

NAVAL SURFACE WARFARE CENTER DAHLGREN DIVISION

SCIENCE AND TECHNOLOGY - RESEARCH AND DEVELOPMENT - TEST AND EVALUATION



ELECTROMAGNETIC & SENSOR SYSTEMS DEPARTMENT Combining Spore Germination and Hot Air Treatment to Reduce the Costs and Time and Lower the Temperature for Hot Air Decontamination

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> > Presented by

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Objective

Buhr et al. 2012, 2014, 2015; Herzberg 2012; Koch et al. 2001



<u>Objective</u>: The objective is to add a spore germination step prior to hot air decontamination in order to reduce decontamination temperature and time requirements from the current Joint Biological Agent Decontamination System (JBADS) requirements, which are \geq 75°C, \geq 72h, 70-90% relative humidity (RH), down to \leq 60°C for \leq 24h.

Need for Decontamination:

Commercial aircraft temperature materials are typically tested at 140°F (60°C). This does not imply that materials won't survive higher temperature, but the cost of changing the test temperature for qualifying all materials is high. DoD accepts risk of higher temperatures.

Lower decontamination temperature and times translates to lower costs, higher practicality, increased applications





B. anthracis Spore, Germination and Outgrowth

Buhr et al. (2012) – Dr. Derrell McPherson; *B. anthracis* germinated spore TEM courtesy Dr. Chris Cote (USAMRIID)



B. anthracis – Dormant spore, germinated spore, outgrown spore



B. atrophaeus (Bg) Dormant spore







B. atrophaeus (Bg) Outgrown spore







Standardized *Bacillus* Spore Preparation for Macrobacillus Spores Containing Exosporia





Anonymous. 2018. Standard Practice for Evaluating Efficacy of Vaporous Decontaminants Against *Bacillus* Spores Contained Within 0.2µm Filter-capped Tubes. ASTM Standard Practice E3092.

Cote, C., Buhr, T., Bernhards, C.B., Fetterer, D.P., Welkos, S.L., Bozue, J.A., McPherson, D., Fountain, A.W. III, Gibbons, H.S. and the *Bacillus anthracis* Spore Irradiation Consortium. (2018) A Standard Method to Inactivate *Bacillus anthracis* Spores to Sterility Using Irradiation. Submitted to Applied and Environmental Microbiology.

Plaut, R., Staab, A., Munson, M., Gebhardt, J., Klimko, C., Quirk, A., Cote, C., Buhr, T., Rossmaier, R., Bernhards, R., Love, C., Berk, K., Abshire, T., Rozak, D., Beck, L., Stibitz, S., Goodwin, B., Smith, M., Sozhamannan, S. (2018) Construction and Characterization of an Avirulent *Bacillus anthracis* Strain Carrying Molecular Assay Targets as a Surrogate for Irradiation-inactivated Virulent Spores. Emerging and Infectious Diseases, in review: Emerging Infectious Diseases 17-1646-R1.





Original Test Design was based on a single germinant application with an estimated germination temperature of 35°C, unknown time, followed by 60°C, 1 h

- What is the optimized germinant solution?
- Is germination efficiency sufficient to meet decontamination test requirements?
 - $\geq 7 \log_{10}$ spores/test to show at least a 6 \log_{10} spore inactivation
 - $\geq 8 \log_{10} \text{ spores/m}^2 \text{ (coupon tests are 2.5e10 } \log_{10} \text{ spores/m}^2 \text{)}$
- Is heat inactivation needed?
- What are the germination limits (Temperature, RH, Time)?
 - Target temperature is 35°C. Is germination efficient at >35°C?
 - Target time is unknown, but short duration (<20h is a target).
 - Target RH is unknown.
- What are the heat inactivation limits (Temperature, RH, Time)?
 - Target temperature is 60°C with a range of 50-60°C.
 - Target time is 1 h. This is short duration.
 - Target RH is unknown.
- Can a single germinant application be used?
- Can a fogger be used?





Germination for Optimized <u>Solution Testing</u> with KOH. Results: Heat Inactivation is Required; Quantitative Detection in Field is a Capability

Gap

Buhr et al. Quarterly Report (2016)

Spore germination assayed by serial dilution and plating shows total \log_{10} colony forming unit (CFU) ml⁻¹ and heat-resistant CFU ml⁻¹ survival of *B. anthracis* Δ Sterne, *B. thuringiensis* Al Hakam or *B. thuringiensis* kurstaki cry- HD 1 after incubation at 35°C and 200 rpm for 0, 1, 2, 4 and 24 hours in the 0.1M L-Alanine + 1mM Inosine + 40mM CaCl2 + 40mM DPA + 20mM Tris Base + 70mM KOH germination solution. Data is the mean of three independent experiments.

	Inoculation			Tin	ne points	(h)	
Species	Titer log10 (Spores ml ⁻¹)	30 min, 65°C Heat	0	1	2	4	24
	5.2	no	5.2	4.9	4.7	4.4	0.6
	5.2	yes	5.1	2.2	1.0	0.1	0.2
	6 1	no	6.1	5.7	6.0	5.6	1.4
Ba∆St	0.1	yes	6.0	2.7	1.7	0.7	0.2
	7 0	no	7.2	6.6	6.9	6.8	3.0
	1.2	yes	6.7	3.5	2.8	2.3	24 0.6 0.2 1.4 0.2 3.0 1.6 1.3 0.0 4.2 1.1 5.8 4.9 0.0 0.0 0.0 2.7 0.2 4.3 2.2
	5 2	no	5.2	5.2	5.2	5.0	1.3
	5.2	yes	5.3	0.4	0.0	0.0	0.0
R+AH	61	no	6.1	6.2	6.1	5.7	4.2
DIAN	0.1	yes	6.2	1.2	1.2	0.6	1.1
	73	no	7.3	7.2	7.0	6.8	5.8
	7.5	yes	7.4	2.2	1.4	1.7	4.9
	5.2	no	5.2	5.2	5.3	4.5	0.0
Btk cry- HD1	5.2	yes	5.2	0.7	0.2	0.0	0.0
	6.2	no	6.2	6.2	6.2	5.6	2.7
	0.2	yes	6.2	1.8	1.0	0.6	0.2
	7.2	no	7.2	7.3	7.2	7.0	4.3
	1.2	yes	6.9	2.4	1.9	1.5	2.2



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Two Cycles of Germination and Heat Inactivation shows total \log_{10} CFU survival of *B. anthracis* Δ Sterne spores dried on to nylon webbing. Germination occurred at 35°C, 90% RH in 0.1M L-Alanine + 1mM Inosine + 40mM CaCl2 + 40mM DPA + 20mM Tris Base + 70mM KOH and heat inactivation was at 60°C, 90% RH. Five independent replicates were tested.

	Single Germination and Decontamination Step	Repeat Germination and Decontamination Steps										
	Test A	Test B	Test C	Test D	Test E	Test F	Test G	Test H				
Sample	2h Germination	2h Germination	2h Germination	2h Germination	2h Germination	2h Germination	2h Germination	2h Germination				
		1h Decon;	1h Decon;	1h Decon;	1h Decon;	1h Decon;	1h Decon;	1h Decon;				
	1h Decon	2h Germination	16h Germination	16h Germination	20h Germination	20h Germination	24h Germination	24h Germination				
		1h Decon	1h Decon	No Decon	1h Decon	No Decon	1h Decon	No Decon				
Test - Nylon	2.4 ± 0.5	1.4 ± 0.5	0.6 ± 0.3	0.8 ± 0.9	0.5 ± 0.6	0.6 ± 1.0	0.2 ± 0.3	0.5 ± 0.7				
Dry Extraction Control - Nylon	6.7 ± 0.0	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.8 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1				
Wet Extraction Control - Nylon	6.5 ± 0.1	6.6 ± 0.1	6.4 ± 0.1	6.4 ± 0.2	6.3 ± 0.2	6.5 ± 0.1	6.6 ± 0.1	6.5 ± 0.2				
Test - Solution Control	7.3 ± 0.3	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.2	7.3 ± 0.2	7.2 ± 0.1	7.3 ± 0.0	7.3 ± 0.1				
Ca-DPA Media 1st Germination	6.0 ± 0.1	6.0 ± 0.1	6.1 ± 0.1	6.0 ± 0.1	6.1 ± 0.1	6.0 ± 0.1	6.1 ± 0.1	6.1 ± 0.1				
Ca-DPA Media 2nd Germination	NA	0.8 ± 0.5	0.7 ± 1.0	0.0 ± 0.0	0.4 ± 0.6	0.0 ± 0.0	0.4 ± 1.0	0.0 ± 0.0				
RT - Nylon	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1	6.8 ± 0.1				
RT - Solution Control	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1				



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Germinant combinations with lower salt concentrations.

Germination Medium	Description
0.1M L-Alanine + 1mM Inosine + 40mM CaCl2 + 40mM DPA + 20mM Tris Base + 70mM KOH, pH 8.3	Optimized germination medium
0.1M L-Alanine + 1mM Inosine + 20mM CaCl2 + 20mM DPA + 10mM Tris Base + 35mM KOH, pH 8.3	Optimized germination medium with salts reduced to 50%.
0.1M L-Alanine + 1mM Inosine + 40mM CaCl2 + 40mM DPA, pH 8.3 w/ Tris Base	Optimized germination medium with no KOH added. Tris base was used to achieve pH of 8.3.
0.1M L-Alanine + 1mM Inosine, pH 8.3 w/ Tris Base	0.1M L-Alanine + 1mM Inosine only with Tris Base used to achieve pH of 8.3.
0.1M L-Alanine + 1mM Inosine, pH 6.1	0.1M L-Alanine + 1mM Inosine only with no pH adjustment performed.







	2 h germination; 1 h decontamination								
	Test A	Test B	Test C	Test D					
Sample	Optimized germination medium with KOH	Optimized germination medium 50% salts	Optimized germination medium w/o KOH	0.1M L- Alanine + 1mM Inosine pH 8.3					
Test - Nylon	3.2 ± 0.7	3.9 ± 0.5	3.1 ± 0.8	4.7 ± 0.9					
Dry Extraction Control - Nylon	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1					
Wet Extraction Control - Nylon	6.7 ± 0.1	6.7 ± 0.1	6.7 ± 0.1	6.7 ± 0.1					
Test - Solution Control	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1	7.3 ± 0.1					
RT - Nylon	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1					
RT - Solution Control	7.2 ± 0.0	7.2 ± 0.0	7.2 ± 0.0	7.2 ± 0.0					







Two-cycle germination + heat inactivation comparing four germination media. Total log_{10} CFU survival of *B. anthracis* Δ Sterne spores on nylon. Germination was tested at 35°C, 90% RH and heat inactivation at 60°C, 90% RH. Five independent replicates were tested per condition.

	2 h germination; 1 h decontamination / 16 h germination, 1 h decontamination									
Sample	Test A	Test B	Test C	Test D	Test E	Test F	Test G			
	1 st germination: Optimal germination medium; 2 nd germination: Optimal germination medium	1 st germination: Optimal germination medium 50% salts; 2 nd germination: Optimal germination medium 50% salts	1 st germination: Optimal germination medium w/o KOH; 2 nd germination: Optimal germination medium w/o KOH	1 st germination: 0.1M L- Alanine + 1mM Inosine pH 8.3; 2 nd germination: 0.1M L- Alanine + 1mM Inosine pH 8.3	1 st germination: Optimal germination medium; 2 nd germination: 0.1M L- Alanine + 1mM Inosine pH 8.3	1 st germination: Optimal germination medium 50% salts; 2 nd germination: 0.1M L- Alanine + 1mM Inosine pH 8.3	1 st germination: Optimal germination medium w/o KOH; 2 nd germination: 0.1M L- Alanine + 1mM Inosine pH 8.3			
Test - Nylon	1.6 ± 0.7	1.8 ± 0.4	1.2 ± 0.9	2.6 ± 0.8	1.3 ± 0.9	2.4 ± 0.7	1.8 ± 0.9			
Dry Extraction Control - Nylon	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1			
Wet Extraction Control - Nylon	6.6 ± 0.1	6.6 ± 0.1	6.6 ± 0.1	6.6 ± 0.1	6.6 ± 0.1	6.6 ± 0.1	6.6 ± 0.1			
Test - Solution Control	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1			
RT - Nylon	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1	6.9 ± 0.1			
RT - Solution Control	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1	7.2 ± 0.1			



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Fog tests to date: No spore inactivation.

Conclusion after fog testing: Fogging is not a good application method. Can't get enough water into the system. Spraying is the only current option.







Estimated material quantity and costs for applying a single application of the optimized germinant solution without KOH to military equipment.

Quantities are based upon a 0.2 ml coupon⁻¹ or 2 l m⁻² from preliminary spray testing.

Note: these are with Sigma prices. Bulk wholesale prices should be significantly lower, probably 10x less.

	Est Surface	Germinant Quantity for Single					Tris	Total
Application	Area	Application	L-Ala	Inosine	DPA	CaCl ₂	Base	Cost
Application	(m^2)	(Liters)	(kg)	(kg)	(kg)	(kg)	(kg)	(\$K)
C-130J (Interior)	250	500	4.45	0.13	3.34	4.38	7.57	3.285
C-130J (Exterior)	400	800	7.13	0.21	5.35	7.01	12.11	5.255
HMMWV (Exterior)	50	100	0.89	0.03	0.67	0.88	1.51	0.657
M1126 Skryker								
(Exterior)	100	200	1.78	0.05	1.34	1.75	3.03	1.314
Abrams Tank								
(Exterior)	125	250	2.23	0.07	1.67	2.19	3.79	1.642





B. anthracis Δ Sterne spore germination after incubation of spores over time and at different temperatures at spore concentrations of 7.0-7.3 log₁₀ (1-2e7) spores ml⁻¹. The colored lines represent log₁₀ survival of spores evaluated after mixing spores with germination medium and then heat treatment at 60°C, 1h or 65°C, 30 min at times of 0h (black), 1h (blue), 2h (red), 4h (green) and 24h (purple).





What are the Limits for Germination Temperature, Time, and Spore Concentration?



	Inoculation	Heatshock		Time p	oints (h)		
Temp	Titer log ₁₀	60°C for 60 min	1	2	4	24	
	5.2	no	5.4	5.3	5.4	4.7	
		3.5	yes	1.3	0.8	0.0	0.0
	63	no	6.3	6.4	6.3	6.0	
	0.5	yes	2.2	1.6	1.1	0.0	
1500	74	no	7.3	7.3	7.3	7.3	
15°C	7.4	yes	3.1	2.7	2.0	1.3	
	84	no	8.4	8.3	8.3	8.4	
	0.4	yes	4.6	3.4	2.5	1.6	
	0.4	no	9.4	9.3	9.3	9.4	
	9.4	yes	6.3	4.7	3.7	2.1	
	Inoculation Titer log10 5.3 6.3 7.4 8.4 9.4 5.2 6.1 7.2 8.2 9.2 5.2 6.1 7.2 8.2 9.2 5.2 6.1 7.1 8.2	no	5.1	5.1	5.1	2.4	
		yes	1.0	0.7	0.0	0.0	
	61	no	6.1	6.1	6.2	4.4	
	0.1	yes	1.9	1.4	0.5	0.0	
2500	7.2	no	7.1	7.2	7.2	6.3	
25°C	1.2	yes	3.0	2.6	1.8	2.0	
	82	no	8.1	8.2	8.2	8.1	
	0.2	yes	4.2	3.5	2.8	2.6	
	0.2	no	9.2	9.2	9.2	9.1	
	5.3 no 6.3 yes 6.3 yes 7.4 yes 7.4 yes 8.4 no 9.4 yes 9.4 yes 9.4 yes 5.2 no 9.4 yes 6.1 no 7.2 yes 6.1 yes 7.2 no 9.2 yes 6.2 no 9.2 yes 6.2 no 9.2 no	6.6	5.1	4.3	4.1		
	5.2	no	5.2	5.1	3.3	0.5	
	5.2	yes	0.9	0.6	0.0	0.0	
	62	no	6.2	6.2	4.7	1.5	
	0.2	yes	1.8	1.5	0.5	0.0	
2500	71	no	7.2	7.2	6.1	2.6	
	/.1	yes	2.8	2.4	2.0	1.5	
	82	no	8.2	8.3	7.9	4.3	
	0.2	yes	4.3	4.1	4.1	4.2	
	0.2	no	9.2	9.2	9.1	8.0	
	9.2	Ves	79	8.0	8.0	8.1	



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What are the Limits for Heat Inactivation Temperature, ie 50°C versus 60°C?



	Inoculation	60°C for							
Temp	Titer	60 min	1	2	4	24	48	7 d	
	5.2	Yes	3.2	2.6	1.7	0.3	0.0	0.0	
	6.0	Yes	3.5	3.0	2.8	1.8	1.3	0.0	
	7.1	Yes	5.0	4.7	4.2	3.1	2.6	1.6	
35°C	8.2	Yes	4.2	4.0	4.0	3.9	3.8	3.8	
	9.3	yes	7.7	7.7	7.8	7.8	7.7	7.8	
Tomp	Inoculation	50°C for							
Temp	Titer	60 min	1	2	4	24	48	7 d	
	5.1	yes	3.2	3.0	2.3	0.8	0.0	0.0	
Ba∆St 35°C	6.0	yes	3.9	4.2	3.6	2.0	1.3	0.0	
	7.0	yes	6.6	5.3	4.6	3.3	2.6	1.7	
	8.3	yes	6.9	7.0	7.2	4.6	3.8	3.7	
	9.3	yes	9.1	8.9	8.3	7.8	7.7	7.7	







- Optimal Germination solution was established
- Goal of $\leq 60^{\circ}$ C and ≤ 24 hours should be achievable
- Repeat application of germinant, ie. germination for 0.5-2 hours followed by 60°C for 1 hour (No RH) followed by germination and heat treatment is the most reliable approach
- Spray application appears to be the best approach
- Test results for Spore Concentration, Germination Temperature, and Germination Time suggests that Germination has wider applicability than originally thought
- What is needed?
 - •Statistics
 - •Repeat application on coupons at lower germination temperatures
 - Addition of humic acid to system
 - •Modeling of germination temperature, time and spore concentration
 - •Identifying Heat Susceptible Molecular Targets may be Helpful
 - •Field testing





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- U Chicago Loyola Dr Adam Driks and Mark Khemmani
- Numerous Academians and Government Agencies have worked on germination
- Numerous Wide Area Decontamination collaborators have worked on Germination

