Final Report

Study Title	Tebuconazole – Independent Laboratory Validation in Water
Study Guideline(s)	EPA 850.6100 (2012) SANCO/3029/99 rev 4 (2000)



INTRODUCTION

The objective of this study was to independently validate the analytical method 14162.6113, for measuring residues of Tebuconazole in surface and ground water by LC-MS/MS, in accordance with EPA 850.6100 (2012) and SANCO/3029/99 rev 4 (2000) guidelines.

Control samples of Fountains Abbey and Borehole water were fortified with Tebuconazole at 0.1 and 1 μ g/L in quintuplicate and analysed. Samples were diluted into calibration range with acetonitrile: water (20:80 v/v).

To assess matrix effects, calibration standards were prepared in acetonitrile: water (20:80 v/v) and in the final extract of untreated water.

Samples were analysed for Tebuconazole using liquid chromatography with tandem mass spectrometry detection (LC-MS/MS).

Matrix effects, linearity and specificity of the method were determined. Precision and accuracy was calculated at each validation level in Fountains Abbey and Borehole water. One primary and one confirmatory LC-MS/MS transition were analysed for Tebuconazole.

MATERIALS AND METHODS

Test Substance

Test substance name:	Tebuconazole
CAS Number:	107534-96-3
Molecular Formula:	$C_{16}H_{22}ClN_3O$
Structure:	
Molecular Mass:	307.82 g/mol
Purity:	98.2%
Batch Number:	201706010
Storage Conditions:	Room Temperature (15-30°C)
Recertification Date:	09 August 2020 ¹

¹ The test substance was manufactured in June 2017 and was certified on 03 July 2018 with an expiry date of June 2019. It was re-analysed on 09 August 2018 with a recertification date of 09 August 2020. The certified purity had not altered significantly between the two analysis dates, and was therefore considered to be stable for the duration of this study.

Test Matrix

Control samples of water were sourced by Smithers Viscient (ESG). The waters used were CS 14/18 Fountains Abbey (surface) and CS 13/18 Borehole (ground) water.

Water characterisation data are listed in the following table:

Water Name	Water Type	Suspended Solids (mg/L)	Conductivity (µS/cm)	Hardness (mg/L CaCO ₃)	рН	Dissolved Organic Carbon (mg/L)
Fountains Abbey	Surface	34	154	86	7.44	11.2
Borehole	Ground	2	436	349	8.0	0.00

Reagents

- Acetonitrile
- Water
- 0.1% Formic acid in water
- 0.1% Formic acid in acetonitrile

HPLC grade, VWR Milli-Q with LCPAK polisher, In House MS grade, Honeywell MS grade, Honeywell

Equipment

- Shimadzu Nexera series HPLC system with AB Sciex API 5000 MS/MS detector.
- HPLC column: Waters XBridge BEH C18, 2.5 μ m, 2.1 \times 50 mm
- Analytical balance
- Positive displacement pipettes
- Volumetric flasks
- Glass jars
- Amber glass vials
- Disposable glass vials
- HPLC vials

Analytical Method

Analytical method 14162.6113 was supplied by Smithers Viscient, Wareham on behalf of the sponsor. The method was re-written as SMV 3202240-01D to take into account minor differences in instrumentation, reagents and consumables before validation, and re-issued as SMV 3202240-01V after validation. The complete analytical procedure is presented in Appendix 6.

Preparation of Reagents

Acetonitrile: water (20:80 v/v) was prepared by mixing 100 mL HPLC grade acetonitrile with 400 mL Milli-Q water.

Preparation of Stock Solutions

Primary stock solutions of Tebuconazole in acetonitrile were prepared at 1000 μ g/mL under GLP Study No. 3202239 (Tebuconazole – Independent Laboratory Validation in Soil). These stocks were used for the matrix assessment, but not for the validation batch because they failed correlation (not within ± 5% difference from the mean peak area).

Primary stock solutions of Tebuconazole were prepared in volumetric flasks under this study, as described in the following table:

Stock ID	Amount Weighed (mg)	Purity (%)	Solvent	Final Volume (mL)	Concentration (µg/mL) ¹
Stock 1	10.60	08.2	A aatomitrila	10.410	1000
Stock 2	10.83	90.2	AcetoIIIthie	10.635	1000

¹Corrected for Purity.

Duplicate stocks were prepared for correlation purposes.

A secondary stock solution of Tebuconazole in acetonitrile was prepared at $10 \mu g/mL$ under study 3202239. This stock was used for the matrix assessment, but not for the validation batch because the primary stock failed correlation.

A secondary stock solution was prepared in a volumetric flask under this study, as described in the following table:

Stock Concentration	Volume Taken	Solvent	Final Volume	Concentration
(µg/mL)	(mL)		(mL)	(µg/mL)
1000	0.1	Acetonitrile	10	10

Secondary stock solutions were stored refrigerated in amber glass bottles and given a nominal expiry of one month.

Sub-stock solutions were prepared in volumetric flasks as described in the following table:

Stock Concentration	Volume Taken	Solvent	Final Volume	Concentration
(µg/mL)	(mL)		(mL)	(µg/mL)
10	0.01	Acatomitrila	10	0.01
0.01	1	Acetomume	10	0.001^{1}

¹Equivalent to 1 μ g/L.

Sub-stock solutions were prepared on the day of use and stored refrigerated until the corresponding analysis was complete.

Matrix Matched Standards for Matrix Assessment

Matrix matched standards of Tebuconazole were prepared in control water final extract as described in the following tables:

Fountains Abbey water

Stock Concentration	Volume Taken	Solvent	Final Volume	Concentration
(µg/L)	(mL)		(mL)	(µg/L)
1	0.05	Surface water final	5	0.01
1	0.05	Surface water final	5	0.01
1	0.05	Extract	5	0.01

Borehole water

Stock Concentration	Volume Taken	Solvent	Final Volume	Concentration
(µg/L)	(mL)		(mL)	(µg/L)
1	0.05	Cround water final	5	0.01
1	0.05	Ground water final	5	0.01
1	0.05	extract	5	0.01

Non-Matrix Matched Standards for Matrix Assessment

Non-matrix standards of Tebuconazole were prepared in acetonitrile: water (20:80 v/v) as described in the following table:

Stock Concentration (µg/L)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
1	0.05	A actomitrila, water	5	0.01
1	0.05	Acetomurile: water (20.90 w/w)	5	0.01
1	0.05	(20.80 V/V)	5	0.01

Preparation of Calibration Standards

Calibration standards of Tebuconazole were prepared in acetonitrile: water (20:80 v/v) as described in the following table:

Stock Concentration (µg/L)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
10	0.25		10	0.25
0.25	0.72		1	0.18
0.25	0.4		1	0.1
0.25	0.26	Acetonitrile: water (20:80 v/v)	1	0.065
0.25	0.18		1	0.045
0.25	0.12		1	0.03
0.25	0.08		1	0.02
0.25	0.05		1	0.0125
0.25	0.032		1	0.008
0.25	0.02]	1	0.005

A single set of calibration standards was prepared for each validation batch and injected twice, interspersed with and bracketing the samples.

Sample Fortification

5 mL of water was measured into a glass jar. Quintuplicate water samples were fortified at the LOQ ($0.1 \mu g/L$) and at $10 \times LOQ$ ($1 \mu g/L$) with a Tebuconazole standard in acetonitrile. Duplicate control water samples and a reagent blank (acetonitrile: water (20:80 v/v)) was also prepared, as described in the following tables:

Fountains Abbey water

Sample ID	Sample Volume (mL)	Stock Concentration (µg/L)	Volume Added (mL)	Fortified Concentration (µg/L)
Reagent Blank A ¹	5	N/A	N/A	N/A
Control A ²	5	N/A	N/A	N/A
Control C-D	5	N/A	N/A	N/A
F0.1 A-E	5	10	0.05	0.1
F1 A-E	5	10	0.5	1

N/A = Not applicable.

¹ Milli-Q water was used for the reagent blank.

² Control A was used for the matrix assessment.

Borehole water

Sample ID	Sample Volume (mL)	Stock Concentration (µg/L)	Volume Added (mL)	Fortified Concentration (µg/L)
Reagent Blank B ¹	5	N/A	N/A	N/A
Reagent Blank C ²	5	N/A	N/A	N/A
Control B ³	5	N/A	N/A	N/A
Control E-F	5	N/A	N/A	N/A
Control G-H ²	5	N/A	N/A	N/A
F0.1 F-J	5	10	0.05	0.1
$F0.1 \text{ K-O}^2$	5	10	0.05	0.1
F1 F-J	5	10	0.5	1

N/A = Not applicable.

¹Milli-Q water was used for the reagent blank.

² Reagent Blank C, Control G-H and F0.1 K-O were used for the second validation attempt at the LOQ, which was repeated due to unacceptably high levels of Tebuconazole in F0.1 G and H. As a precaution, the glass jars were rinsed with acetonitrile prior to use for the second attempt.

³Control B was used for the matrix assessment.

Sample Dilution

The water was made to 50 mL volume with acetonitrile: water (20:80 v/v). The diluted sample was transferred into an HPLC vial for analysis. Diluted samples were stored refrigerated in case further analysis was required. The dilution procedure is summarised in the following tables:

Sample ID	Fortified	Sample Volume	Final Volume	Dilution Factor
	Concentration	(mL)	(mL)	
	(µg/L)			
Reagent Blank A	N/A	5	50	10
Control A ¹	N/A	5	50	10
Control C-D	N/A	5	50	10
F0.1 A-E	0.1	5	50	10
F1 A-E	1	5	50	10

Fountains Abbey water

N/A = Not applicable.

¹ Control A was used for the matrix assessment.

Borehole water

Sample ID	Fortified	Sample Volume	Final Volume	Dilution Factor
	Concentration	(mL)	(mL)	
	(µg/L)			
Reagent Blank B	N/A	5	50	10
Reagent Blank C ¹	N/A	5	50	10
Control B ²	N/A	5	50	10
Control E-F	N/A	5	50	10
Control G-H ¹	N/A	5	50	10
F0.1 F-J	0.1	5	50	10
F0.1 K-O ¹	0.1	5	50	10
F1 F-J	1	5	50	10

N/A = Not applicable.

¹Reagent Blank C, Control G-H and F0.1 K-O were used for the second validation attempt at the LOQ.

²Control B was used for the matrix assessment.

Instrument Conditions

LC-MS/MS analysis was performed using the following instrument conditions:

HPLC Parameters:

Instrument:	Shimadzu Nexera series HPLC system				
Column#:	Waters XBridge BEH	C18, 2.5 µm, 2.1 × 50 µ	mm		
Mobile Phase A#:	0.1% Formic acid in w	ater			
Mobile Phase B#:	0.1% Formic acid in a	cetonitrile			
Flow Rate:	0.5 mL/min				
Gradient:	Time (min)	Mobile Phase A (%)	Mobile Phase B (%)		
	0.00	80	20		
	0.50	80	20		
	3.00	0	100		
	3.50	0	100		
	3.51	80	20		
	5.00	80	20		
Run Time:	5.0 minutes				
Column Temperature:	40°C				
Autosampler Temperature:	10°C				
Injection Volume:	50 µL				
Retention Time:	Approx. 2.5 minutes				
Valco Valve Diverter:	Time (min)		Position		
	0		A (to waste)		
	1		B (to MS)		
	4		A (to waste)		

MS/MS Parameters:

Instrument:	AB Sciex API 5000 Triple Quadrupole Mass Spectrometer			
Ionisation Type#:	Electrospray (ESI)			
Polarity#:	Positive			
Scan Type#:	Multiple reaction mo	onitoring (MRM)	
Ion Spray Voltage:	5000 V			
Collision Gas (CAD):	8			
Curtain Gas (CUR):	25			
Gas Flow 1 (GS1):	40			
Gas Flow 2 (GS2):	40			
Vaporiser Temperature (TEM):	500°C			
Interface Heater (ihe):	On			
Entrance Potential (EP):	10			
Collision Exit Potential (CXP):	13			
Transition Name:	MRM Transition	Declustering	Collision	Dwell Time (ms)
	Ions Monitored	Potential	Energy	
		(DP)	(CE)	
Tebuconazole (Primary):	308.4/70.0	120	36	75
Tebuconazole (Confirmatory):	308.4/125.0	120	54	75

Parameters marked # may not be modified. Minor adjustments to the remaining parameters may be required in order to fully optimise the system.

LC-MS/MS data were collected using Analyst 1.6.2.

Calculation of Results

LC-MS/MS data were calculated using Analyst 1.6.2. When the calibration fit is linear as in this study, Analyst uses the following formula to calculate the concentration of test substance present in the sample extract:

x = (y - c) / m

Where:

x = concentration of test substance in sample extract ($\mu g/L$)

y = peak area due to test substance

c = y intercept on calibration graph

m = gradient of the calibration graph

The calibration line used a 1/x weighting.

The concentration of test substance in the sample is calculated as follows:

Sample concentration ($\mu g/L$) = extract concentration ($\mu g/L$) × dilution factor

Dilution factor = final extract volume (mL) / volume of water in final extract (mL)

Procedural recovery from fortified samples is calculated as follows:

Recovery (%) = sample concentration / fortified concentration $\times 100$

The Limit of Detection (LOD) was measured from the peak height of the lowest calibration standard and the height of the baseline noise in a control sample and was calculated as follows:

LOD ($\mu g/L$) = 3 × height of control baseline noise × dilution factor × calibration standard concentration ($\mu g/L$) / peak height of calibration standard

The Method Detection Limit (MDL) based upon the sample concentration equivalent to the lowest calibration standard was calculated as follows:

MDL (μ g/L) = lowest calibration standard × dilution factor

For the correlation of stocks, each stock was diluted to a concentration near the middle of the calibration line. The diluted stocks were injected five times each, and the mean peak areas calculated. The correlation was then assessed using the following equation:

% Correlation from the mean =
$$\frac{(A - B)}{(A + B)} \times 100$$

Where A and B are the mean peak areas for each of the two stocks. The acceptance criterion is not more than \pm 5% difference from the overall mean.

Validation Pass Criteria

The validation was deemed acceptable if the following criteria were met for the primary and confirmatory transitions monitored:

Mean Recovery and Precision

Recovery and precision were acceptable if each fortification level had a mean recovery between 70 and 110% and a %RSD (relative standard deviation) $\leq 20\%$.

Specificity/Selectivity

Specificity was acceptable if no significant interference at the retention time of Tebuconazole was found in the control samples at > 30% of the LOQ.

Linearity

The linear range was acceptable if the lowest calibration standard concentration was $\leq 80\%$ of the equivalent LOQ final extract concentration and the highest calibration standard concentration was $\geq 120\%$ of the $10 \times LOQ$ extract concentration (after dilution if applicable). The correlation coefficient (r) was acceptable if it was ≥ 0.995 .

LOD (Limit of Detection) Assessment

An estimate of the LOD was made at $3 \times$ baseline noise for primary and confirmatory transitions.

MDL (Method Detection Limit)

The MDL was calculated as the sample concentration equivalent to the lowest calibration standard.

Matrix Assessment

An assessment of matrix effects was made by comparison of the peak areas for triplicate standards prepared in acetonitrile: water (20:80 v/v) and in untreated water final extracts. This was assessed for the primary and confirmatory transitions.

Results were presented as a % difference from the mean non-matrix matched standard value.

A difference of $\geq 20\%$ was considered significant.

The lowest calibration point was $\leq 80\%$ of the LOQ final extract concentration and the highest calibration point was $\geq 120\%$ of the $10 \times \text{LOQ}$ final extract concentration.

Limit of Quantification (LOQ)

The LOQ based upon the lowest level validated confirmed the LOQ to be 0.1 μ g/L for Tebuconazole in Fountains Abbey surface water and Borehole ground water.

Limit of Detection (LOD)

The LOD based upon the sample concentration equivalent to $3 \times$ baseline noise was calculated in Fountains Abbey and Borehole water (primary and confirmatory transitions). The LOD values are presented in the summary tables at the beginning of the results section.

Method Detection Limit (MDL)

The MDL for Tebuconazole was calculated to be 0.05 μ g/L, based upon a lowest standard concentration of 0.005 μ g/L and a control dilution factor of 10.

Matrix Effects

An assessment of matrix effects was made by comparison of peak areas from standards prepared in control water final extracts against standards prepared in acetonitrile: water (20:80 v/v). The difference from the mean non-matrix standard area was calculated.

Matrix effects were insignificant (< 20% difference from non-matrix standards) for Tebuconazole in Fountains Abbey and Borehole water, therefore non-matrix matched calibration standards were used for the method validation.

Validation Attempts

The first validation attempt for Fountains Abbey water was acceptable at both levels.

The first validation attempt for Borehole water failed at the 0.1 μ g/L level (LOQ) but was acceptable at the 1 μ g/L level. The validation was repeated at the LOQ using glass jars which had been rinsed with acetonitrile and passed.

Method Issues

There were no issues found with the primary method (14162.6113) or recommendations for improvement.

REFERENCES

EPA Ecological Effects Test Guideline OCSPP 850.6100: Environmental Chemistry Methods and Associated Independent Laboratory Validation (January 2012).

EC Guideline SANCO/3029/99 rev 4: Residues: Guidance for generating and reporting methods of analysis in support of pre-registration data requirements for Annex II (part A, Section 4) and Annex III (part A, Section 5) of Directive 91/414 (11 July 2000).

Smithers Viscient (Wareham) GLP Study No. 14162.6113: Environmental Chemistry Method: Validation of the Analytical Method for the Determination of Tebuconazole in Aqueous Matrices by LC-MS/MS.

Smithers Viscient (ESG) Ltd. GLP Study No. 3202239: Tebuconazole – Independent Laboratory Validation in Soil.

APPENDICES

Appendix 6 Analytical Procedure

Analytical Procedure

Procedure Title	Determination of Tebuconazole in Ground Water and Surface Water by LC-MS/MS
Procedure Code	SMV 3202240-01V
Issue Date	11 December 2018

1 of 12

Page Number

The methodology described in this procedure has been validated in Borehole Ground Water and Fountains Abbey Surface Water at 0.1 and 1 μ g/L.



REVISION HISTORY

SMV 3202240-01V New method issued following independent laboratory validation of Smithers Viscient, Wareham study 14162.6113

SAFETY PRECAUTIONS

Operators should take the normal precaution of wearing gloves, laboratory coats and safety glasses when handling compound and matrix samples.

Safety assessments (Control of Substances Hazardous to Health, COSIIII) have been made of those procedural steps involving preparation of solutions, reagents and analysis of matrix samples. Appropriate safety codes have been included in the text and are defined in the section titled General Handling Control Categories.

The hazards and risks of the substances hazardous to health used in this method have been considered. Provided the method is accurately followed and the control measures specified in the method are correctly used, there should be no foreseeable hazards to health.

INTRODUCTION

This method describes the procedure for determining concentrations of Tebuconazole in ground water and surface water by LC-MS/MS. Water is diluted into calibration range with acetonitrile: water (20:80 v/v). The extracts are quantified by LC-MS/MS.

Matrix effects for Tebuconazole in ground water and surface water were determined by comparing peak areas of calibration standards prepared in control water final extract and in acetonitrile: water (20:80 v/v).

Matrix effects are considered significant if the matrix matched standard area is $\geq 20\%$ different from the non-matrix standard area.

If matrix effects are significant then matrix matched calibration standards should be used.

- 2 -

HPLC grade, VWR

Milli-Q (with LCPAK polisher)

LC-MS grade, Honeywell

LC-MS grade, Honeywell

APPARATUS, MATERIALS, REAGENTS AND SOLUTIONS

Apparatus and Glassware

- Shimadzu Nexera series HPLC system with AB Sciex API 5000 MS/MS detector
- IIPLC column: Waters XBridge BEII C18, 2.5 µm, 2.1 × 50 mm
- Analytical balance
- Positive displacement pipettes
- Volumetrie flasks
- Glass jars
- Amber glass vials
- Disposable glass vials
- HPLC vials

Equivalent equipment may be used if required

Materials

- Acetonitrile
- Water
- 0.1% Formic acid in water
- 0.1% Formic acid in acetonitrile

Equivalent materials may be used if required

Reagents

Acetonitrile: water (20:80 v/v) Acetonitrile: water (20:80 v/v) is prepared by mixing 100 mL HPLC grade acetonitrile with 400 mL Milli-Q water.

Reagent volumes may be scaled as appropriate.

Standard Solution Preparation [1b, 4a] Primary Standard Stock

Prepare duplicate stock solutions of Tebuconazole at 1000 μ g/mL in acetonitrile. Accurately weigh ≥ 10 mg test substance, corrected for purity and transfer into a 10 mL volumetric flask. Adjust the volume to give exactly 1000 μ g/mL. Transfer into a mber glass bottles. The primary stocks should be stored refrigerated and given a nominal expiry date of 3 months.

Standard Correlation

Dilute the duplicate primary stocks to the mid-point of the calibration line. Correlate the standard solutions by injecting each of the two calibration standards 5 times into the LC-MS/MS. Ensure that the two solutions are injected alternately in the run sequence. The results for the correlation should be + 5% of the overall mean calculated by peak areas.

Review of Results

Review the data and document the correlation calculations. If the correlation is out of specification, either repeat the injections, re-dilute, or prepare two new stock

- 3 -

standards and repeat the procedures in sections << *Initial Weighing of Stock* Solutions>> to << *Review of Results*>>.

If the acceptance criteria from section <<*Standard Correlation*>> have been met, then the calibration solutions are acceptable for use. If required, fortification solutions for method validation will be made from the same stock standard, or its dilutions, from which the calibration line has been prepared.

Secondary Standard Stocks

Prepare secondary stock solutions of Tebuconazole in volumetric flasks as described in the following table:

Primary Stock Concentration	Volume Taken (mL)	Solvent	Final Volume	Secondary Stock Concentration
(μg/m L)			(mL)	(µg/mL)
1000	0.1	Acetonitrile	10	10

Transfer into amber glass bottles. The secondary stocks should be stored refrigerated and given a nominal expiry date of 1 month.

Sub-Stocks

Prepare sub-stock solutions of Tebuconazole in volumetric flasks as described in the following table:

Secondary Stock Concentration (µg/mL)	Volume Taken (mL)	Solvent	Final Volume (mL)	Sub-Stock Concentration (µg/mL)
10	0.01	Acatomitrila	10	0.011
0.01	1	Acetoniune	10	0.0012

¹ Equivalent to 10 µg/L.

² Equivalent to 1 μ g/L.

Transfer into disposable glass vials. The sub-stock solutions should be prepared on the day of use.

Matrix Matched Standards for Matrix Assessment

Prepare surface water and ground water matrix matched standards of Tebuconazole in disposable glass vials as described in the following table:

Stock Concentration (µg/L)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
1	0.05		5	0.01
1	0.05	Water final extract	5	0.01
1	0.05		5	0.01

- 4 -

Non-Matrix Matched Standards for Matrix Assessment Prepare non-matrix matched standards of Tebuconazole in disposable glass vials as described in the following table:

Stock Concentration (µg/L)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
1	0.05	A astanitsilar motor	5	0.01
1	0.05	(20.80 m/m)	5	0.01
1	0.05	(20.80 V/V)	5	0.01

Calibration Standards

Prepare calibration standards of Tebuconazole as described in the following table:

Stock Concentration (µg/L)	Volume Taken (mL)	Solvent	Final Volume (mL)	Concentration (µg/L)
10	0.25		10	0.25
0.25	0.72		1	0.18
0.25	0.4	Acetonitrile:	1	0.1
0.25	0.26		1	0.065
0.25	0.18		1	0.045
0.25	0.12	water (20:80 v/v) ¹	1	0.03
0.25	0.08		1	0.02
0.25	0.05		1	0.0125
0.25	0.032		1	0.008
0.25	0.02		1	0.005

If matrix matched calibration standards are required, use water final extract as the solvent.

A single set of calibration standards should be prepared for each validation batch and injected twice, interspersed with and bracketing the samples.

- 5 -

PROCEDURES

All procedures will be carried out in compliance with departmental SOPs, following departmental safety procedures in conjunction with COSHH assessments.

All work should be carried out under the minimum control categories listed under the safety precautions section. Additional controls are listed with the individual steps of the procedure.

Note: it is recommended that glass jars are rinsed with solvent before use.

Fortification of Control Samples for Method Validation [1b, 4a]

Measure 5 mL of either ground water or surface water into a glass jar. Fortify samples with Tebuconazole standard in acetonitrile as shown in the following table:

Γ	Number of	Sample Type	Stock	Volume	Sample	Fortified
	Replicates		Concentration	Added	Volume	Concentration
	-		(µg/L)	(mL)	(mL)	(µg/L)
	1	Reagent blank	N/A	N/A	N/A	N/A
Γ	2	Control	N/A	N/A	5	N/A
Γ	5	LOQ	10	0.05	5	0.1
Γ	5	$10 \times LOQ$	10	0.5	5	1

N/A = Not Applicable.

Sample Dilution [1b, 4a]

- 1. Measure 5 mL of water into a glass jar.
- 2. Dilute into calibration range with acetonitrile: water (20:80 v/v).
- 3. Transfer into an HPLC vial for analysis.

The recommended dilution procedure is given in the following table:

Sample type	Fortified Concentration (µg/L)	Sample Volume (mL)	Final Volume (mL)	Dilution Factor
Reagent blank	N/A	N/A	50	10
Control ¹	N/A	5	50	10
LOQ	0.1	5	50	10
$10 \times LOQ$	I	5	50	10

 $\rm N/A$ = Not Applicable. ¹ Use control extract for matrix matched calibration standards, if required.

- 6 -

LC-MS/MS CONDITIONS

HPLC Parameters:

Instrument:	Shimadzu Nexera series HPLC system			
Column#:	Waters XBridge BEH C18, 2.5 µm, 2.1 \sigma 50 mm			
Mobile Phase A#:	0.1% Formic acid in water			
Mobile Phase B#:	0.1% Formic acid in acetonitrile			
Flow Rate:	0.5 mL/min			
Gradient:	Time (min)	Mobile Phase A (%)	Mobile Phase B (%)	
	0.00	80	20	
	0.50	80	20	
	3.00	0	100	
	3.50	0	100	
	3.51	80	20	
	5.00	80	20	
Run Time:	5.0 minutes			
Column Temperature:	40°C			
Autosampler Temperature:	10°C			
Injection Volume:	50 µL			
Retention Time:	Approx. 2.5 minutes			
Valco Valve Diverter:	Time (min)		Position	
	0		A (to waste)	
	1		B (to MS)	
	4		A (to waste)	
MS/MS Parameters				

MS/MS Parameters:

Instrument:	AB Sciex API 5000	Triple Quadrup	ole Mass Sp	ectrometer
Ionisation Type#:	Electrospray (ESI)			
Polarity#:	Positive			
Scan Type#:	Multiple reaction monitoring (MRM)			
Ion Spray Voltage:	5000 V			
Collision Gas (CAD):	8			
Curtain Gas (CUR):	25			
Gas Flow 1 (GS1):	40			
Gas Flow 2 (GS2):	40			
Vaporiser Temperature (TEM):	500°C			
Interface Heater (ihe):	On			
Entrance Potential (EP):	10			
Collision Exit Potential (CXP):	13			
Transition Name:	MRM Transition	Declustering	Collision	Dwell Time (ms)
	Ions Monitored	Potential	Energy	
		(DP)	(CE)	
Tebuconazole (Primary):	308.4/70.0	120	36	75
Tebuconazole (Confirmatory):	308.4/125.0	120	54	75

Parameters marked # may not be modified. Minor adjustments to the remaining parameters may be required in order to fully optimise the system.

- 7 -

CALCULATION OF RESULTS

All peak measurements and calculations are performed on Analyst 1.6.2. From the measured peak area, where the calibration fit is linear as in this study, Analyst uses the following formula to calculate the concentration of test substance present in the sample extract.

$$x = \frac{(y-c)}{m} \times DF$$

Where:-

x = concentration of test substance in sample (μ g/L)

y = area of peak due to test substance

m – gradient

 $c = \mathbf{\tilde{Y}}$ intercept on calibration graph

DF = sample dilution factor

Procedural recovery data from fortified samples are calculated via the following equation:

Recovery(%) =
$$\frac{A}{S} \times 100$$

Where:-

A = concentration found in fortified sample (μ g/L)

S = concentration added to fortified sample ($\mu g/L$)

- 8 -

METHOD CRITERIA

For the analysis by LC-MS/MS to be considered successful the following criteria should be met.

- At least 5 calibration standards will be used in the determination of the calibration line.
- The correlation coefficient (r) for the calibration line will be ≥ 0.995 with a 1/x weighting.
- All sample extracts will be within the appropriate range of calibration standards.
- Mean recovery from fortified samples will be considered acceptable within the range of 70 to 110%.
- The control sample should not contain interference > 30% of the LOQ.

-9-

CATEGORY		CONTROL
Main Division		Name and Specification
1		GLOVES
	a	Disposable latex
	b	Disposable nitrile
	¢	Rubber gloves
	d	Specific type for the job (see assessment giving details)
2		PROTECTIVE CLOTHING
	a	Laboratory coat or equivalent
	b	Disposable overalls
	с	Oversleeves
	d	Overshoes
	e	Plastic apron
3		EYE/FACE PROTECTION
	a	Safety glasses to BS 2092/2 C or better
	b	Face shield to BS 2092/2 C or better
	¢	Safety goggles to BS 2092/2 C or better
4		ENGINEERING CONTROLS
	a	Open bench in ventilated area
	b	Fume cupboard to BS 7258
	c	Laminar flow cabinet to BS 5295 Class 1
	d	Re-circulating fume chamber
	e	Radioisotope lab
	f	Biohazard lab
	g	Glove box
5		RESPIRATORY PROTECTIVE EQUIPMENT
	a	Disposable filtering facemask (HSE approved),
		1 - organic vapour
		11 - dust
		iii – combination organic vapour/dust
	1-	MUST SPECIFY TYPE
	b	Powered respirators/neimets with safety visor to BS 2092/2 C
	_	or better (HSE approved)
(¢	Respirator with specified canister (HSE approved)
7		ALLEDCIC DEBSONG DECHDITED (CIVE DETAILS)
/		ALLERGIC PERSONS PROHIBITED (SPECIFY ALLERGY)
8		KEFEK TO MATERIAL SAFETY DATA SHEET
9		KNOWN OR SUSPECTED REPRODUCTIVE HAZARD TO
10		FITTER SEA (must specify details)
10		POISON ensure antidote is available and is within its expiry date (must specify details)

GENERAL HANDLING CONTROL CATEGORIES

- 10 -