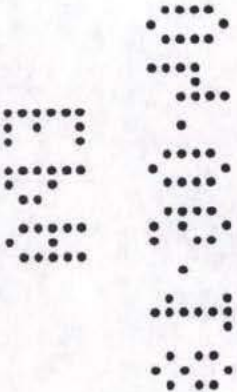


INTRODUCTION

The objective of this study was to independently validate the analytical method 11106.6116, for measuring residues of Ipconazole in soil and sediment of differing USDA Textural Classification in accordance with the EPA guideline OCSPP 850.6100 (2012): Environmental Chemistry Methods and Associated Independent Laboratory Validation.

Control samples of Calwich Abbey sediment and Speyer 5M soil were fortified with Ipconazole Technical (containing a mixture of Ipconazole cc and ct isomers) at concentrations of 50 and 500 µg/kg (total Ipconazole) in quintuplicate and analysed. Samples were extracted with acetonitrile: water: formic acid (90:10:0.1 v:v:v) followed by dilution into the calibration range with methanol: water (50:50 v:v). Control extracts from sediment and soil were used to prepare matrix matched standards, and were analysed against non-matrix standards to assess matrix effects. Samples were analysed using LC-MS/MS.

Matrix effects, linearity and specificity of the method were determined. Precision and accuracy was calculated at each validation level in each soil for total Ipconazole (primary and confirmatory).



MATERIALS AND METHODS

Test Substances

Test substance name: **Ipconazole Technical**
CAS number: 125225-28-7
IUPAC name: (1*RS*,2*SR*,5*RS*;1*RS*,2*SR*,5*SR*)-2-(4-chlorobenzyl)-5-isopropyl-1-(1*H*-1,2,4-triazol-1-ylmethyl)cyclopentanol
Molecular formula: C₁₈H₂₄ClN₃O
Sponsor lot number: 89010
Purity: 96.7% w/w (as total Ipconazole), 89.7% w/w (as Ipconazole cc), and 7.0% w/w (as Ipconazole ct)
Molecular mass: 333.9 g/mol
Storage conditions: Room Temperature (15-30°C)
Expiry date: 24 November 2019

Test substance name: **Ipconazole cc**
CAS number: 115850-69-6
Sponsor lot number: G-00328
Purity: 99.5% w/w
Storage conditions: Room Temperature (15-30°C)
Expiry date: 12 September 2021

Test substance name: **Ipconazole ct**
CAS number: 115937-89-8
Sponsor lot number: G-00329
Purity: 99.7% w/w
Storage conditions: Room Temperature (15-30°C)
Expiry date: 09 September 2021

Certificates of Analysis for the test substances are presented in Appendix 1.

Test System

Control samples of soil and sediment with differing USDA Textural Classification were sourced by Smithers Viscient (ESG). The soils used were Calwich Abbey sediment (silt loam) and Speyer 5M soil (sandy loam).

Soil characterisation data are listed in the table below:

Soil Name	Test System Code	Textural Class ¹	% Sand, Silt, Clay ²	CEC (meq/100 g)	% Organic Carbon	pH in H ₂ O	pH in 0.01M CaCl ₂
Calwich Abbey	CS 55/17	silt loam	29, 57, 14	17.4	4.7	7.7	7.3
Speyer 5M	CS 31/17	sandy loam	59, 30, 11	15.7	1.0	8.3	7.3

^{1,2}USDA classification.

The certificates of analysis for each soil are presented in Appendix 2.

Reagents

Acetonitrile	HPLC grade, Honeywell
Methanol	HPLC grade, Honeywell
Water	Milli-Q with LCPAK polisher, In House
Formic acid	ACS reagent, Honeywell
0.1% Formic acid in water	LC-MS grade, Honeywell
0.1% Formic acid in acetonitrile	LC-MS grade, Honeywell

Equivalent or better reagents may have been used.

Equipment

Shimadzu Nexera series HPLC system with AB Sciex API 5000 MS/MS detector.

Analytical Method

Analytical method 11106.6116 was supplied by the sponsor. The method used LC-MS/MS analysis.

Preparation of Reagents

Acetonitrile: water: formic acid (90:10:0.1 v/v/v) was prepared by mixing 1800 mL acetonitrile with 200 mL water and 2 mL formic acid.

Methanol: water (50:50 v/v) was prepared by mixing 500 mL methanol with 500 mL water.

Preparation of Stock Solutions

Primary stock solutions of Ipconazole Technical, Ipconazole cc and Ipconazole ct were prepared as described in the table below:

Stock ID	Test substance	Amount Weighed (mg)	Purity (%)	Solvent	Final Volume (mL)	Concentration (µg/mL) ¹
Stock 1	Ipconazole cc	10.91	99.5	Methanol	10.856	1000
Stock 2		10.38			10.328	1000
Stock 3	Ipconazole ct	10.18	99.7		10.150	1000
Stock 4		10.44			10.409	1000
Stock 5	Ipconazole	11.14	96.7 ²		10.773	1000
Stock 6	Technical	11.02			10.657	1000

¹Corrected for Purity.

²Purity is 96.7% w/w (as total Ipconazole), 89.7% w/w (as Ipconazole cc), and 7.0% w/w (as Ipconazole ct).

Primary stocks were stored refrigerated (2-8°C) in amber glass bottles and given a nominal three month expiry.

Secondary stock solutions of Ipconazole Technical, Ipconazole cc and Ipconazole ct were prepared as described in the table below:

Stock ID	Test substance	Stock Concentration (µg/mL)	Volume Taken (mL) ¹	Solvent	Final Volume (mL) ²	Concentration (µg/mL)
Stock 1	Ipconazole cc	1000	0.1	Methanol	10	10
Stock 2		1000	0.1		10	10
Stock 3	Ipconazole ct	1000	0.1		10	10
Stock 4		1000	0.1		10	10
Stock 5	Ipconazole	1000	1		10	100
Stock 6	Technical	1000	1		10	100

Secondary stocks were stored refrigerated (2-8°C) in amber glass bottles and given a nominal one month expiry.

Stock solutions of Ipconazole Technical for sample fortification and mixed stock solutions of Ipconazole cc and Ipconazole ct for calibration standard preparation were prepared as described in the table below:

Test Substance	Secondary Stock Concentration (µg/mL)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/mL)
Ipconazole cc	10	0.25	Methanol	50	0.05
Ipconazole ct	10	0.0175			0.0035
Ipconazole Technical	100	1		10	10
	10	1		10	1
	1	0.1		10	0.01

Stock solutions for sample fortification and calibration standard preparation were prepared on the day of use and stored refrigerated until the analysis was complete.

Preparation of Calibration Standards

Mixed calibration standards of Ipconazole cc and Ipconazole ct were prepared in methanol: water (50:50 v/v) as described in the table below:

Mixed Stock Concentration (µg/L) ¹	Volume Taken (mL)	Final Volume (mL)	Concentration (µg/L) ¹
50/3.5 ²	0.1	10	0.5/0.035
0.5/0.035	0.8	1	0.4/0.028
0.5/0.035	0.6	1	0.3/0.021
0.5/0.035	0.4	1	0.2/0.014
0.5/0.035	0.2	1	0.1/0.007
0.5/0.035	0.1	1	0.05/0.0035

¹ Concentrations expressed as Ipconazole cc/Ipconazole ct concentrations.

² 0.05/0.0035 µg/mL is equivalent to 50/3.5 µg/L.

Calibration standards were prepared on the day of analysis and discarded when analysis was complete. A single set of calibration standards was prepared for each

validation batch, which was analysed once before the samples and once after the samples. When samples required re-injection due to failure, the same calibration standards were used as the initial injection, so that the calibration standards and sample extracts were equally aged. Suitability of aged calibration standards was verified by an acceptable correlation coefficient.

Matrix Matched and Non-Matrix Matched Standards

In order to assess any possible matrix effect, matrix matched standards of Ipconazole Technical were prepared in control sample final extract for soil and sediment in triplicate. Non-matrix standards were prepared in methanol: water (50:50 v/v) in triplicate as described in the table below:

Fortification Stock Concentration (µg/mL)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
0.01	0.1	Methanol: water (1:1 v/v)	10	0.1
0.01	0.1		10	0.1
0.01	0.1		10	0.1
0.01	0.1	Calwich Abbey sediment final extract Control A	10	0.1
0.01	0.1		10	0.1
0.01	0.1		10	0.1
0.01	0.1	Speyer 5M soil final extract Control B	10	0.1
0.01	0.1		10	0.1
0.01	0.1		10	0.1

Sample Preparation and Fortification

The moisture content of the soil was determined and the weight of wet soil equivalent to 5 g dry weight was calculated. The required amount of wet soil (±0.005 g) was weighed into 50 mL centrifuge tubes. A single control soil and sediment was prepared for matrix assessment. Quintuplicate soil samples were fortified at the LOQ (50 µg/kg) and at 10 × LOQ (500 µg/kg) with Ipconazole Technical. Duplicate control soils and a reagent blank (without soil) were also prepared, as described in the following tables:

Recovery samples in silt loam sediment

Sample ID	Dry Weight (g)	Fortification Stock Concentration (µg/mL)	Volume Added (mL)	Fortified Concentration (µg/kg)
Reagent Blank A	N/A	N/A	N/A	N/A
Control A ¹	5	N/A	N/A	N/A
Control C-D	5	N/A	N/A	N/A
F50 A-E	5	1	0.25	50
F500 A-E	5	10	0.25	500

N/A = Not applicable.

¹ Control A was used for matrix assessment only.

Recovery samples in sandy loam soil

Sample ID	Dry Weight (g)	Fortification Stock Concentration (µg/mL)	Volume Added (mL)	Fortified Concentration (µg/kg)
Reagent Blank B	N/A	N/A	N/A	N/A
Control B ¹	5	N/A	N/A	N/A
Control E-F	5	N/A	N/A	N/A
F50 F-J	5	1	0.25	50
F500 F-J	5	10	0.25	500

N/A = Not applicable.

¹ Control B was used for matrix assessment only.

Soil Extraction

The samples were extracted twice with 20 mL acetonitrile: water: formic acid (90:10:0.1 v/v/v) by shaking at 150 rpm for 30 minutes and centrifuging at 3000 rpm for 10 minutes. The extracts were combined and made to 50 mL volume with acetonitrile: water: formic acid (90:10:0.1 v/v/v). Samples were then further diluted into calibration range with methanol: water (50:50 v:v). The extraction and dilution procedures are detailed in the table below:

Extraction and dilution in silt loam sediment

Sample ID	Nominal Soil Concentration (µg/kg)	Dry Weight (g)	Extract Volume (mL)	Extract Dilution (mL to mL)	Overall Dilution Factor	Nominal Extract Concentration after Dilution (µg/L)
Reagent Blank A	N/A	N/A ¹	50	0.2 to 10	500	N/A
Control A	N/A	5	50	1 to 50	500	N/A
Control C-D	N/A	5	50	0.2 to 10	500	N/A
F50 A-E	50	5	50	0.2 to 10	500	0.1
F500 A-E	500	5	50	0.075 to 10	1333	0.375

N/A = Not applicable.

¹ A dry weight of 5 g was used for calculation of the overall dilution factor in the Reagent Blank.

F500 A-E were re-diluted because an error was suspected with the original dilution.

Extraction and dilution in sandy loam soil

Sample ID	Nominal Soil Concentration (µg/kg)	Dry Weight (g)	Extract Volume (mL)	Extract Dilution (mL to mL)	Overall Dilution Factor	Nominal Extract Concentration after Dilution (µg/L)
Reagent Blank B	N/A	N/A ¹	50	0.2 to 10	500	N/A
Control B	N/A	5	50	1 to 50	500	N/A
Control E-F	N/A	5	50	0.2 to 10	500	N/A
F50 F-J	50	5	50	0.2 to 10	500	0.1
F500 F-J	500	5	50	0.075 to 10	1333	0.375

N/A = Not applicable.

¹ A dry weight of 5 g was used for calculation of the overall dilution factor in the Reagent Blank.

Instrument Conditions

LC-MS/MS analysis was performed using the following instrument conditions:

HPLC Parameters:

Column	XBridge C18 3.5 μ m 2.1 \times 50 mm		
Mobile Phase A	0.1% Formic acid in water		
Mobile Phase B	0.1% Formic acid in acetonitrile		
Flow Rate	0.4 mL/min		
Gradient	Time (min)	Mobile Phase A (%)	Mobile Phase B (%)
	0	50	50
	4	50	50
	7	30	70
	7.1	0	100
	9	0	100
	9.1	50	50
Run Time	10.5 minutes		
Column Temperature	40°C		
Autosampler Temperature	5°C		
Injection Volume	10 μ L		
Retention Time	Approx. 3.7 minutes (Iponazole ct) Approx. 3.9 minutes (Iponazole cc)		
Valco Valve Diverter	Time (min)	Position	
	0	A (to waste)	
	1	B (to MS)	
	9.5	A (to waste)	

MS/MS Parameters:

Instrument	AB Sciex API 5000 Triple Quadrupole Mass Spectrometer		
Ionisation Type	Electrospray (ESI)		
Polarity	Positive		
Scan Type	Multiple reaction monitoring (MRM)		
Ion Spray Voltage	4000		
Collision Gas (CAD)	5		
Curtain Gas (CUR)	25		
Gas Flow 1 (GS1)	40		
Gas Flow 2 (GS2)	40		
Vaporiser Temperature (TEM)	500		
Interface Heater (ihe)	On		
Entrance Potential (EP)	10		
Collision Exit Potential (CXP)	13		
Declustering Potential (DP)	100		
Compound Name	MRM Transition Ions Monitored	Collision Energy (CE)	Dwell Time (ms)
Iponazole cc/ct (Primary)	334 - 70	35	200
Iponazole cc/ct (Confirmatory)	336 - 70	42	200

LC-MS/MS data was collected using Analyst 1.6.2.

Calculation of Results

Results were calculated using Analyst 1.6.2. When the calibration fit is linear, Analyst uses the following formula to calculate the concentration of test substance present in the sample extract:

$$x = (y - c) / m$$

Where:

x = concentration of test substance in sample extract ($\mu\text{g/L}$)

y = peak area due to test substance

c = y intercept on calibration graph

m = gradient of the calibration graph

The concentration of test substance in the initial sample was calculated as follows:

$$\text{Sample concentration } (\mu\text{g/kg}) = \text{Extract concentration } (\mu\text{g/L}) \times \text{Dilution factor}$$

$$\text{Dilution factor} = \text{Final extract volume (mL)} / \text{soil weight in final extract (g)}$$

Procedural recovery from fortified samples was calculated as follows:

$$\text{Recovery (\%)} = \text{Sample concentration} / \text{Fortified concentration} \times 100$$

95% confidence intervals were calculated for each validation level as follows:

$$\text{95\% confidence interval } (\pm) =$$

$$1.96 \times \text{standard deviation of results} / \text{square root of the number of replicate results}$$

Grubbs test for outliers was calculated as follows:

$$G \text{ value} = (\text{suspect result} - \text{mean result}) / \text{standard deviation of results}$$

If the G value is greater than the critical value (1.715 for a sample size of 5) the result is an outlier with a significance of 0.05.

The limit of detection (LOD) based upon the sample concentration equivalent to three times the baseline noise of a control sample was calculated as follows:

$$\text{LOD} = 3 \times \text{height of control baseline noise} \times \text{control dilution factor} \times \text{calibration standard concentration } (\mu\text{g/L}) / \text{height of calibration standard peak}$$

The relative proportions of the cc and ct isomers in Ipconazole technical were calculated by dividing the isomer % purity by the total % purity stated on the Certificate of Analysis in Appendix 1.

Validation Pass Criteria

The validation was deemed acceptable if the following criteria were met for both the primary and confirmatory transitions monitored for total Ipconazole:

Mean Recovery and Precision – Recovery and precision were acceptable if each fortification level had a mean recovery between 70 and 120% and a % RSD (relative standard deviation) $\leq 20\%$.

Specificity/Selectivity – Specificity was acceptable if the amounts found in blank samples were $\leq 30\%$ of the limit of quantification (LOQ).

Linearity – Linearity was acceptable if the lowest calibration standard concentration was $\leq 20\%$ lower than the lowest sample nominal concentration and the highest calibration standard was $\geq 20\%$ higher than the highest nominal sample concentration (after dilution). If matrix effects were determined to be significant, matrix matched standards would be used. The correlation coefficient (r) was acceptable if it was ≥ 0.99 .

Limit of Detection (LOD) Assessment

An estimate of the LOD was made at $3 \times$ baseline noise of control soil and sediment for Ipconazole cc and Ipconazole ct (primary and confirmatory).

Method Detection Limit (MDL) Assessment

The MDL was calculated as the sample concentration equivalent to the lowest calibration standard (after dilution).

Matrix Assessment

An assessment of matrix effects was made by comparison of peak areas from control matrix final extracts fortified in triplicate with Ipconazole Technical against methanol: water (1:1 v/v) fortified in triplicate with Ipconazole Technical.

Results were presented as a % difference from the mean non-matrix standard value for Ipconazole cc and Ipconazole ct (primary and confirmatory).

A difference of $\geq 20\%$ was considered significant.

Variations to the Method

Small variations to the given method are listed as follows:

- Reagent supplier
- Equipment supplier
- Soil type
- Stock concentrations
- Scaling of reagents and solutions
- Optimisation of MRM transitions, MS voltages and gas pressures
- LC-MS software
- Use of measuring cylinder, rather than a volumetric flask to make extracts to volume
- Not re-centrifuging soil final extracts
- Using a different type of mass spectrometer
- Not using the same injector rinse solvents on the HPLC

These small variations demonstrated robustness of the method when transferred to an independent laboratory environment, and did not adversely affect the validation results.