

# Disinfection of Biofilm-Associated *Legionella pneumophila*

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2019 U.S. EPA International Decontamination Research and  
Development Conference



## Legionella spp.

### 66 species (serogroups)

<i>L. adelaidensis</i>	<i>L. fairfieldensis</i>	<i>L. lytica</i>	<i>L. sainthelensi</i> (2)
<i>L. anisa</i>	<i>L. fallonii</i>	<i>L. maceachernii</i>	<i>L. santicrucis</i>
<i>L. beliardensis</i>	<i>L. feeleii</i> (2)	<i>L. massiliensis</i>	<i>L. shakespearei</i>
<i>L. birminghamensis</i>	<i>L. geestiana</i>	<i>L. micdadei</i>	<i>L. spiritensis</i>
<i>L. bozemanii</i> (2)	<i>L. genomospecies</i>	<i>L. monrovica</i>	<i>L. steelei</i>
<i>L. brunensis</i>	<i>L. gormanii</i>	<i>L. moravica</i>	<i>L. steigerwaltii</i>
<i>L. busanensis</i>	<i>L. gratiana</i>	<i>L. nagasakiensis</i>	<i>L. saoudiensis</i>
<i>L. cardiaca</i>	<i>L. gresilensis</i>	<i>L. nautarum</i>	<i>L. taurinensis</i>
<i>L. cherrii</i>	<i>L. hackeliae</i> (2)	<i>L. norrlandica</i>	<i>L. thermalis</i>
<i>L. cincinnatiensis</i>	<i>L. impletisoli</i>	<i>L. oakridgensis</i>	<i>L. tucsonensis</i>
<i>L. clemsonensis</i>	<i>L. israelensis</i>	<i>L. parisiensis</i>	<i>L. tunisiensis</i>
<i>L. donaldsonii</i>	<i>L. jamestowniensis</i>	<i>L. pittsburghensis</i>	<i>L. wadsworthii</i>
<i>L. drancourtii</i>	<i>L. jeonii</i>	<i>L. pneumophila</i> (15)	<i>L. waltersii</i>
<i>L. dresdenensis</i>	<i>L. jordanis</i>	<i>L. quateirensis</i>	<i>L. worsleiensis</i>
<i>L. drozanskii</i>	<i>L. lansingensis</i>	<i>L. quinlivanii</i> (2)	<i>L. yabuuchiae</i>
<i>L. dumoffii</i>	<i>L. londiniensis</i>	<i>L. rowbothamii</i>	
<i>L. erythra</i> (2)	<i>L. longbeachae</i> (2)	<i>L. rubrilucens</i>	

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19 species  
associated with  
human disease

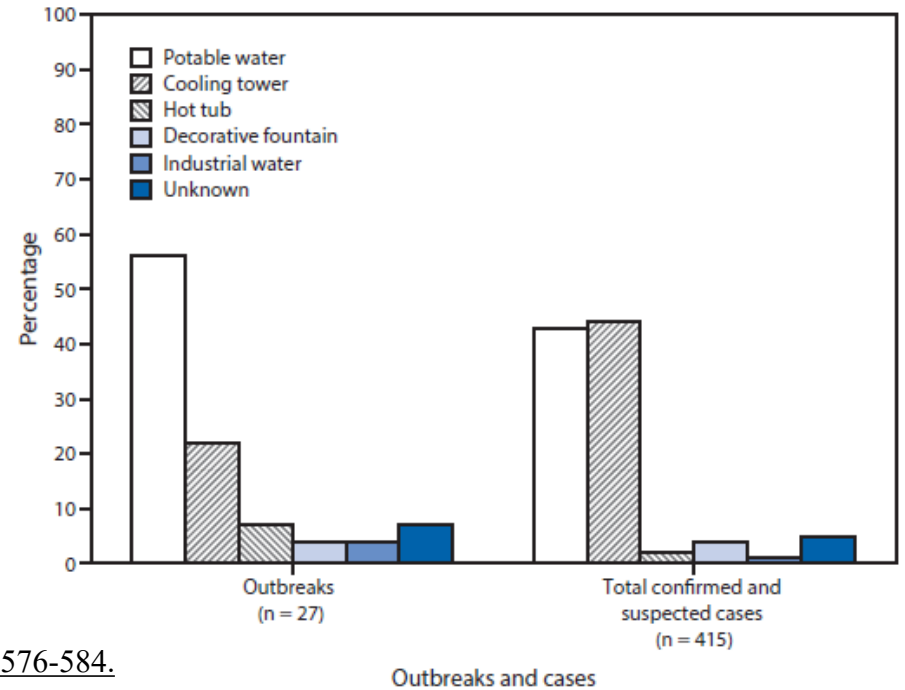
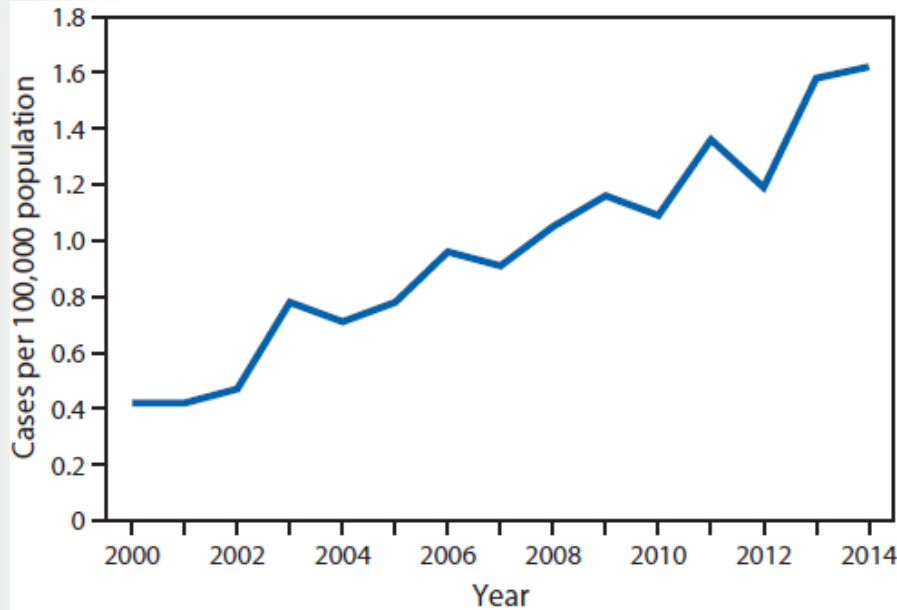
Dresden panel  
of mAbs (17),  
groups Lp into  
sg 1 to 15

Lück, C. et al. 2013.  
Typing Methods for  
*Legionella*, p. 119-  
148. In C.  
Buchrieser and H.  
Hilbi (ed.),  
*Legionella*.

sg1, >80% clinical  
isolates



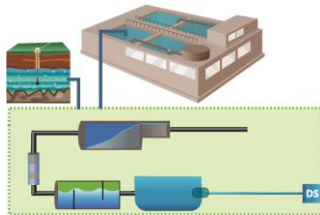
# Legionnaires Disease – A Public Health and Economic Burden



Garrison, L. E., et al. (2016). *MMWR Morb Mortal Wkly Rep* **65**(22): 576-584.  
Collier, S. A., et al. (2012). *Epidemiol Infect* **140**(11): 2003-2013.

- Incidence rate up from 0.4 to 1.6 cases per 100,000 people; exposure to potable water responsible for the majority of outbreak cases
- Between 2004-2007, LD cases had associated healthcare costs of \$9.4 million (Medicaid and Medicare patients)

Treated water - reclaimed (i.e. sewage treatment plant), ground, and surface

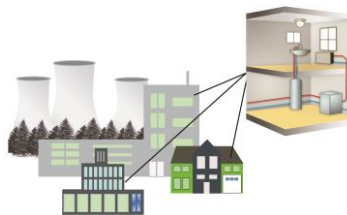


■  $3 \times 10^3 - 8 \times 10^4$  CFU L<sup>-1</sup>

■  $290 - 2.5 \times 10^3$  CE L<sup>-1</sup>

- |                          |                          |
|--------------------------|--------------------------|
| <i>L. anisa</i>          | <i>L. micdadei</i>       |
| <i>L. adelaidensis</i>   | <i>L. pneumophila</i>    |
| <i>L. bozemanii</i>      | <i>L. quateirensis</i>   |
| <i>L. donaldsonii</i>    | <i>L. sterigerwaltii</i> |
| <i>L. dumoffi</i>        | <i>L. wadsworthii</i>    |
| <i>L. fairfieldensis</i> | <i>L. waltersii</i>      |
| <i>L. fallonii</i>       | <i>L. worsleiensis</i>   |
| LLAP 2, 3, 4, 7, 8       |                          |
| <i>L. londiniensis</i>   |                          |

Distributed water - premise plumbing, e.g. residences, apartments, recreational facilities, hospitals, and cooling towers



■  $43 - 6 \times 10^5$  CFU L<sup>-1</sup>

■  $< 2 \times 10^3 - 8 \times 10^5$  GU L<sup>-1</sup>

- |  |                          |
|--|--------------------------|
| ■ $1.6 \times 10^5 - 2 \times 10^6$ CE L <sup>-1</sup> |                          |
| <i>L. anisa</i>  | <i>L. longbeacheae</i>   |
| <i>L. bozemanii</i>                                    | <i>L. lytica</i>         |
| <i>L. cherrii</i>                                      | <i>L. maceachernii</i>   |
| <i>L. dumoffi</i>                                      | <i>L. micdadei</i>       |
| <i>L. erythra</i>                                      | <i>L. pneumophila</i>    |
| <i>L. feelii</i>                                       | <i>L. rubrilucens</i>    |
| <i>L. gormanii</i>                                     | <i>L. saintcrucis</i>    |
| <i>L. gresilensis</i>                                  | <i>L. sterigerwaltii</i> |
| <i>L. hackeliae</i>                                    |                          |

Soil – garden, compost, potting



■  $10^2 - 10^8$  CFU g<sup>-1</sup>

■  $10^4 - 10^6$  GU g<sup>-1</sup>

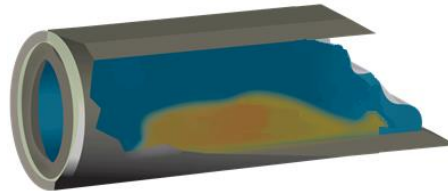
- |                           |                         |
|---------------------------|-------------------------|
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| <i>L. cincinnatiensis</i> | <i>L. oakridgensis</i>  |
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| <i>L. gormanii</i>        | <i>L. saintcrucis</i>   |
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Papadakis et al. 2018. Int J Environ Res Public Health  
 Schalk et al. 2014. Int J Infect Dis.  
 van Heijnsbergen et al. 2016. Appl Environ Microbiol.  
 van Heijnsbergen et al. 2014. J Appl Microbiol.

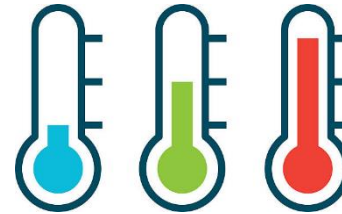
# Conditions within Premise Plumbing



generation of respirable, DW-derived aerosols



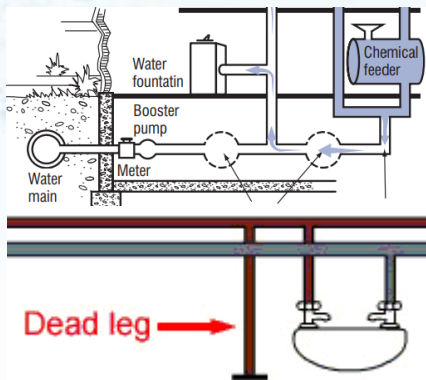
biofilms and sediments



amenable growth temperatures

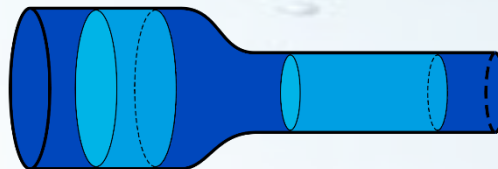


low to absent residual

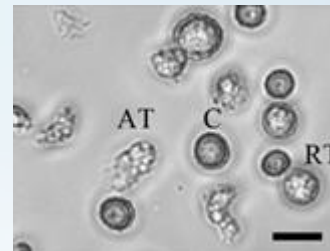


cross-connections

nutrients (metals, minerals, microbes, etc.)



high surface area to volume ratios



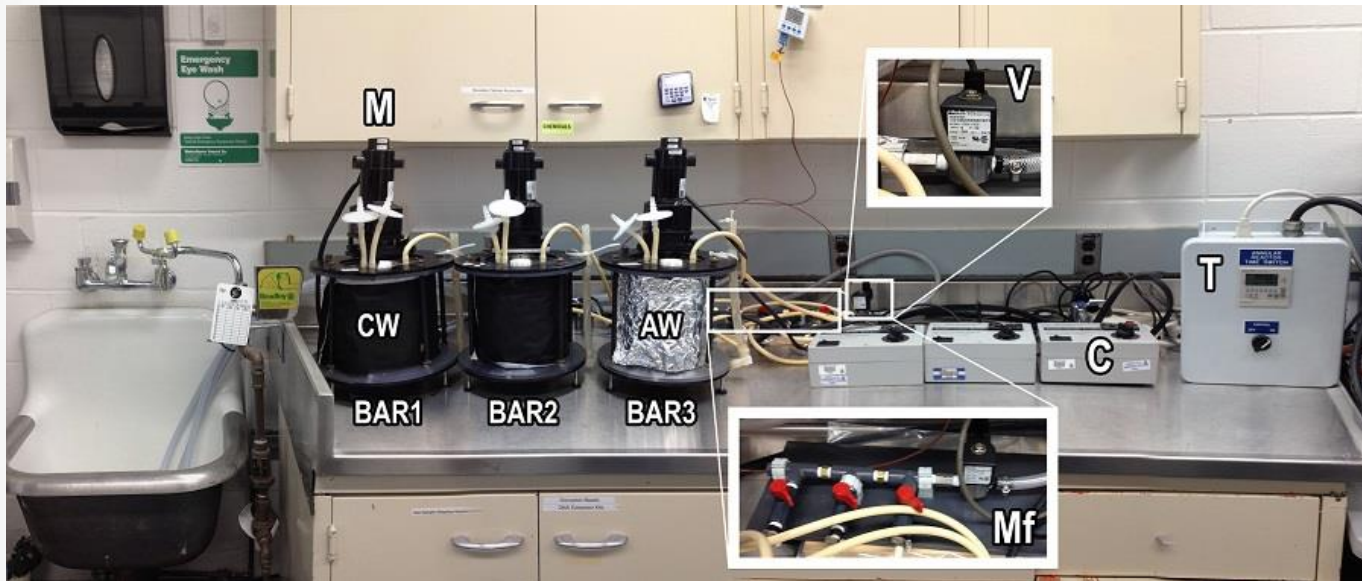
eukaryotic hosts



water stagnation



Project goal: determine the disinfection efficacy of chlorine and monochloramine, on DW biofilm-associated *L. pneumophila* established on copper (Cu) and polyvinylchloride (PVC) surfaces



BAR – biofilm annular reactor (1, 2, and 3)

M – motor

CW – cloth wrap

AW – aluminum wrap

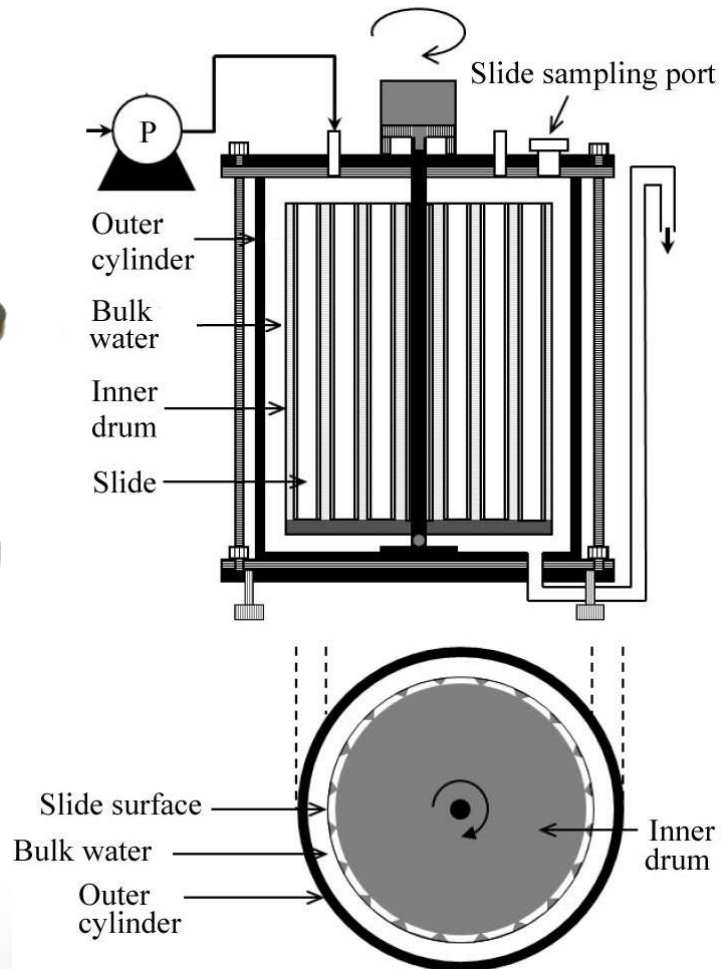
Mf – manifold

V – valve

C – controller

T – timer

- Developed mature (1.5-2 yo) drinking water biofilms on Cu and PVC surfaces
- Conditions set to mimic private residential water flow conditions
- Monitor water quality: TOC, HPC, chlorine decay, pH, temperature, hardness, turbidity, organics, inorganics, metals, etc.

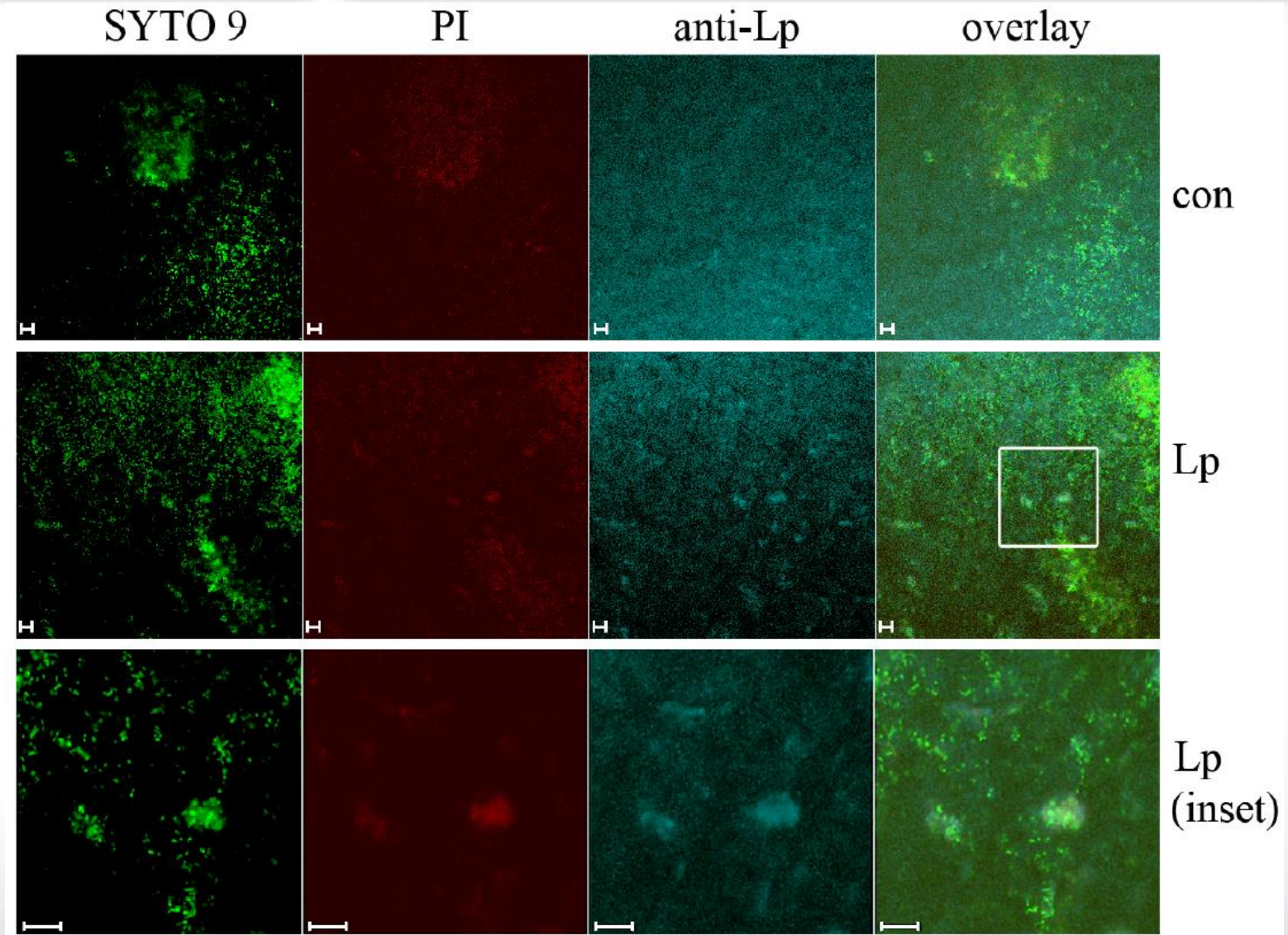




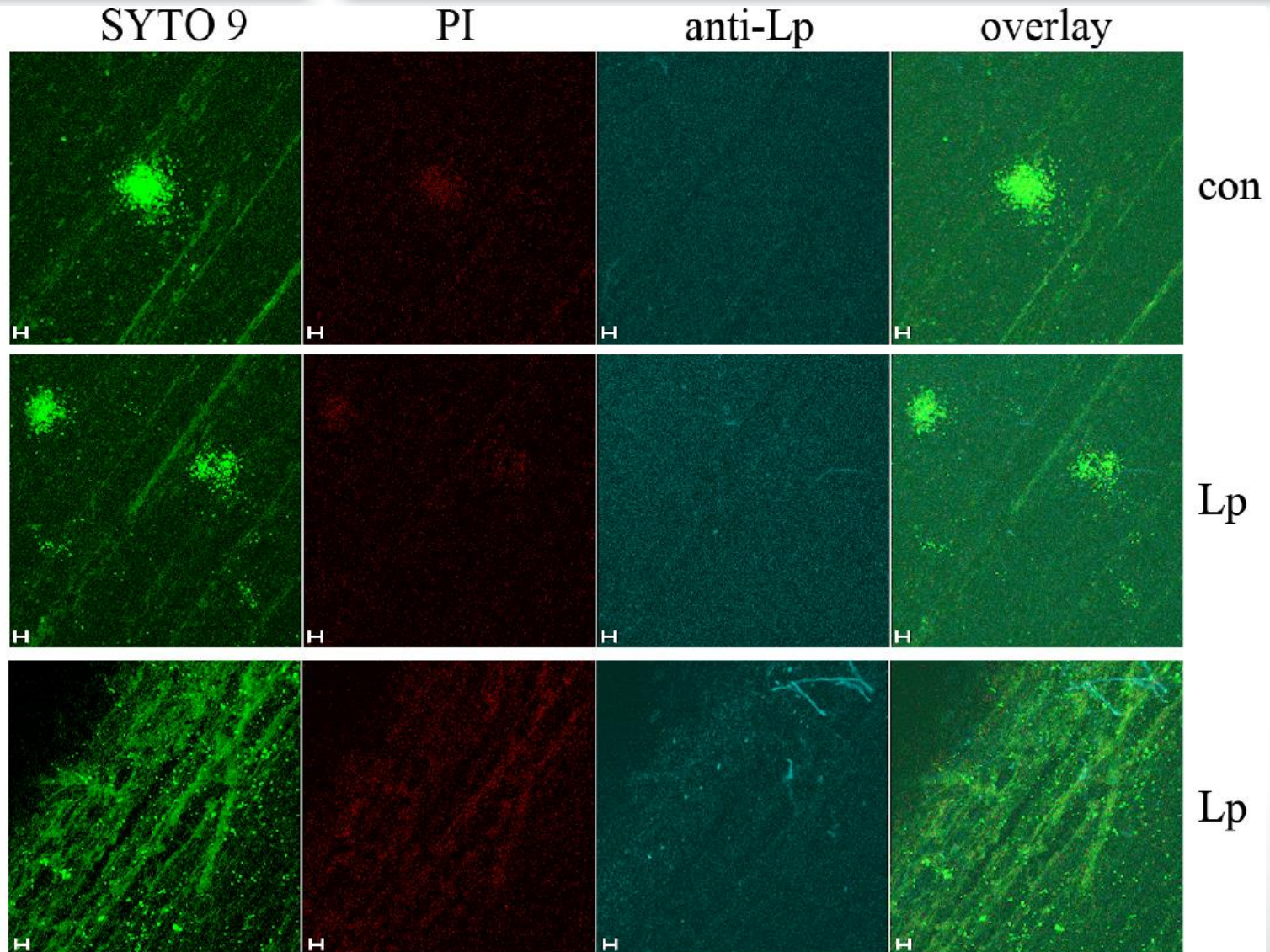
## *L. pneumophila* Inoculation Method



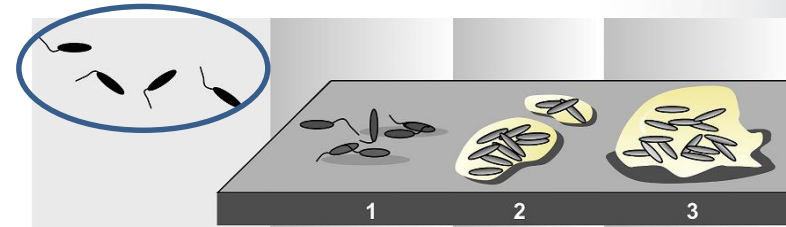
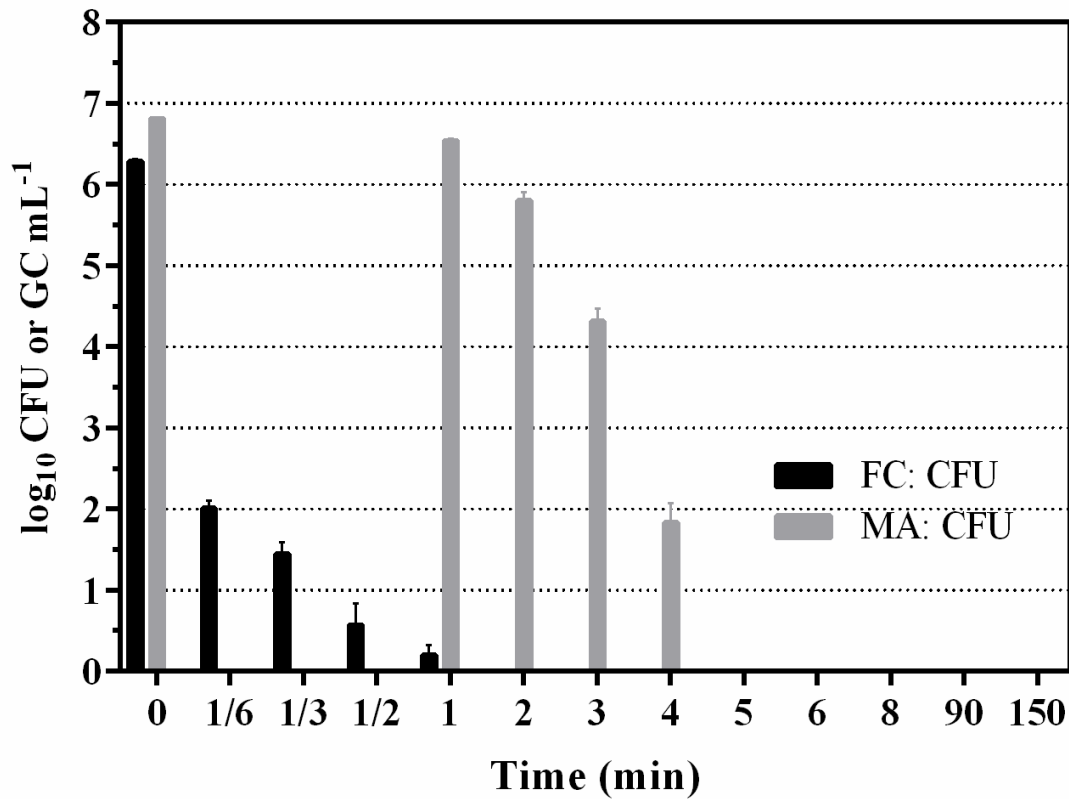
- slides removed from reactors
- suspended in beakers with  $10^6$  CFU mL<sup>-1</sup> Lp sg 1 in 0.22  $\mu$ m filtered, UV dechlorinated (5 d) drinking water (dfH<sub>2</sub>O)
- batch mode for 5 d
- slides rinsed twice in 250mL dfH<sub>2</sub>O
- incubated for additional 24 h in dfH<sub>2</sub>O







# Inactivation of Planktonic *L. pneumophila*



21°C, pH 8 conditions

Free chlorine (FC):  $2.1 \pm 0.1 \text{ mg L}^{-1}$

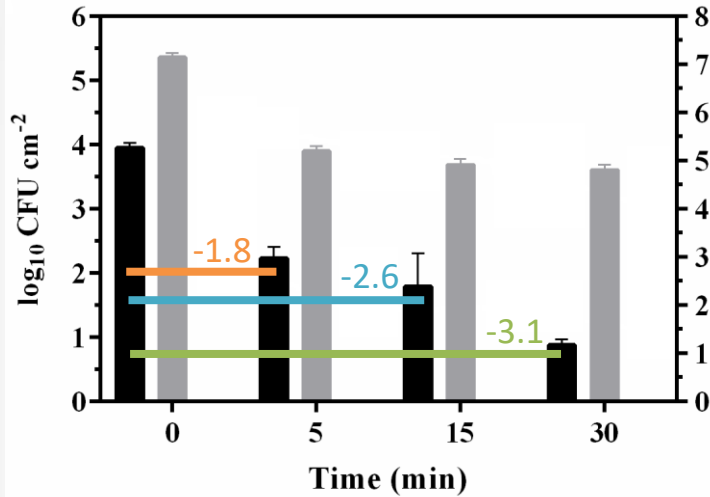
Monochloramine (MA):  $2.2 \pm 0.1 \text{ mg L}^{-1}$

Inactivation Ct value for:	FC	MA
2- $\log_{10}$	-	5.35
3- $\log_{10}$	0.11	6.58
4- $\log_{10}$	0.30	7.81

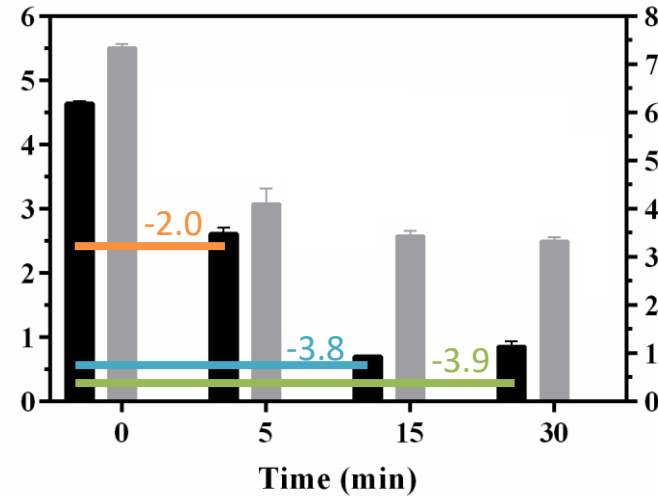


# Inactivation of Lp-Colonized Cu and PVC DW Biofilms

### Cu

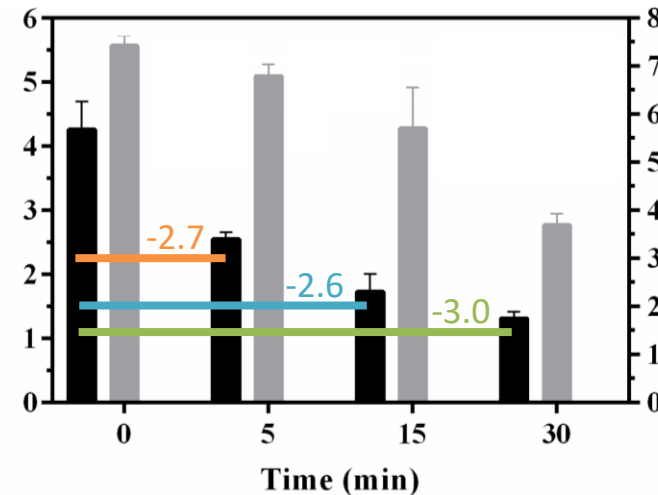
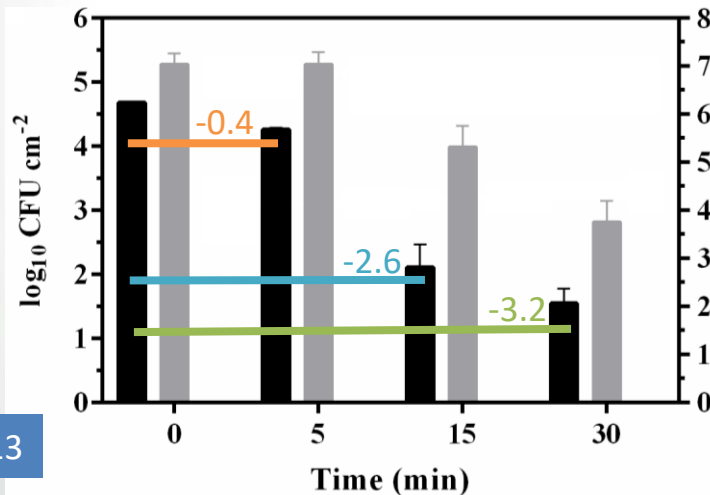


### PVC



Free chlorine

■ Lp CFU  
■ HPC CFU



Mono-chloramine



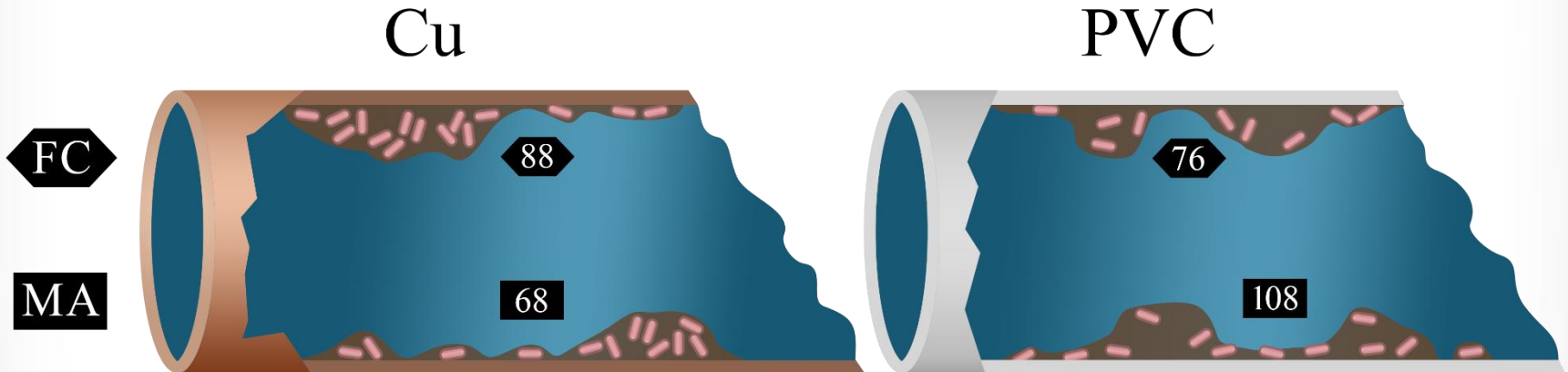


# Ct values for Planktonic- and Biofilm-Lp inactivation

Inactivation Ct value for:	Planktonic Lp		Drinking Water Biofilm-associated Lp			
	Free chlorine	Monochloramine	←Lower Ct		Higher Ct→	
2-log <sub>10</sub>	-	5.35	Free chlorine PVC 8.86	Free chlorine Cu 13.18	Monochloramine PVC 17.16	Monochloramine Cu 34.86
3-log <sub>10</sub>	0.11	6.58	Free chlorine PVC 36.11	Free chlorine Cu 50.83	Monochloramine Cu 55.38	Monochloramine PVC 62.8
4-log <sub>10</sub>	0.30	7.81	Free chlorine PVC 63.67	Monochloramine Cu 75.90	Free chlorine Cu 88.48	Monochloramine PVC 108.44

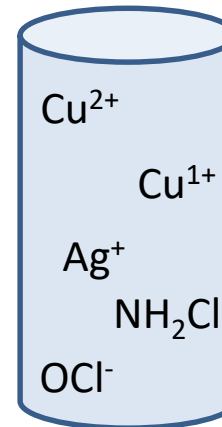
- 2- and 3-log reduction biofilm-Lp: free chlorine more effective at inactivation
- For 4-log reduction: Effectiveness seemed to be substratum and disinfectant dependent
  - Biofilm penetration? (Cu vs. PVC biofilm structures)
  - Chemical interactions between disinfectant and substratum?

- For planktonic *L. pneumophila*, free chlorine (FC) was more effective at inactivation compared to monochloramine (MA) treatment
- For biofilm-associated *L. pneumophila* and 4-log inactivation levels, MA was more effective on Cu biofilms, while FC was more effective on PVC biofilms



- Free chlorine treatment negatively impacted 16S rRNA gene transcript levels and may act synergistically with Cu surfaces to further reduce their levels (data not presented here)

Current study: determining the effectiveness of metal ions and various water quality parameters on *L. pneumophila* inactivation (bench scale and pilot scale studies)



- $\text{Cl}_2$  + Cu/Ag metal biocides effectiveness
- monitor water quality
- pathogen quantification (culture and molecular)
- microbial community analyses (bulk and biofilm phase)

Bench scale: evaluate combinations of microbes and inactivating agents

Evaluating decontamination technologies for drinking water distribution systems



## Ongoing & Planned Work

- **Characterization of contaminant spread via aerosolization and inhalation**
  - Assess efficiency of drinking water to air partitioning for *L. pneumophila*
  - Determine their viability and concentrations within the aerosols (and infectivity in murine models)

- **Mechanisms of Pathogen Mobility and Adherence to Infrastructure Surfaces**

NRMRL  
Darren Lytle, PhD  
Jill Holle  
Christy Muhlen

NRMRL ORISE  
Min Tang



Claressa Lucas, PhD  
Brian Raphael, PhD

Buse, Hoelle, Muhlen, Lytle 2018 FEMS ML

Understanding contaminant mobility (surface charge, hydrophobicity) and adhesion to pipe walls (microbalance) can help design effective decontamination strategies and/or engineering operations to minimize pathogen occurrence

➤ **Evaluating the effectiveness of flushing for controlling *Legionella pneumophila* growth in building water systems and reducing their levels in premise plumbing**

- Conduct longitudinal study on the impacts of flushing and thermal inactivation on *L. pneumophila*, *Pseudomonas aeruginosa*, and NTMs in hot water heaters, plumbing, and fixtures
- Study design: four hot water heaters, two of conventional design and two prototypes with passive control of microbial growth

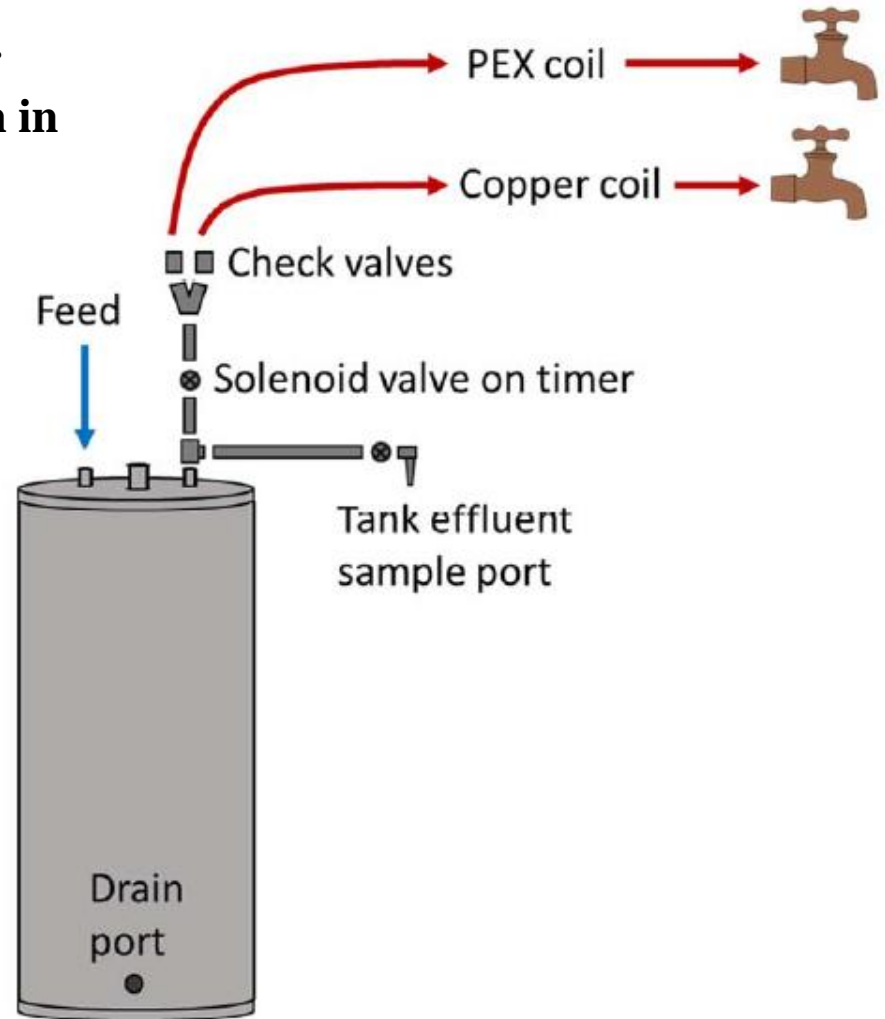


Figure 4. Hot Water Experiment Plumbing





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*L. pneumophila* sgl  
drinking water isolate  
grown on BCYE agar plates

