and LC/MS/MS Confirmatory Method for Residues of CL 299263, CL 312622 and CL 354825 in surface water" and BASF Method D1102; "Method for Determination of BAS 720 H and its Metabolites CL 312622 and CL 354825 Residues in Soil Using LC-MS/MS". This was done for the validation of an Analytical Method for the Determination of BAS 720 H and 2 Metabolites in Water.

Principle of the Method

The test systems ground water and surface water will be prepared for isolation of residues by solid phase extraction (SPE). The estimated limit of quantitation (LOQ) in ground water and surface water is 0.025 μ g/L for BAS 720 H and its metabolites Reg. No. 4110603 and Reg. No. 4110542.

In this study, two parent-daughter ion transitions will be determined by LC-MS/MS. One transition will be used for evaluation, the other one for confirmation.

7.2 Extraction

Add 10 g of the water to a 15 mL centrifuge tube followed by the addition of 0.5 mL of 2 N HCl. For fortification experiments specimens are fortified at this step.

7.3 Conditioning of C₁₈ SPE columns

Attach a Varian Bond Elut C-18 column on to a vacuum manifold by washing the column subsequently with 3 mL of each of the following solutions: hexane, DCM, methanol, water and 0.01 N HCl.

In a separate vacuum manifold, prepare the SCX column by eluting 3 mL of methanol through the column. Fill the column with 2 mL of methanol and set aside.

7.4 C-18 SPE Column only

Load the whole acidified specimen extract from step 6.2 to the C-18 column carefully. Wash the column with 3 mL of water followed by 3 mL of hexane. Stop the column when the liquid level passes just below the frit. Discard the wash.

Elute the above C-18 column with 3 mL of DCM and collect into a 15 mL glass centrifuge vial. Evaporate the DCM eluent at 40°C with a gentle nitrogen stream. Save the glass vial.

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7.5 C-18 column couples with SCX SPE column

Attach the C-18 column to the top of the SCX column (still containing the 2 mL of methanol from step 6.3)

Add additional 3 mL of methanol to the top of C-18 column and eluate. Detach and discard the C-18 column from the SCX column.

7.6 SCX SPE column only

Wash the SCX column with 3 mL of methanol and discard the wash.

Elute the SCX column with 6 mL of water/methanol (20:80, v/v) into the same glass vial used to collect the eluent from the C1-8 column. Swirl the solvent to combine and mix with a vortex.

7.7 Sample preparation for Analysis

Evaporate the combined eluent from step 6.6 using a nitrogen evaporator with the water bath set to 70°C until dryness.

Reconstitute once completely dry, all specimens in 0.1 % HCOOH in water/methanol (80/20, v/v). Vortex to obtain an homogeneous solution. The specimens are ready for analyses.

Final determination was performed by LC-MS/MS monitoring two parent daughter ion transitions. One transition was used for evaluation, the other one for confirmation.

7.8 Equipment and Materials

7.8.1 General Laboratory Equipment

- shaking machine Model SM 25 Bühler

- centrifuge Model Heraeus Megafuge R16, Thermo Fisher

analytical balance Sartorius, model CPA225D-OCE top loading balance Sartorius, model CPA 32025 + GS 3200-2

ultra sonic bath Bandelin, model DK 102

vortex Model reax 2000, Heidolph

Heidolph, model 11
nitrogen evaporator Model 112 Organomation

drying oven Modle FD 115, WTB Binder ultra pure water unit Millipore, model Synergy UV

centrifuge tubes Neolab, PP, 50 mL and 15 mL

test tube for GPC sampler glass 100x18x1 mm, with perforated PE-stopper,

Pasteur pipette Burdich glass

volumetric pipette different sizes volumetric flask different sizes

beaker different sizes
glass bottle different sizes
graduated cylinder different sizes

Dessicator
SPE manifold
Supelco

- injection vial, 1.8 mL, with
PTFE sealed crimp-on caps
Burdich GmbH

microvial
 sample vial
 500 μL fitting into the injection vial
 20 mL, Burdich

adapter mciropipettes BASF Study ID: 391110

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7.8.2 Reagents

dichloromethane Roth, No. T162.1

- methanol LGC Promochem SO-3041-B025

ultra pure water
 sodium hydroxide
 formic acid
 Millipore, Synergy UV
 VWR, No. 1.06498
 Merck 1.00264

- 1 mol/L Titrisol Merck

C₁₈ SPE cartridges (0.2 g, 3 mL) Bond Elute Varian, No. 12102025
 SCX SPE cartridges Isolute / Biotage 530-0020-B

7.8.3 Preparation of the Mixtures

2 mol/L HCl 1 mol HCl (Titrisol) in 500 mL ultrapure water

Washing solution 5 C-18 5 mL 2 mol/L HCl in 1 L ultrapure water

clean-up 0.01 mol/L HCl

Elution solvent 200 mL ultrapure water with 800 mL methanol

SCX SPE clean-up

0.1 % HCOOH in ultra pure 1 mL conc. formic acid in 1 L water

water

Diluent 800 mL 0.1 % HCOOH in water with 200 mL methanol

Mobile Phase I 1 mL conc. formic acid in 1 L ultrapure water

MobilePhase II 1 mL conc. formic acid in 1 L methanol

7.8.4 Final Determination of BAS 720 H, Reg. No. 4110603 and Reg. No. 4110542

The residues of BAS 720 H and its metabolites were determined using LC-MS/MS equipment. The reported LC-MS/MS conditions are general. They may vary as they were adapted in order to obtain appropriate detector sensitivity and peak separation.

HPLC System:

System: Agilent, No. 1200
Degasser: Agilent, G1322A
Pump: Agilent, G1311A

Injection system: CTC Analytics, HTC Pal

Column oven with 6 port valve: Agilent, G1316A Control module: Agilent, G4208A

HPLC-Conditions:

Analytical Column: Zorbax Eclipse XDB-C18

length: 50 mm interior diameter: 4.6 mm particle size: 1.8 µm Manufacturer: Agilent

Oven temperature: 40 °C

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Injection volume: 20 µL

Temperature of sample thermostat: 10 °C

Split (detector/waste): ---

Mobile Phase I:

Mobile Phase II:

0.1 % formic acid in water

0.1 % formic acid in methanol

Flow rate: see table below Stop time: 15 minutes

Gradient time table:	Time	Mobile	Mobile	Flow
		Phase I	Phase II	
	[min]	[%]	[%]	[mL/min]
	0	80	20	0.6
	0.5	80	20	0.6
	3.0	50	50	0.6
	7.0	5	95	0.6
	7.10	5	95	0.6
	10.0	5	95	0.6
	10.1	80	20	0.6
	15.0	80	20	0.6

Switching times for valve in column oven: 0 - 3.5 min (waste)

3.5 - 9.5 min (LC-MS/MS)

9.5 - 15 min (waste)

Retention times:

BAS 720 H: 5.2 min Reg. No.4110603: 7.1 min Reg. No. 4110542: 4.7 min

LC-MS/MS System:

System: Applied Biosystems, API 4000

Vacuum pump: HS602

Data system: Applied Biosystems, Analyst, version 1.5.1

LC-MS/MS Conditions:

		BAS 720 H
Scan type:	MRM	MRM
Polarity:	positive	positive
Curtain gas:	10	10
Collision gas:	8	8
Ionization voltage:	5500	5500
Temperature:	500	500
Entrance potential:	10	10
Declustering potential:	86	86
Collision energy:	31	43
Collision cell exit potential:	14	16
Transition used for	$306.3 \rightarrow 261.2$	$306.3 \rightarrow 86.3$
evaluation:	(Quantification)	(Confirmation)

Rea	. No.	411	0603

Scan type:	MRM	MRM
Polarity:	positive	positive
Curtain gas:	10	10
Collision gas:	8	8
Ionization voltage:	5500	5500
Temperature:	500	500
Entrance potential:	10	10
Declustering potential:	96	96
Collision energy:	29	41
Collision cell exit potential:	16	10
Transition used for	$278.1 \rightarrow 233.1$	$278.1 \rightarrow 165.0$
evaluation:	(Quantification)	(Confirmation)

Reg. No. 4110542

	- 3	
Scan type:	MRM	MRM
Polarity:	positive	positive
Curtain gas:	10	10
Collision gas:	8	8
Ionization voltage:	5500	5500
Temperature:	500	500
Entrance potential:	10	10
Declustering potential:	71	71
Collision energy:	31	27
Collision cell exit potential:	16	6
Transition used for	$306.3 \rightarrow 261.2$	$306.3 \rightarrow 264.2$
evaluation:	(Quantification)	(Confirmation)

7.9 Preparation of Solutions for Fortification and Calibration

Immediately after preparation, the stock, fortification and calibration solutions were stored in capped vials protected from light in a refrigerator at 2-8 °C.

7.9.1 Stock and Fortification Solutions

Three stock solutions of BAS 720 H, Reg. No 4110603 and Reg. No 4110542 were prepared on 26 February 2013.*

Example:

On 26 February 2013, a stock solution of BAS 720 H, Reg. No 4110603 and Reg. No. 4110542 were prepared each in a volumetric flask by dissolving 11.07 mg in Methanol, 10.58 mg in Methanol and 11.89 mg in Methanol. This resulted in a concentration of 1.101 mg/mL, 1.039 mg/mL and 1.051 mg/mL taking into account a purity of 99.5 %, 98.2 and 88.4 % respectively.

^{*} The stock solutions of BAS 720 H, Reg. No. 4110603 and Reg. No. 4110542 were prepared on 26 February 2013 and used for the BASF study 391104_1. These same stock solutions were simultaneously used for the study 391110.

The stock solution prepared on 26 February 2013 was dissolved daily fresh on 21 March 2013 with methanol to concentrations of 10.06 to 0.2012 μ g/mL for BAS 720 H, 10.04 to 0.2008 μ g/mL for Reg. No. 4110603 and 10.02 to 0.2004 μ g/mL for Reg. No. 4110542 which were used for fortification.

These were used for the preparation of calibration solutions and fortification of the specimens.

7.9.2 Calibration Solutions

Example:

On 26 February 2013,a diluted stock solution of BAS 720 H, Reg. No. 4110603 and Reg. No 4110542 (concentration: 1.006, 1.004 and 1.002 μ g/mL respectively) were dissolved in a 0.1 % HCOOH in H₂O/MeOH (80/20, v/v) solution to concentrations ranging from 0.0503 ng/mL to 2.515 ng/mL (BAS 720 H), 0.0502 to 2.515 ng/mL (Reg. No. 4110603) and 0.0501 to 2.505 ng/mL (Reg. No. 4110542) which were prepared for calibration.

No matrix matched calibration standards were necessary and therefore not used in this study.

Comparison between solvent standards and matrix-matched standard solutions

A solvent standard was compared against a matrix-matched standard for all analytes, Results are presented in the following table:

Analyte	Imazamox	Imazamox	Reg. No. 4110603	Reg. No. 4110603	Reg. No. 4110542	Reg. No. 4110542
Mass	306 > 261	306 > 86	278 > 233	278 > 165	306 > 261	306 > 264
transition						
m/z						
C std [ng/mL]	0.2515	0.2515	0.2505	0.2505	0.2510	0.2510
Peak area	6381	2649	3165	1721	9099	3003
Solvent						
Matrix	surface wat	er				
Peak area	6227	3038	3639	1668	9173	3050
Matrix						
Signal	-2	15	15	-3	1	2
Enhancement/						
Suppresion						
by the matrix						
for the analyte						
Matrix	ground water	er				
Peak area	7042	2826	3326	1522	8966	2877
Matrix						
Signal	10	7	6	-12	-1	-4
Enhancement/						
Suppresion						
by the matrix						
for the analyte						

^{*}A negative value indicates signal suppression, while a positive value means signal enhancement

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7.10 Calculations

The detector signals were registered and integrated using the data system outlined in chapter 6.8.4. The peak area was taken into account to determine the BAS 720 H, Reg. No. 4110603 and Reg. No 4110542 amount in the specimens. The calibration curves were calculated from the area of the calibration solutions with their corresponding concentrations of the analytes using equation (1).

$$y = a + bx$$

where y: peak area [integration units iu]

x: amount of analyte [ng]a: ordinate intercept [iu]

b: slope [iu/ng]

The amount of BAS 720 H, Reg. No. 4110603 and Reg. No. 4110542 in the specimen was calculated using the transformed equation (1):

$$x = \frac{y - a}{b}$$

The concentration C of BAS 720 H, Reg. No. 4110603 and Reg. No. 4110542 in the specimen was calculated from x using equation (3):

(3)
$$C = \frac{x \cdot V_E \cdot A_1}{V_i \cdot A_2 \cdot W}$$

where x : amount of analyte [ng]

C : analysed concentration of the respective analyte in the

specimen [µg/L]

 V_E : final volume (1 and 5 mL) A_1 : total extract (10 mL) V_i : injection volume (20 μ L)

A₂: aliquot (10 mL)

W: specimen weight (for 10 mL)

The recovery data was calculated according to equation (4):

(4)
$$R = \frac{C \cdot 100}{C_{for}}$$

where R: recovery [%]

C: analysed concentration of the respective analyte in the fortified

specimen

Cfor: nominal concentration of respective analyte in the fortified

specimen

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7.11 **Example for Calculation**

The fortified specimen of ground water 130206571-A was extracted and analysed on 21 March 2013. The parameters of the calibration curve for BAS 720 H on the basis of the detected mass transition m/z 306 \rightarrow 261 was a = 6.923 (ordinate intercept), b = 1455807.07 derived from equation (1). The peak area acquired of the analyte BAS 720 H corresponded to 6702 counts.

The amount of BAS 720 H was calculated according to equation (2):

$$x = \frac{6702 - 6.923}{1455807} = 0.0046 [ng]$$

The residue concentration was calculated according to equation (3):

$$C_R = \frac{0.0046 \cdot 10 \cdot 1}{10 \cdot 1 \cdot 20} = 0.023 \left[\frac{\mu g}{L} \right]$$

where

X: 0.0046 [ng]

 C_R : analysed concentration of the respective analyte

 $[\mu g/L]$

V_E: 1 [mL] A₁: 10 [mL] V_i: 20 [μL]

A₂: 10 [mL] 10 [mL]

The recovery data were calculated according to equation (4).

$$R = \frac{0.023 \cdot 100}{0.025} = 91\% \tag{4}$$

where

recovery rate [%] R: C_R : 0.023 [µg/L] 0.025 [µg/L]

The tabulated values were rounded values based on calculations with the exact raw data.

The difference between the old and the fresh stock solutions of BAS 720 H was 3.4 %, proving that solutions of the reference item were stable for the duration of the laboratory work.

Stability of the Reference Item Reg. No. 4110603 during the Study (m/z 278→ 233)

Fresh Sto	Fresh Stock Standard Solution			Old Stock Standard Solution		
Stock Solution	prepared on	26 Mar 2013	Stock Solution	prepared on	26 Feb 2013	
Standard Solution	diluted on	26 Mar 2013	Standard Solution	diluted on	26 Mar 2013	
	The date of analysis was 26 Mar 2013					
Concentration	Peak		Concentration	Peak		Percentage
[ng/mL]	Area	Quotient	[ng/mL]	Area	Quotient	[%]
1.002	36169	36097	1.004	36788	36641	101.5

The difference between the old and the fresh stock solutions of Reg. No. 4110603 was 1.5 %, proving that solutions of the reference item were stable for the duration of the laboratory work.

Stability of the Reference Item Reg. No. 4110542 during the Study (m/z $306 \rightarrow 261$)

1.1.0 = 0.00						
Fresh Sto	Fresh Stock Standard Solution			Old Stock Standard Solution		
Stock Solution	prepared on	26 Mar 2013	Stock Solution	prepared on	26 Feb 2013	
Standard Solution	diluted on	26 Mar 2013	Standard Solution	diluted on	26 Mar 2013	
		The date of an	alysis was 26 Mar 20	013		
Concentration	Peak		Concentration	Peak		Percentage
[ng/mL]	Area	Quotient	[ng/mL]	Area	Quotient	[%]
1.004	13349	13296	1.002	13010	12984	97.7

The difference between the old and the fresh stock solutions of Reg. No. 4110542 was 2.3 %, proving that solutions of the reference item were stable for the duration of the laboratory work.

Since no matrix-matched standards were used within this study, these were not tested for stability during laboratory work.

8.7 Stability tests with Specimen extracts

In order to test the stability of the reference items in the final extracts (after clean-up treatments), an aliquot from these specimens fortified at 10times LOQ of the matrices ground water and surface water were diluted and analysed after 7 days. All stability analyses were determined using the quantification and confirmation ion transitions for all substances.

Storage temperature of the final specimen extracts: 2-8 °C

Storage period: 21.03.2013 (Extraction) to 28.03.2013 (Re-analyses)

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11 Tables

Table 1: Detailed Information about the Specimen used for Fortification

Matrix]	Specimen number	
	Specimen received Specimen homogenisation		SGS INSTITUT FRESENIUS
surface water ground water	1	n.a. n.a.	130160769 130206571

The specimen of surface water was stored at 2-8 °C and the specimen of ground water was stored deep frozen (\leq -18 °C) prior to extraction.

Table 2: Preparation of the Fortified Specimens

ground water

IF-Specimen No.	Weight of Specimen	Volume of Fortification Solution	Concentration of Fortification Solution	Concentration of Fortification Solution	Concentration of Fortification Solution	Date of Specimen Preparation
	[g]	[mL]	lmazamox [µg/L]	Reg. No. 4110603 [µg/L]	Reg. No. 4110542 [µg/L]	
130206571-1	10.05					21-Mar-2013
130206571-2	10.00					21-Mar-2013
130206571-A	10.00	0.125	0.025	0.025	0.025	21-Mar-2013
130206571-B	10.00	0.125	0.025	0.025	0.025	21-Mar-2013
130206571-C	10.02	0.125	0.025	0.025	0.025	21-Mar-2013
130206571-D	10.01	0.125	0.025	0.025	0.025	21-Mar-2013
130206571-E	10.00	0.125	0.025	0.025	0.025	21-Mar-2013
130206571-F	10.01	0.125	0.25	0.25	0.25	21-Mar-2013
130206571-G	10.01	0.125	0.25	0.25	0.25	21-Mar-2013
130206571-H	10.04	0.125	0.25	0.25	0.25	21-Mar-2013
130206571-K	10.02	0.125	0.25	0.25	0.25	21-Mar-2013
130206571-L	10.03	0.125	0.25	0.25	0.25	21-Mar-2013

surface water

IF-Specimen No.	Weight of Specimen	Volume of Fortification Solution	Concentration of Fortification Solution	Concentration of Fortification Solution	Concentration of Fortification Solution	Date of Specimen Preparation
	[g]	[mL]	lmazamox [µg/L]	Reg. No. 4110603 [µg/L]	Reg. No. 4110542 [µg/L]	
130160769-1	10.00					21-Mar-2013
130160769-2	10.02					21-Mar-2013
130160769-A	10.01	0.125	0.025	0.025	0.025	21-Mar-2013
130160769-B	10.01	0.125	0.025	0.025	0.025	21-Mar-2013
130160769-C	10.00	0.125	0.025	0.025	0.025	21-Mar-2013
130160769-D	10.00	0.125	0.025	0.025	0.025	21-Mar-2013
130160769-E	10.02	0.125	0.025	0.025	0.025	21-Mar-2013
130160769-F	10.00	0.125	0.25	0.25	0.25	21-Mar-2013
130160769-G	10.00	0.125	0.25	0.25	0.25	21-Mar-2013
130160769-H	10.02	0.125	0.25	0.25	0.25	21-Mar-2013
130160769-K	10.00	0.125	0.25	0.25	0.25	21-Mar-2013
130160769-L	10.01	0.125	0.25	0.25	0.25	21-Mar-2013

Table 3: **Water Parameters**

Parameter Determined	Result					
surface water (130160769)						
temperature	4.9 °C					
Redox potential	308 mV					
Conductivity	82 μS/cm					
Water hardness	0.0169 mmol/L					
pH value	7.81					
Dissolved organic Carbon (DOC)	0.931 mg/L					
ground water (130206571)						
temperature	6.4 °C					
Redox potential	252 mV					
Conductivity	45 μS/cm					
Water hardness	0.0049 mmol/L					
pH value	6.00					
Dissolved organic Carbon (DOC)	0.116 mg/L					