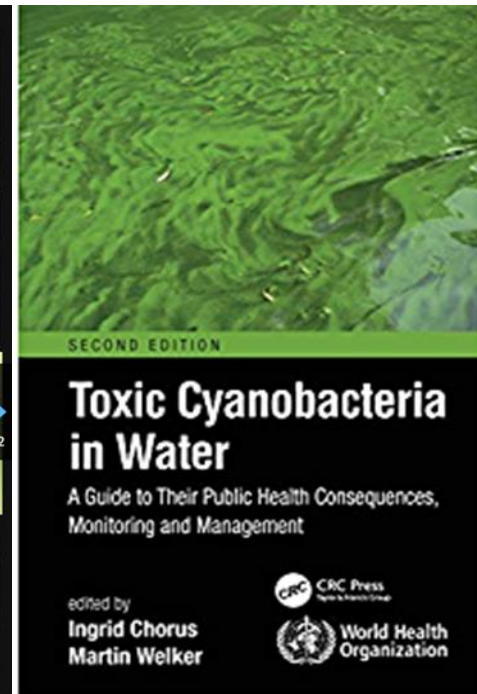


Cyanotoxins in freshwater fish and shellfish **understanding** risk assessment and management



WHO TCiW Book launch 2021



<https://doi.org/10.1201/9781003081449>

Understanding the hazard

Chapter 2: Introduction to cyanotoxins

Chapter 3: Introduction to cyanobacteria

Chapter 5:

Preventing hazardous exposure to cyanotoxins

5.1: Drinking-water

5.2: Recreation and workplace

5.3: Food

5.4: Renal dialysis

5.5: Dietary supplements

Controlling cyanobacteria and cyanotoxin occurrence

Chapter 6: Assessing & managing risks

Chapter 7
Catchment

Chapter 8
Waterbody

Chapter 9
Sites of use

Chapter 10
Water treatment

Monitoring cyanobacteria and cyanotoxins

Chapter 11: Planning monitoring programmes

Chapter 12: Site inspection and sampling

Chapter 13: Analysing cyanobacteria and water chemistry

Chapter 14: Analysing cyanotoxins

Chapter 15: Public surveillance, communication and participation

Introduction and role of public authorities (chapters 1 and 15)

... Cyanotoxins are **among the most toxic** naturally occurring compounds...

... **widespread occurrence**; contact may be difficult to avoid

... **less public awareness** of risk than for other chemicals

... **control indirect and complex**– through controlling eutrophication: successful approaches need to be site-specific

Introduction and role of public authorities (chapters 1 and 15)

Context of toxic Cyanobacteria – **similar** to other chemicals in water:

... lack of quantitative data (dose estimates) for human exposure...

...the contribution of **toxic chemicals in water to morbidity and mortality is rarely acute**, ... impacts on health are less visible and less clearly attributable to chemicals.

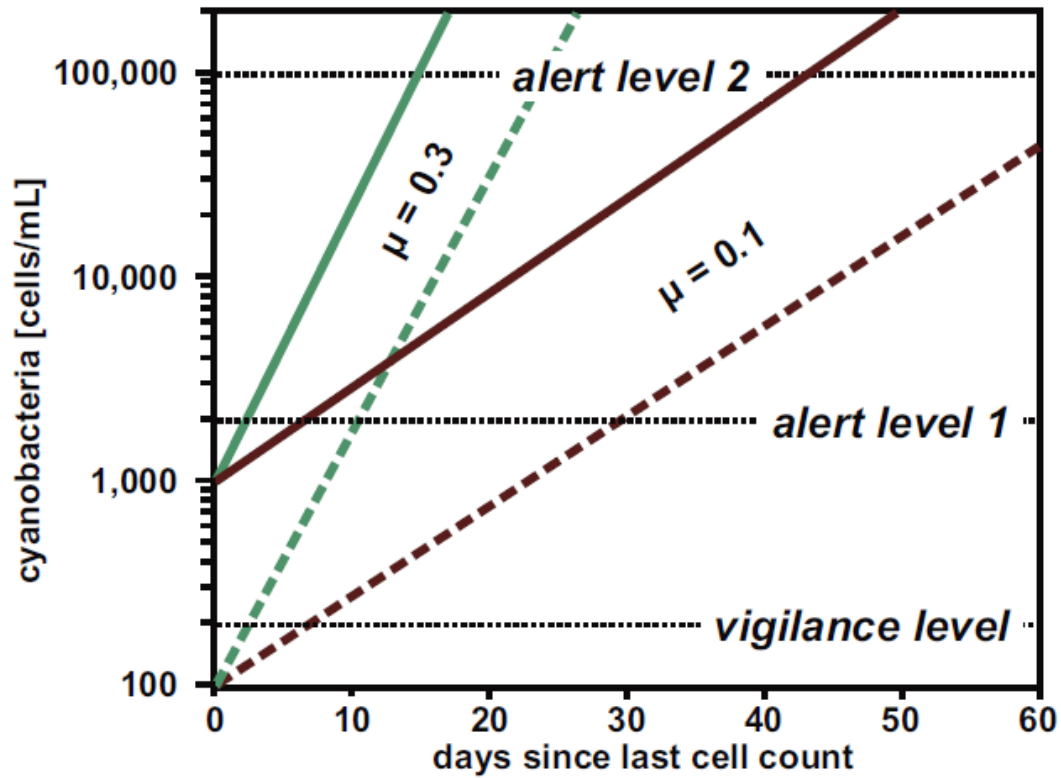
... unclear whether cyanotoxins or other hazards were the cause of symptoms reported

... **quite clear data from animal experiments**

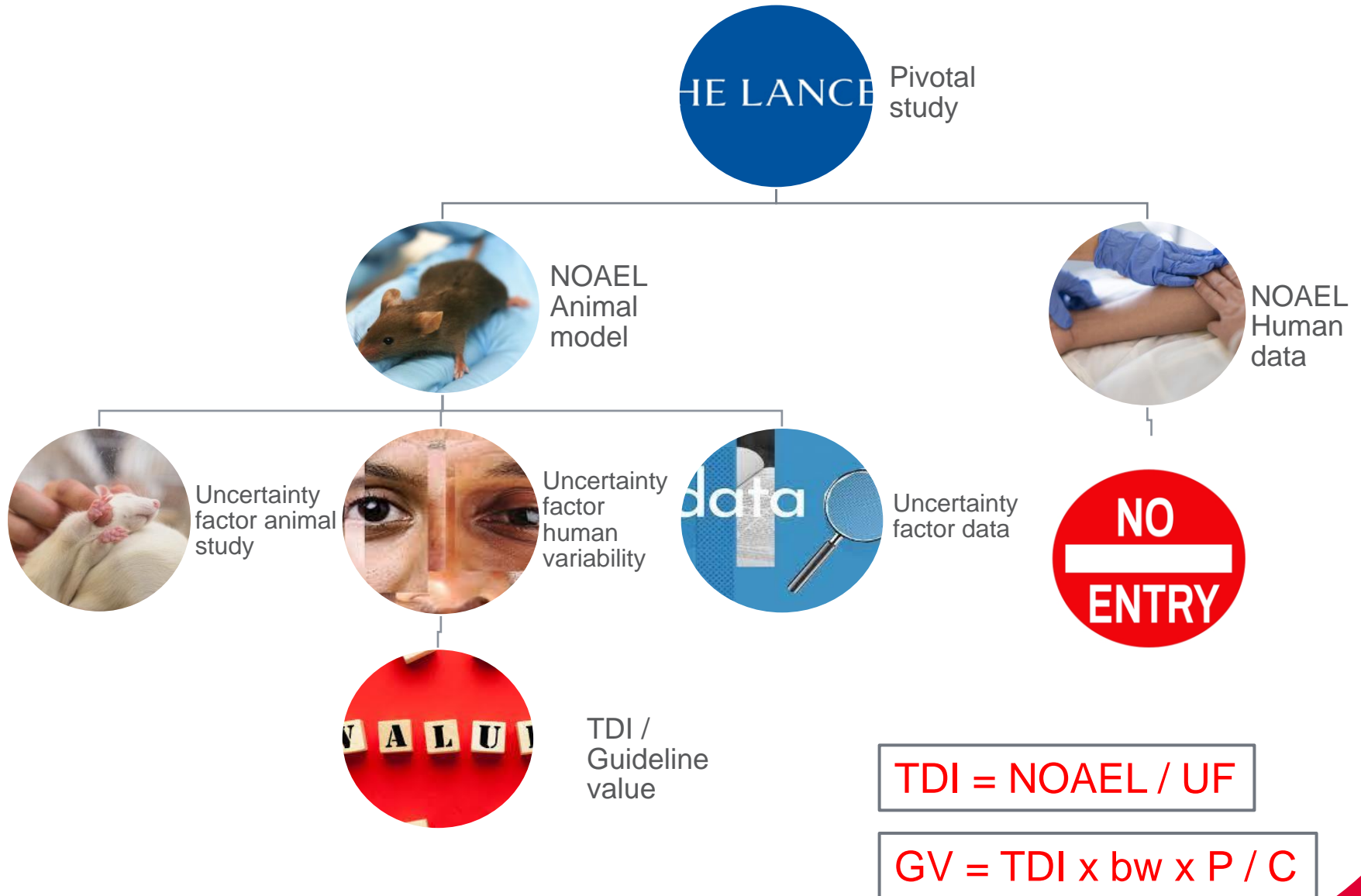
Like for other chemicals in water:

WHO guideline value not because of widespread cause of illness – but because of significant toxicity and water as relevant exposure pathway





Derivation GV and TDI



Analysis of Epidemiological data and Human Reports

Can individual cases support the derivation of GV?

- “... mere **co-occurrence of cyanotoxins and unspecific symptoms** (skin irritation, gastro-intestinal, etc. ...) is not indicative of the known cyanotoxins having caused the symptoms.....
- Human effects ascribed to the presence of cyanotoxins in the various vectors (drinking- or recreational water, food) such as gastrointestinal illness or skin irritation may well be due to other unknown cyanobacterial metabolites or closely linked to **pathogens or other substances associated with the bloom**
- **Epidemiological studies: indicative value only** because (i) uncertain, retrospective(!) exposure estimates; (ii) other contaminants in surface water; (iii) demographic info is limited
- Important to understand **cause–effect relationships** and then **dose-response relationship** (for the quantitative aspects – then role in GV)



The toxicity of cyanobacterial toxins in the mouse: I Microcystin-LR

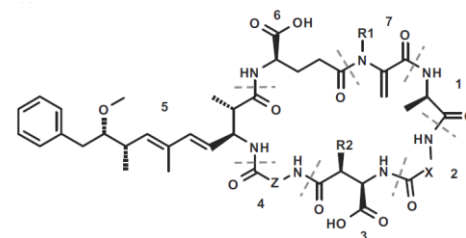
JK Fawell^{*1}, RE Mitchell¹, DJ Everett² and RE Hill²

¹WRC National Centre for Environmental Toxicology, Medmenham, Bucks, SL7 2HD, England; ²Quintiles Toxicology/Pathology Services, Bromyard Road, Ledbury, Herefordshire, HR8 1LH, England

Table 3 Thirteen week study – liver microscopic pathology

Dose level ($\mu\text{g kg}^{-1}$ bodyweight per day)	Control (0)	NOAEL 40	LOAEL 200	1000
Males				
Number examined	15	15	15	15
Acute inflammation	0	1	0	0
Chronic inflammation	1	2	4	15
Congestion	3	0	0	1
Hepatocyte vacuolation	5	5	6	3
Haemosiderin deposits	0	0	0	15
Hepatocyte degeneration	0	0	1	14
Females				
Number examined	15	15	15	15
Autolysis	0	0	0	1
Chronic inflammation	5	8	8	14
Congestion	0	0	0	1
Hepatocyte vacuolation	5	5	11	8
Haemosiderin deposits	0	0	1	14
Hepatocyte degeneration	0	0	1	9

Fawell et al. (1999): Mice of both sexes given MC-LR by gavage at 40 $\mu\text{g/kg}$ bw per day for 13 weeks did not show treatment-related effects in the parameters measured. Only slight hepatic damage was observed at the lowest observed effect level (LOAEL) of 200 $\mu\text{g/kg}$ bw per day in a limited number of treated animals, whereas at the highest dose tested (1 mg/kg bw per day), all the animals showed hepatic lesions, consistent with the known action of MC-LR.



Derivation provisional **TDI** MC-LR

$$\text{TDI}_{\text{chronic}} = \frac{\text{NOAEL}}{\text{UF}} = \frac{40}{1000} \frac{\mu\text{g}}{\text{kg}} / \text{d} = 0.04 \frac{\mu\text{g}}{\text{kg}} / \text{d}$$

$\text{TDI}_{\text{MC,chronic}}$ = tolerable daily intake for chronic exposure

NOAEL = no-observed-adverse-effect level (40 $\mu\text{g}/\text{kg}$ bw per day, based on Fawell et al., 1999)

UF = uncertainty factor (1000 = 10 for interspecies variation \times 10 for intraspecies variation \times 10 for database deficiencies, including use of a subchronic study)

PROVISIONAL

Derivation provisional **life term drinking water GV** MC-LR

$$GV_{\text{chronic}} = \frac{\text{NOAEL} * \text{bw} * P}{\text{UF} * C} = \frac{40 * 60 * 0.8}{1000 * 2} \mu\text{g} / \text{L} = 0.96 \mu\text{g} / \text{L} \approx 1 \mu\text{g} / \text{L}$$

GV_{chronic} = guideline value for chronic (lifetime) exposure

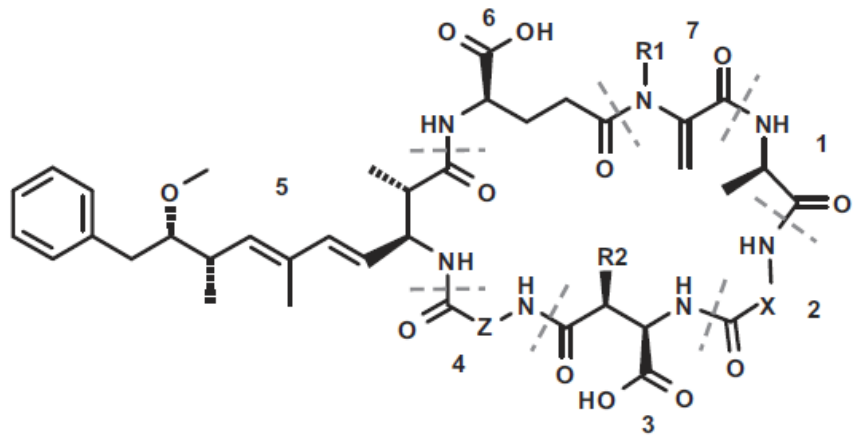
NOAEL = no-observed-adverse-effect level (40 $\mu\text{g}/\text{kg}$ bw per day, based on Fawell et al., 1999)

bw = body weight (default = 60 kg for an adult)

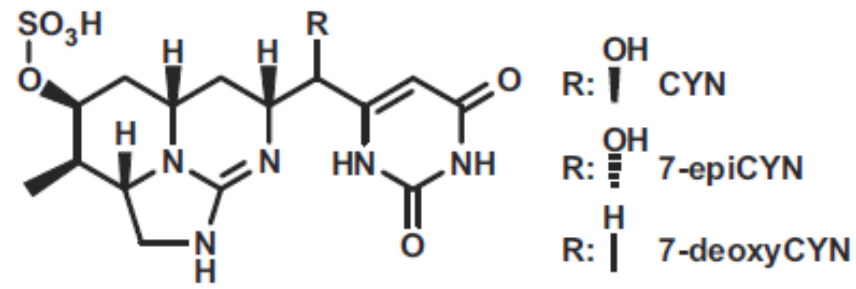
P = fraction of exposure allocated to drinking-water (80%, as other sources of exposure such as air, food and soil are considered minor for lifetime exposure)

UF = uncertainty factor (1000 = 10 for interspecies variation \times 10 for intraspecies variation \times 10 for database deficiencies, including use of a subchronic study)

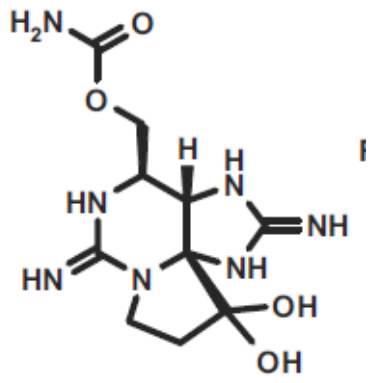
C = daily drinking-water consumption (default = 2 L for an adult).



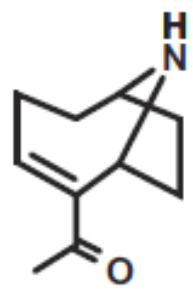
Microcystin



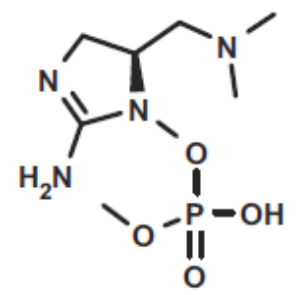
Cylindrospermopsin



Saxitoxin



Anatoxin-a



Anatoxin-a (s)

Cyanobacterial toxins

Hepatotoxic cyclic peptides: microcystins and nodularins

- Characteristic Adda amino acid
- > 250 variants for MCYST
- Nomenclature based variable amino acids positions 2 and 4
- Water soluble
- OATP membrane transporter to enter cell (liver)
- Slow detoxication liver
- Inhibition protein phosphatases, destabilization cytoskeleton
- Tumour promoting
- Producers MCYST across all cyanobacteria, main genera *Microcystis*, *Planktothrix*, *Dolichospermum*, *Nostoc*
- *Nodularia* for NODLN
- Produced via nonribosomal peptide synthetase and polyketide synthase pathway
- Cell quota dependent on temperature, light, pH, N, P, Fe, salinity
- Positive correlation growth rate and MC content
- Tens of $\mu\text{g} / \text{L}$ in lakes, up to > 100 mg / L in scums
- Dynamics linked to strain variation

Cyanobacterial toxins

Cylindrospermopsin

- **Alkaloid**
 - 4 structural variants
 - Hydrophobic
 - Various modes action, low dose inhibition protein synthesis, ROS
 - Genotoxic
 - Unclear whether CYN production constitutive or not
 - Species from Nostocales, Oscillatoriales, genera *Raphidopsis* (*Cylindrospermopsis*), *Chrysochloris*, *Aphanizomenon*, *Anabaena*, *Umezakia*
 - *Planktonic and benthic*
 - *Per unit biovolume 0.6 – 3.5 µg / mm³*
- In lake water 10 – 20 µg / L
 - Substantial share (up to 90 %) CYN found **extracellularly** (in contrast MC)
 - Environmental conditions change cell quota, ratio between variants and intra vs extracellular

Cyanobacterial toxins

Anatoxin-a

- Secondary amine **alkaloids**
- Anatoxin-a, homoanatoxin, dihydroanatoxin-a
- Pre- and postsynaptic **depolarizing agent in nerve cells**, increased simulation
- Brain and cardiovascular system target organs
- Muscular paralysis and respiratory failure
- **Inadequate data GV**, bounding value = **provisional health-based reference value**, NOAEL 98 µg / kg bw
- Producers *Dolichospermum*, *Anabaena*, *Oscillatoria*, *Planktothrix*, *Raphidopsis* ...
- Environmental factors like light and temperature 2-7 fold variation ATX
- Like MCYST **worldwide distribution**
- USA concentrations in literature 35 – 1170 µg / L
- Important production benthic cyanobacteria like *Phormidium*

Cyanobacterial toxins

Saxitoxins

- Shellfish poisoning toxins (PSP)
 - 57 analogues
 - Mainly hydrophylic
 - Na, Ca and K channel blocking in neural and cardiac cells, prevents electrical transmission
 - Twitching, convulsions, muscle and respiratory paralysis
 - LOAEL based upon review human cases PSP: STX = 1.5 $\mu\text{g}/\text{kg}$ bw, UF of 3, NOAEL estimated = 0.5 $\mu\text{g}/\text{kg}$ bw (acute reference dose)
 - Allocation factor drinking water vs food, risk co-incident exposure
 - Produced Nostocales, Oscillatoriales, including..
- *Dolichospermum*, *Aphanizomenon*, *Raphidiopsis*, *Planktothrix*, *Microseira* (*Lyngbya*)
 - High cellular content *Dolichospermum circinale* (Murray-Darling 1000 km bloom)

Chapters 2.5 – 2.10 – Other toxins and metabolites

E. TESTAI, N. J. OSBORNE, N. CHERNOFF, E.J. FAASEN, D.J. HILL, M. WELKER, T. KALOUDIS, A. HUMPAGE

- 2.5 Anatoxin-a(S) – no GV
- 2.6 Marine dermatoxins – no GV (skin exposure)
- 2.7 β -Methylamino-L-alanine (BMAA)
 - No convincing evidence for the role of BMAA as cause of neurodegenerative disease under realistic exposure scenarios
- 2.8 Cyanobacterial lipopolysaccharides
 - "...cyanobacterial LPS are not likely to pose health risks to an extent known from toxins like microcystins or cylindrospermopsins, in particular, when considering plausible exposure pathways."

Chapters 2.5 - 2.10 – Other toxins and metabolites

E. TESTAI, N. J. OSBORNE, N. CHERNOFF, E.J. FAASEN, D.J. HILL, M. WELKER, T. KALOUDIS, A. HUMPAGE

- 2.9 Cyanobacterial taste and odour compounds in water
 - T&O can be used as an early warning for cyanobacteria, but not all cyanobacteria produce T&O
 - No relation between T&O and toxin production

- 2.10 Unspecified toxicity and other cyanobacterial metabolites
 - Compounds produced by cyanobacteria showing bioactivity in various test systems (e.g. peptides)
 - Toxic effects of cyanobacteria extracts that cannot be attributed to the well-known compounds

Derivation provisional **life term drinking water GV** MC-LR

$$GV_{\text{chronic}} = \frac{\text{NOAEL} \cdot \text{bw} \cdot P}{\text{UF} \cdot C} = \frac{40 \cdot 60 \cdot 0.8}{1000 \cdot 2} = 1 \mu\text{g} / \text{L}$$

GV_{chronic}
 NOAEL
 on Fav
 bw = bod
 P = frac
 source
 lifetim

however

sed
 her
 for

UF = uncertainty factor (1000 = 10 for interspecies variation \times 10 for intraspecies variation \times 10 for database deficiencies, including use of a subchronic study)

C = daily drinking-water consumption (default = 2 L for an adult).

Review

Accumulation of cyanobacterial toxins in freshwater “seafood” and its consequences for public health: A review

Bas W. Ibelings^{a,b,*}, Ingrid Chorus^c

^a *Eawag, Swiss Federal Institute of Aquatic Sciences and Technology, Centre of Ecology, Evolution and Biogeochemistry, Seestrasse 79, CH-6047 Kastanienbaum, Switzerland*

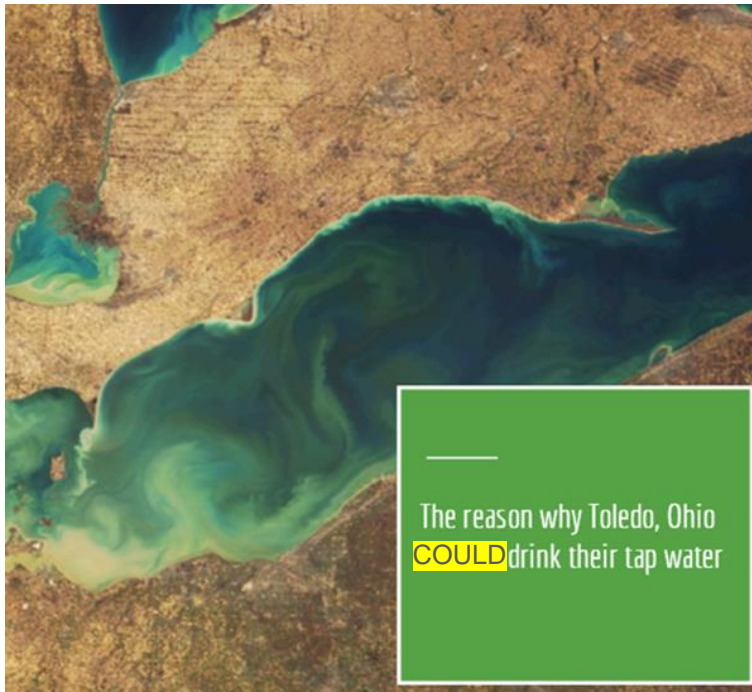
Tolerable doses to microcystin-LR in relation to frequency and duration of exposure

Temporal pattern of exposure and ensuing Tolerable Intake (TI)	Assumptions	Tolerable Intake per kg	Tolerable Intake for a 10 kg child	Tolerable Intake for a 75 kg adult	Guideline value for food ($\mu\text{g kg}^{-1}$)	
					AF = 1	AF = 0.2
Acute TI	NOAEL of 250 $\mu\text{g/kg}$ and day, extrapolation factors of 100 (Fromme et al., 2000)	2.5 μg per kg and single exposure	25 μg per single exposure	190 μg per single exposure	Adult: 1900, Child: 250	Adult: 380, Child: 50
Seasonal TDI	NOAEL of 0.4 $\mu\text{g/kg}$ and day, extrapolation factors of 100 (Chorus and Bartram, 1999, adapted)	0.4 μg per kg and day	4 μg per day	30 μg per day	Adult: 300, Child: 40	Adult: 60, Child: 8
Lifetime TDI	NOAEL of 0.4 $\mu\text{g/kg}$ and day, extrapolation factors of 100 and uncertainty factor of 10 (Chorus and Bartram, 1999)	0.04 μg per kg and day	0.4 μg per day	3 μg per day	Adult: 30, Child: 0.4	Adult: 6, Child: 0.08

Derivation provisional **short term drinking water GV** MC-LR

$$GV_{\text{short-term}} = \frac{\text{NOAEL} * bw * P}{UF * C} = \frac{40 * 60 * 1.0}{100 * 2} \mu\text{g} / \text{L} = 12 \mu\text{g} / \text{L}$$

UF = uncertainty factor (100 = 10 for interspecies variation × 10 for intraspecies variation)



WHO: short-term guideline values are necessary to avoid mis-interpretation of lifetime GVs !

Derivation provisional **recreational-waters GV** MC-LR

SUM!

$$GV_{\text{recreation}} = \frac{\text{NOAEL} * \text{bw}}{\text{UF} * \text{C}} = \frac{40 * 15}{100 * 0.25} \mu\text{g} / \text{L} = 24 \mu\text{g} / \text{L}$$

$GV_{\text{recreation}}$ = guideline value for recreational exposure

NOAEL = no-observed-adverse-effect level (40 $\mu\text{g}/\text{kg}$ bw per day, based on Fawell et al., 1999)

bw = body weight (default = 15 kg for a child)

UF = uncertainty factor (100 = 10 for interspecies variation \times 10 for intraspecies variation)

C = daily incidental water consumption (default = 250 mL for a child).



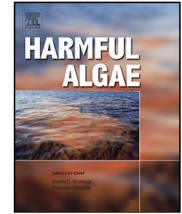


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Current approaches to cyanotoxin risk assessment and risk management around the globe



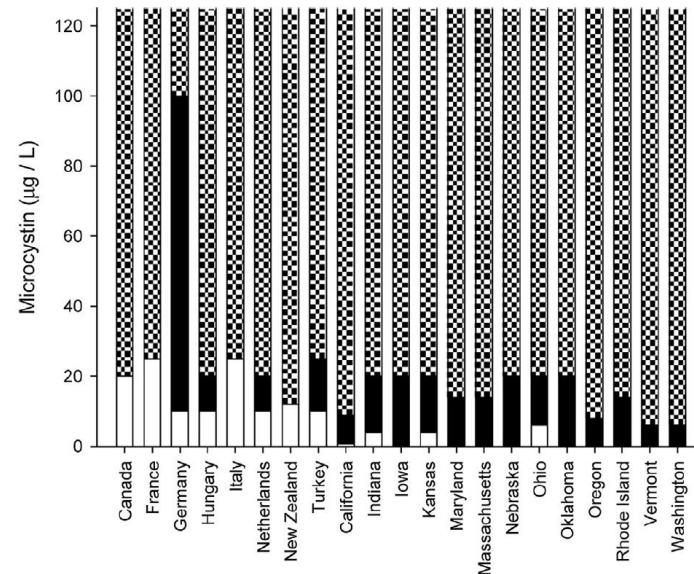
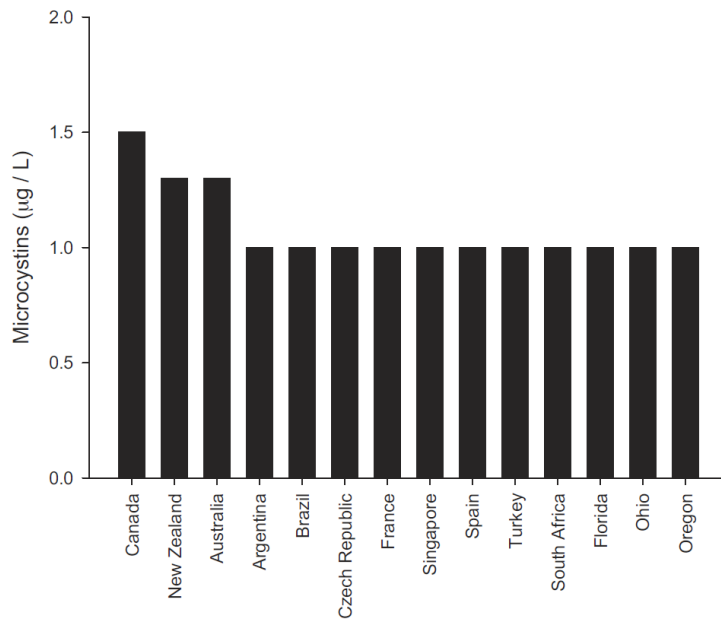
Bas W. Ibelings^{a,*}, Lorraine C. Backer^b, W. Edwin A. Kardinaal^c, Ingrid Chorus^d

^a *Institute F.-A. Forel and Institute of Environmental Sciences University of Geneva, 10 Route de Suisse, 1290 Versoix, Switzerland*

^b *National Center for Environmental Health, 4770 Buford Highway NE, MS F-57, Chamblee, GA 30341, USA*

^c *KWR, Watercycle Research Institute, Groningehaven 7, 3433 PE Nieuwegein, The Netherlands*

^d *German Federal Environment Agency, Corrensplatz 1, 14195 Berlin, Germany*



“Setting the alert levels in the Netherlands is the outcome of intensive discussions between scientists, lake managers and policy makers, in a country known for the highly eutrophic state of its lakes (despite successful restoration efforts ...), where stricter alert levels might result in extended closure of many lakes. Safety clearly must come first, but the protocol used in the Netherlands – in addition to health risks – takes into account the promotion of outdoor activities, feasibility, complexity and costs of monitoring or risk control of cyanobacteria, as well as the ease of communication to the public. Given the large uncertainty in the derivation of TDI for cyanotoxins it is not possible to say whether the higher alert levels in the Netherlands truly result in decreased protection. We merely know it is uncertain.” (p. 68)

The newly derived GV

- GVs be applied to total MCs, total CYNs and total STXs (lack of data for variants)
- **Allocation factors: drinking water 80%** for life time (!), 100% for short time or recreational
- **Life time food or recreation only 20%, short term can be up to 100 %.** Adapt AF local circumstances, be mindful other health-effects
- Water intake: 2L for drinking water
- 250 mL for a child as recreational, daily - incidental - water intake
- UF :
 - MC: 1000 LT, 100 ST, 100 Rec
 - CYN: 1000 LT, 300 ST, 300 Rec
 - STX: 3 Acute, 3 Rec
- Provisional: data base deficiency
- Short-term exposure refers to about 2 weeks, until measures can be implemented to achieve concentrations < lifetime GV

Chapters 2.1 – 2.4

Guideline values / Health-based reference value*

	Microcystin	Cylindrospermopsin	Anatoxin-a	Saxitoxin
Drinking water (life time)	1	0.7	- *	3 **
Short term value (DW)	12	3	30	- 0.3
Bathing water	24	6	60	30

µg / L

- No comprehensive toxicological data available
 - provisional GV/HBRV Health Based Reference Value
 - no life-time HBRV for anatoxin-a*
- Acute drinking water GV for saxitoxin** (derived from acute seafood poisoning)
- Short term values: exceedance of GV for 2 weeks once in a season
- GV can be applied to the sum of all congeners (MC, CYN, ATX, STX)

Potential exposure routes

5.1 Drinking-water and the Alert Levels

Framework (ALF): now includes the *short-term guideline* values

5.2 Recreational exposure and the related ALF. Emphasis is on adapting ALF locally or regionally.

5.3 Food exposure: emphasis to the need of good quality data for assessing exposure

5.4 Renal dialysis: less frequent exposure but at high risk (susceptible subpopulation and kinetics)

5.5 Dietary supplements: increasing consumption expected - good quality data needed

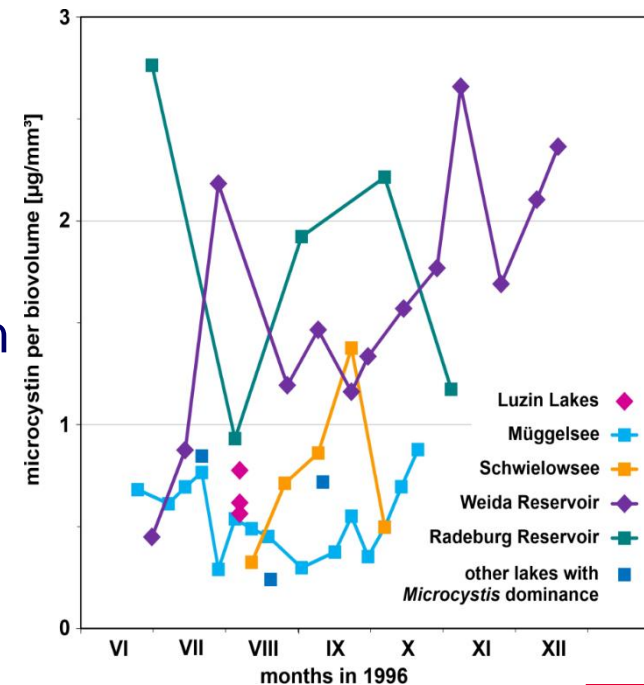


Alert Levels Framework (ALF)

An Alert Levels Framework (ALF) is a **monitoring and management action sequence** (“**decision tree**”), which can be used to provide a rapid response to a cyanobacterial bloom

Adapt ALF to local conditions:

- Parameters used to trigger alerts (e.g. visual assessment, turbidity readings, quantification of cyanobacterial biomass- either biovolume or pigment concentrations-, cell number, cyanotoxin levels)
- Providing they are periodically “calibrated” against toxin levels.
- Indeed, the toxin/biomass ratios can vary not only between waterbodies but also between years for one-and-the-same waterbody



Adapting the ALF to local situation

- Extent and duration of cyanotoxin exposure in relation to other hazards: when pathogens are present and cyanobacterial blooms are short-lived events, a decision might be to tolerate somewhat higher concentrations (as an interim solution) in order to prioritize available capacity and resources on controlling hazards causing the highest risks
- The amount of drinking-water consumed and the **fraction of cyanotoxin allocated to uptake through drinking-water in relation to other exposure pathways**. (GV were derived with an allocation factor=1, assuming each of these exposure to be the dominant source during the short duration)
- Meteorological data, indication of drought or heavy rainfall (often associated to cyanobacterial blooms) should be recorded and considered when following the ALF
- **Local adaptation : consumption freshwater seafood**

Pre-screening water-bodies for elevated risk of blooms and exposure to cyanotoxins:

- Total phosphorus concentrations > 20 µg/L and/or experience of cyanobacterial occurrence
- Intensive recreational activity (Section 5.2.3)

Alternative or complementary entry points for assessment at intervals of about 2 weeks

Assessment by visual site inspection

fairly clear water, slightly turbid, greenish discoloration;
Secchi depth < 1 – 2 m

yes

VIGILANCE LEVEL

yes

Assessment supported by lab analyses

Microscopy showing dominance of cyanobacteria with up to 1 – 4 mm³/L biovolume
or
up to 3 – 12 µg/L chlorophyll-*a* with dominance of cyanobacteria

Assess further characteristics determining its potential to support blooms or scums (Chapter 8)
Assess for cyanobacteria known to be toxin producers (Chapter 3)
If yes, intensify monitoring and/or inform site users about toxic cyanobacteria and how to recognize them (Section 5.2.3; Chapter 15)
Inform relevant authorities

Vigilance Level

Vigilance Level

ALF recreational waters

no

**Pronounced greenish turbidity;
feet barely visible when standing
knee-deep in water**

(Fig. 5.7)

**possibly minor thin green film or
streaks on parts of the surface**

(Fig. 5.6)

Secchi depth < 0.5 – 1 m

ALERT LEVEL 1

Watch for scums

**Investigate further (if possible, conduct
toxin analysis)**

**Inform site users to watch for scums
and avoid activities that can lead to
uptake through mouth or nose,
particularly for children; if this cannot
be controlled keep children out of the
water**

Inform relevant health authorities

no

**Cyanobacterial biovolume
up to 4 – 8 mm³/L**

or

**up to 12-24 µg/L chlorophyll-*a* with
dominance of cyanobacteria**

If toxins are analysed:

≤ 24 µg/L MCs *or*
≤ 6 µg/L CYNs *or*
≤ 60 µg/L ATXs *or*
≤ 30 µg/L STXs



**Alert
Level 1**

**Alert
Level 1**

no

**Visible, thick cyanobacterial scums
covering most of the water surface
in areas used for recreation**

Secchi depth < 0.5 – 1 m

ALERT LEVEL 2

no

Cyanobacterial scum and

> 24 µg/L MCs *or*
> 6 µg/L CYNs *or*
> 60 µg/L ATXs *or*
> 30 µg/L STXs

**Immediate action to prevent contact with scums; possible temporary
prohibition of swimming and other water contact activities**

**Inform site users to stay out of the water and to avoid sports activities
that can lead to scum contact, particularly uptake through mouth or
nose; keep children out of scum**

Inform relevant authorities

Public health follow-up investigation

Alert Levels for biovolume and chlorophyll-a

- The proposed values are derived on the basis of *conservative* – “worst case” - assumptions on ratios of MCs to either biovolume or chlorophyll-a
- Available data on low concentrations of CYN, ATX and STX found in water indicate that the toxin/biomass ratios derived for MCs can be assumed valid for the other cyanotoxins.
- For ATX, GVs are much higher i.e. much less toxic – MCs valid
- However, when Alert Level 2 is exceeded, careful with respect CYN (box 5.1) and STXs: toxin concentrations should be determined for blooms when STX- or CYN-producing species are dominant

Table 5.2 Conservative values for parameters of cyanobacterial biomass indicative of possible occurrence of cyanotoxin concentrations reaching guideline values

Alert Level	Biovolume	Chlorophyll-a	Basis for conservative estimate ^a of toxin/biomass
	MC/BV ≤ 3/l [µg/mm ³]	MC/ Chl.a ≤ 1:1 [µg/µg]	
Alert Level 1 in drinking-water ALF	0.3 mm ³ /L	1 µg/L	GV _{chronic} for MCs in drinking-water: 1 µg/L
Alert Level 2 in drinking-water ALF	4 mm ³ /L	12 µg/L	GV _{short-term} for MCs in drinking-water: 12 µg/L
Alert Level 2 in recreational ALF	8 mm ³ /L	24 µg/L	GV _{recreational} for MCs: 24 µg/L

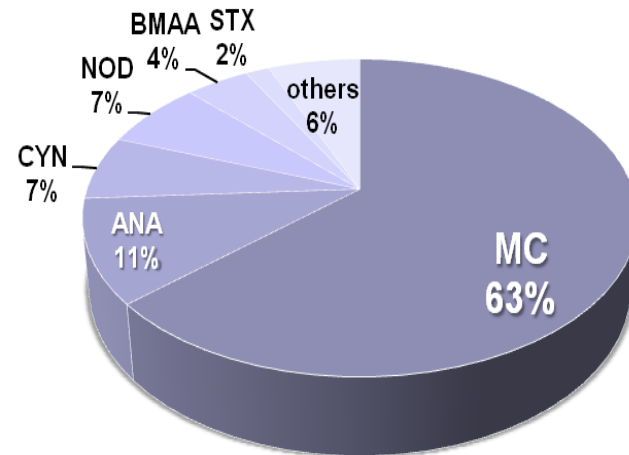
If these Alert Levels are not exceeded, the MCs concentrations are highly unlikely to exceed the respective GVs

Exposure through food



- The great majority of papers deal with MC, and more specifically to **MC-LR**
- Little availability of **robust analytical methods** for matrices other than water
- **Covalently bound MC**
- Only about **25%** of the available studies applied **reliable analytical methods** (especially dated studies do not report recovery, matrix effects etc...)
- **Food preparation (cleaning) important**, cooking inefficient

Occurrence data: distribution among toxins



Testai et al, EFSA Supporting Publications 13(2):998 (2016)

Cyanotoxins in food: potential sources

- Animals grown in aquaculture or harvested as food in freshwater or brackish rivers and lakes
- Blue-green algal food supplements
- Food prepared with contaminated water
- Crops irrigated with contaminated water



Cyanotoxin concentrations in various food sources

Table 5.5 (Continued) Examples of cyanotoxin concentrations in foods

A	B	C	D	E	F	G	H
Organism	Organs or tissue	Cyanotoxin	Content [$\mu\text{g}/\text{kg ww}$]	Max. [μg] ingested with serving of 0.1 kg	% of TI for DW short term (MC, CYN) or acute (STX) exposure	Study type	Reference
Fish							
<i>Aristichthys nobilis</i>	Muscle	MC-RR, MC-YR, MC-LR	177 ²	17.7	74	Field	Chen et al. (2007)
<i>Hypophthalmichthys molitrix</i>	Muscle	MC-RR	0.4 ²	0.0	0.2	Field	Chen et al. (2009b)
<i>Hypophthalmichthys molitrix</i>	Muscle	MC-RR, MC-LR	ND–249 ^c (mean: 39.4)	24.9	104	Field	Chen et al. (2006)
<i>Pomoxis nigromaculatus</i>	Muscle	MC-LR	ND–70	7.0	29	Field	Schmidt et al. (2013)
<i>Cyprinus carpio</i>	Muscle	MC-LR	ND–4	0.4	2	Field	Schmidt et al. (2013)
<i>Cyprinus carpio</i>	Muscle	MC-RR	0.6 ^b	0.1	0.3	Field	Chen et al. (2009b)
<i>Oreochromis niloticus</i>	Muscle	MC-LR ^b	4.2–5.2	0.5	2	Field	Greer et al. (2017)
<i>Geophagus brasiliensis</i>	Muscle	STX _{req}	12–20	2.0	7	Field	Clemente et al. (2010)

For contextualising the health risk due to these concentrations, column “F” relates to the amount of toxin ingested with a serving of 0.1 kg to the percentage of the tolerable intake for short-term (MCs, CYN) or acute (STX) exposure (TI calculated from the NOAEL, UF and bodyweight of 60 kg as given in Chapter 2). Note that these short-term GVs were derived assuming drinking-water to be the major source of exposure to these cyanotoxins, not leaving a proportion to other sources, and that this comparison is intended merely to give a rough estimate of the health relevance of concentrations found in food, not for defining safe levels for food. This table does not take into account the relative toxicities of microcystin congeners.

ND = not detected above the detection limit. STX_{req} = saxitoxin toxicity equivalents.

^a When applied with equal amounts of MC; without MC accumulation was fourfold lower.

^b Secondary technique (i.e., MMPB, ELISA) indicated higher levels may have been present.

^c Converted to wet weight using a wet-to-dry weight ratio of 5.

Cyanotoxin concentrations in various food sources

Table 5.5 Examples of cyanotoxin concentrations in foods

A	B	C	D	E	F	G	H
Organism	Organs or tissue	Cyanotoxin	Content [$\mu\text{g}/\text{kg ww}$]	Max [μg] ingested with serving of 0.1 kg	% of TI for DW short term (MC; CYN) or acute (STX) exposure	Study type	Reference
Crops							
<i>Lactuca sativa</i>	Leaf	MC-LR	2.4–147	14.7	61	Lab	Cordeiro-Araújo et al. (2017)
<i>Lactuca sativa</i>	Unknown	MC-YR, MC-RR	ND–108	10.8	45	Field	Li et al. (2014)
<i>Potamogeton amplifolius</i>	Unknown	MC-RR, MC-LR	ND–68	6.8	29	Field	Li et al. (2014)
<i>Brassica oleracea</i>	Unknown	MC-RR	ND–20	2.0	9	Field	Li et al. (2014)
<i>Brassica rapa</i> var. <i>Parachinensis</i>	Unknown	MC-RR	ND–40	4.0	17	Field	Li et al. (2014)
<i>Brassica oleracea</i> var. <i>Sabellica</i>	Leaf	CYN	3.1	0.3	5	Lab	Kittdler et al. (2012)
<i>Brassica juncea</i>	Leaf	CYN	4.0	0.4	7	Lab	Kittdler et al. (2012)
<i>Lactuca sativa</i>	Leaf	CYN	3.1–8.2	0.8	13	Lab	Cordeiro-Araújo et al. (2017)
<i>Eruca sativa</i>	Leaf	CYN	5.5–11.5	1.1	18	Lab	Cordeiro-Araújo et al. (2017)
<i>Spinacea oleracea</i>	Leaf	CYN	9.5–120	12.0	200	Lab	Llana-Ruiz-Cabello et al. (2019) ^a
<i>Lactuca sativa</i>	Leaf	CYN	2.4–42	4.2	70	Lab	Llana-Ruiz-Cabello et al. (2019) ^a

(Continued)

Cyanotoxin concentrations in various food sources

Table 5.5 (Continued) Examples of cyanotoxin concentrations in foods

A	B	C	D	E	F	G	H
Organism	Organs or tissue	Cyanotoxin	Content [$\mu\text{g}/\text{kg ww}$]	Max. [μg] ingested with serving of 0.1 kg	% of TI for DW short term (MC; CYN) or acute (STX) exposure	Study type	Reference
Molluscs							
<i>Mytilus galloprovincialis</i>	Whole	dmMC-RR ^b	ND–39	3.9	16	Field	Rita et al. (2014)
<i>Patinopecten yessoensis</i>	Whole	MC-LR	ND–4.3	0.43	2	Field	Cui et al. (2018)
<i>Crassostrea virginica</i>	Whole	MC-RR, MC-LR	ND–9.8	1.0	4	Field	Cui et al. (2018)
<i>Mytilus galloprovincialis</i>	Whole	CYN	28.1–41.6	4.2	70	Lab	Freitas et al. (2016)
<i>Anodonta cygnea</i>	Whole	CYN	247	24.7	412	Lab	Saker et al. (2004)
<i>Anodonta cygnea</i>	Whole	STX _{12q}	160–220	22	73	Lab	Pereira et al. (2004)
<i>Alathyrja condola</i>	Muscle	STX _{12q}	144–179	18	60	Lab	Negri & Jones (1995)
Crustaceans							
<i>Astacus astacus</i>	Head/ thorax	[Asp ³ , Dhb ⁷]MC-LR, [Asp ³]MC-RR, [Asp ³ , Dhb ⁷]MC-RR	10	1.0	4	Field	Miles et al. (2013)
<i>Astacus astacus</i>	Tail	[Asp ³ , Dhb ⁷]MC-LR, [Asp ³]MC-RR, [Asp ³ , Dhb ⁷]MC-RR	<1	<0.1	<1	Field	Miles et al. (2013)
<i>Procambarus darkii</i>	Abdomen	MC-RR, MC-LR	1.4–17.1	1.7	7	Field	Rios et al. (2013)
<i>Cherax quadricarinatus</i>	Muscle	CYN	205	20.5	342	Field	Saker & Eaglesham (1999)

(Continued)

Checklist assessing risk cyanotoxin exposure food

- **Blooms** potentially toxic species present?
 - Toxin content cyanobacteria
 - Recent period?
- Are organisms **harvested**?
 - Filter feeders?
 - **Viscera removed**?
- Are crops irrigated?
 - Cyanobacterial cells present on surface?
- Toxins likely from other sources (**drinking water**)
- **Seasonal** or year round? Amounts consumed local setting?
- Clarify possible **toxin dose**
 - Necessary to **restrict access** to food contaminated food?
- Measures for **protection**?
- Raise **public awareness**

Guideline values food



- MCYST 5.6 $\mu\text{g} / \text{kg}$ bw fish and 51 $\mu\text{g} / \text{kg}$ bw molluscs (France)
- CYN 18 $\mu\text{g} / \text{kg}$ bw (AUS), 70 $\mu\text{g} / \text{kg}$ bw (California)
- STX 800 $\mu\text{g} / \text{kg}$ bw (AUS, CAN), 5000 $\mu\text{g} / \text{kg}$ bw (California)

Tolerable doses to microcystin-LR in relation to frequency and duration of exposure

Temporal pattern of exposure and ensuing Tolerable Intake (TI)	Assumptions	Tolerable Intake per kg	Tolerable Intake for a 10 kg child	Tolerable Intake for a 75 kg adult	Guideline value for food ($\mu\text{g kg}^{-1}$)	
					AF = 1	AF = 0.2
Acute TI	NOAEL of 250 $\mu\text{g}/\text{kg}$ and day, extrapolation factors of 100 (Fromme et al., 2000)	2.5 μg per kg and single exposure	25 μg per single exposure	190 μg per single exposure	Adult: 1900, Child: 250	Adult: 380, Child: 50
Seasonal TDI	NOAEL of 0.4 $\mu\text{g}/\text{kg}$ and day, extrapolation factors of 100 (Chorus and Bartram, 1999, adapted)	0.4 μg per kg and day	4 μg per day	30 μg per day	Adult: 300, Child: 40	Adult: 60, Child: 8
Lifetime TDI	NOAEL of 0.4 $\mu\text{g}/\text{kg}$ and day, extrapolation factors of 100 and uncertainty factor of 10 (Chorus and Bartram, 1999)	0.04 μg per kg and day	0.4 μg per day	3 μg per day	Adult: 30, Child: 0.4	Adult: 6, Child: 0.08

What defines a risk?

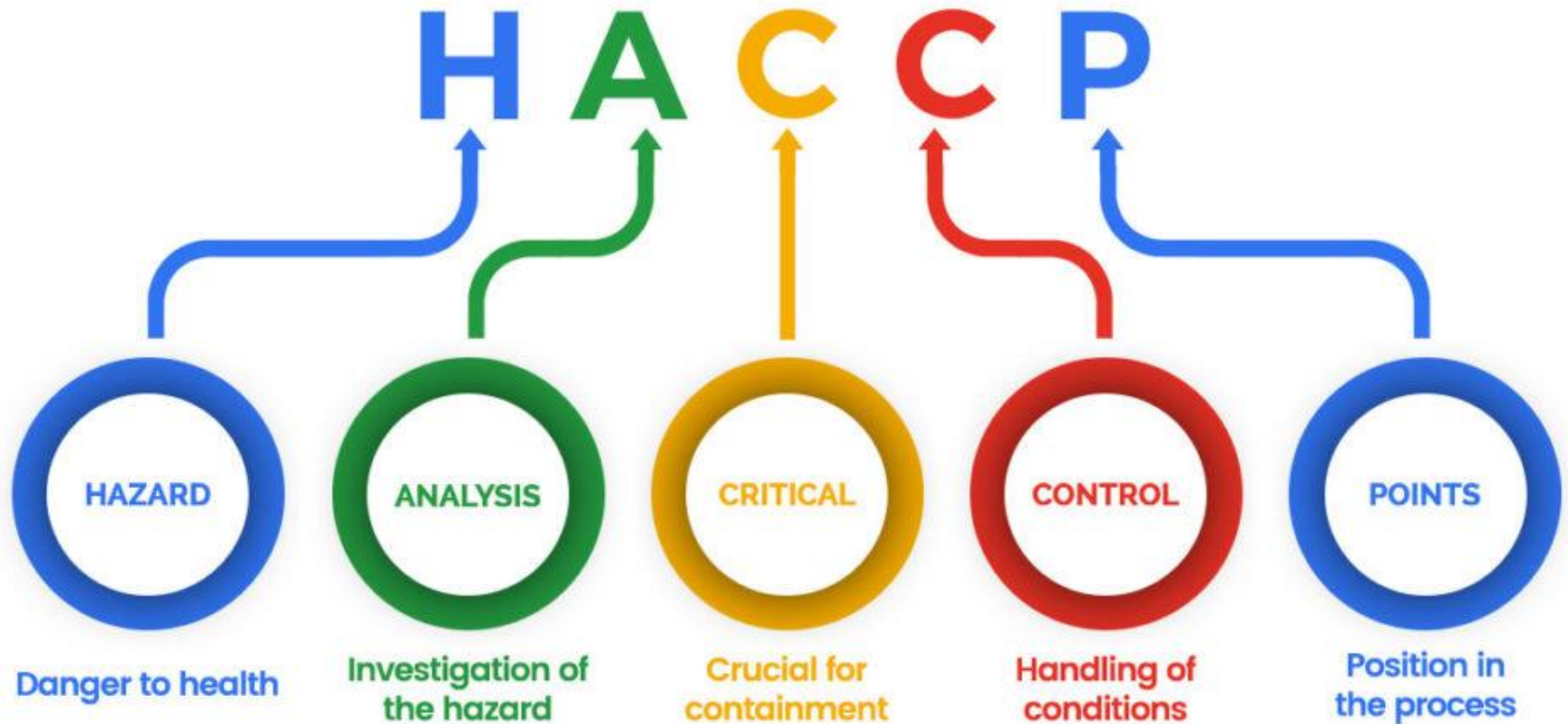
	daily exposure to MC at up to 20-fold the TDI	protein deficient diet	infant undernourishment and starvation
Likelihood of occurrence ↑	dental damage from excessive sugar in diet	daily exposure to MCYST at >100-fold the TDI	infectious diarrhoea from Campylobacter contamination
		protein need covered from fat-rich low quality sources	mercury accumulation from seafood
			Severity of public health consequences →

Multiple Barriers

- Common element in many National and International Water Quality Guides
 - HCCAP
 - Water Safety Plans
 - Drinking Water Guidelines
 - Water Quality Guidelines
- “Catchment to Tap”
 - Systematically assess hazards, risk and control measures
- ***Toxic cyanobacteria in water*** (2nd edition) provides a comprehensive guide to inform state and national guidelines, assess situations and manage risk

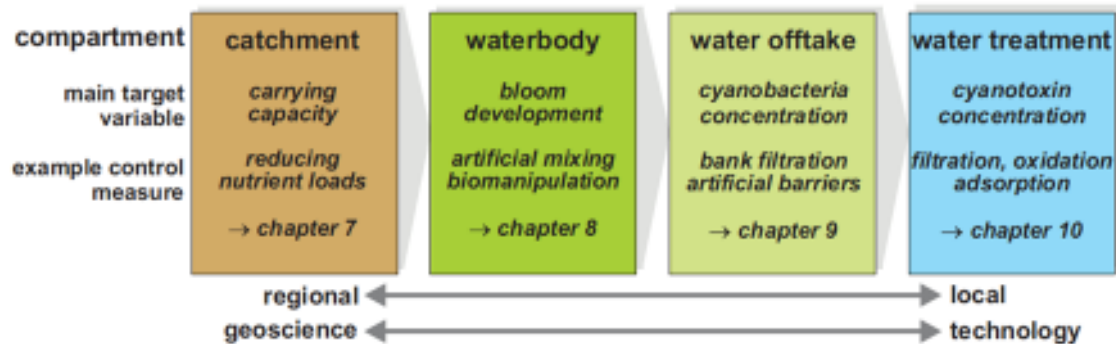


HACCP CERTIFICATION



Critical control points and control measures

- *Toxic cyanobacteria in water* (2nd edition) outlines critical control points and control measures



- Advice to tailor control measures to the local situation and the cyanobacterial risk



Uiminen kielletty

