Study Title

Independent Laboratory Validation (ILV) of the Analytical Method
for the Determination of Benzobicyclon and its Metabolites 1315P-070, 1315P-076,
1315P-570, 1315P-683, and 1315P-960 in Soil by LC-MS/MS

Data Requirements

OCSPP Guideline 850.6100
OCSPP Guideline 850.7100
OCSPP Guideline 860.1340

2.0 MATERIALS AND METHODS

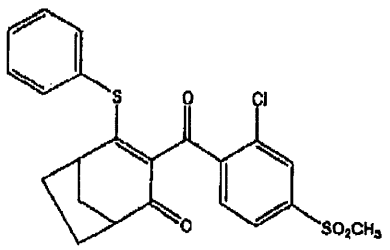
2.1 Study Protocol

This study was performed following the Smithers Viscient protocol entitled "Independent Laboratory Validation (ILV) of the Analytical Method for the Determination of Benzobicyclon and its Metabolites 1315P-070, 1315P-076, 1315P-570, 1315P-683, and 1315P-960 in Soil by LC-MS/MS" (Appendix 1). The methods described in this protocol meet the requirements specified in the OCSPP Guideline 850.6100: Environmental Chemistry Methods and Associated Independent Laboratory Validation (U.S. EPA, 2012), OCSPP Guideline 850.7100 (U.S. EPA, 1996a), SANCO/3029/99 rev. 4 (EC, 2000), SANCO/825/00 rev. 8.1 (EC, 2010), and OSCPP Guideline 860.1340: Residue Analytical Method (U.S. EPA, 1996b).

2.2 Test Substances

The test substance, Benzobicyclon, was received on 10 January 2017 from EAG Laboratories, Hercules, California. The following information was provided:

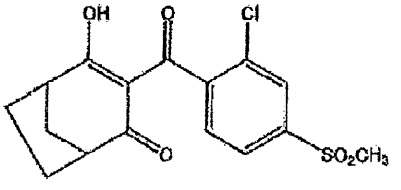
Name:	Benzobicyclon
Chemical Name:	3-[2-chloro-4-(methylsulfonyl)benzoyl]-4-(phenylthio)bicyclo [3.2.1] oct-3-en-2-one
Molecular Weight:	447.0 g/mole
Amount Received:	0.72 g
Lot No.:	1L0108
CAS No.:	156963-66-5
Purity:	99.3% (Certificate of Analysis, Appendix 2)
Expiration Date:	1 April 2018
Chemical Structure:	



Upon receipt at Smithers Viscient, the test substance (SMV No. 8696) was stored in a freezer in the original container. Concentrations were adjusted for the purity of the test substance.

The test substance, 1315P-070, was received on 10 January 2017 from EAG Laboratories, Hercules, California. The following information was provided:

Name:	1315P-070
Synonym:	Benzobicyclon metabolite 1315P-070
Chemical Name:	3-[(2-Chloro-4-methylsulfonyl)benzoyl)]-bicyclo[3.2.1]octane-2,4-dione
Molecular Weight:	354.8 g/mole
Amount Received:	0.59 g
Lot No.:	95Z25
CAS No.:	126656-88-0
Purity:	98.8% (Certificate of Analysis, Appendix 2)
Recertification Date:	24 February 2018
Chemical Structure:	

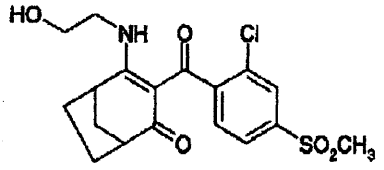


Upon receipt at Smithers Viscient, the test substance (SMV No. 8697) was stored in a freezer in the original container in a dark, ventilated cabinet. Concentrations were adjusted for the purity of the test substance.

The test substance, 1315P-076, was received on 8 January 2016 from PTRL West, Hercules, California. The following information was provided:

Name:	1315P-076
Synonym:	Benzobicyclon metabolite 1315P-076
Chemical Name:	(3-(2-chloro-4-(methylsulfonylbenzoyl)-4-(2-hydroxyethylamino))bicyclo[3.2.1]oct-3-en-2-one
Molecular Weight:	397.9 g/mole

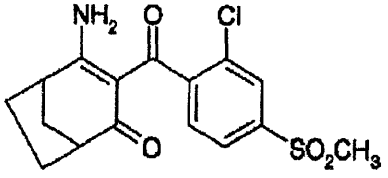
Amount Received:	5 g
Lot No.:	TNA-10-074
CAS No.:	Not Listed
Purity:	99.5% (Certificate of Analysis, Appendix 2)
Expiration Date:	23 February 2019
Chemical Structure:	



Upon receipt at Smithers Viscient, the test substance (SMV No. 8028) was stored at room temperature in a dark, ventilated cabinet in the original container. Concentrations were adjusted for the purity of the test substance.

The test substance, 1315P-570, was received on 8 January 2016 from PTRL West, Hercules, California. The following information was provided:

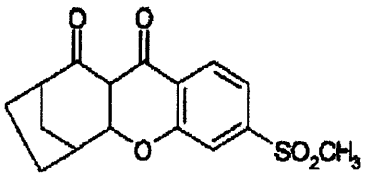
Name:	1315P-570
Synonym:	Benzobicyclon metabolite 1315P-570
Chemical Name:	(3-(2-Chloro-4-methylsulfonylbenzoyl)-4-amino)bicyclo[3.2.1]oct-3-en-2-one
Molecular Weight:	353.8 g/mole
Amount Received:	5 g
Lot No.:	TNA-9-186
CAS No.:	Not Listed
Purity:	99.8% (Certificate of Analysis, Appendix 2)
Recertification Date:	28 February 2019
Chemical Structure:	



Upon receipt at Smithers Viscient, the test substance (SMV No. 8029) was stored at room temperature in the original container in a dark, ventilated cabinet. Concentrations were adjusted for the purity of the test substance.

The test substance, 1315P-683, was received on 5 May 2016 from PTRL West, Hercules, California. The following information was provided:

Name:	1315P-683
Synonym:	Benzobicyclon metabolite 1315P-683
Chemical Name:	3,4-dihydro-2,4-ethylene-6-methylsulfonyl-1*H*-xanthene-1,9(2*H*)-dione
Molecular Weight:	318.35 g/mole
Amount Received:	4.50 g
Lot No.:	TM-8-198
CAS No.:	Not Listed
Purity:	99.77% (Certificate of Analysis, Appendix 2)
Recertification Date:	21 February 2019
Chemical Structure:	

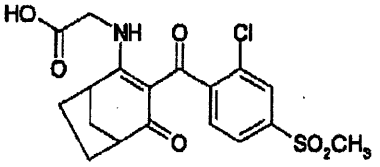


Upon receipt at Smithers Viscient, the test substance (SMV No. 8256) was stored at room temperature in the original container in a dark, ventilated cabinet. Concentrations were adjusted for the purity of the test substance.

The test substance, 1315P-960, was received on 2 February 2017 from EAG Laboratories, Hercules, California. The following information was provided:

Name:	1315P-960
Synonym:	Benzobicyclon metabolite 1315P-960
Chemical Name:	4-(carboxymethylamino)-3-[2-chloro-4-(methylsulfonyl)benzoyl]bicyclo[3.2.1]oct-3-en-2-one
Molecular Weight:	411.86 g/mole
Amount Received:	0.5 g
Lot No.:	H/M-13-57-2
CAS No.:	Not Listed
Purity:	99.48% (Certificate of Analysis, Appendix 2)
Recertification Date:	24 March 2019

Chemical Structure:



Upon receipt at Smithers Viscient, the test substance (SMV No. 8732) was stored in a freezer in the original container. Concentrations were adjusted for the purity of the test substance.

Determination of stability and characterization, verification of the test substance identity, maintenance of records on the test substances, and archival of samples of the test substances are the responsibility of the Study Sponsor.

2.3 Reagents

1.	Acetonitrile:	EMD, reagent grade
2.	Celite:	Fisher, reagent grade
3.	Citric acid:	Fisher, reagent grade
4.	Formic acid:	BDH, reagent grade
5.	Methanol:	EMD, reagent grade
6.	Purified reagent water:	prepared from a Millipore Milli-Q Direct 8 system (meeting ASTM Type II requirements)
7.	Ultra-pure water:	Fisher, reagent grade

2.4 Equipment

1.	Instruments:	AB Sciex API 5000 mass spectrometer equipped with an ESI Turbo V source Shimadzu SIL-20ACHT autoinjector Shimadzu DGU-20A3V vacuum degasser Shimadzu DGU-20A5R vacuum degasser Shimadzu LC-20AD solvent delivery pumps Shimadzu CTO-20A column compartment Shimadzu CBM-20A communications bus Analyst 1.4.2 software for data acquisition

2.	Balances:	Mettler Toledo XSE205DU, Mettler PG-2002-S, and Mettler PJ-3000
3.	Moisture balance:	Mettler Toledo Moisture Balance HB43-S
4.	Shaker table:	VWR Standard Analog Shaker 3500STD
5.	Laboratory equipment:	Volumetric flasks, disposable glass pipets, positive displacement pipets, Whatman #5 filter paper, PTFE syringe filters, stir bars, stir plates, vortexers, 250-mL centrifuge bottles, amber bottles, autosampler vials, and amber glass bottles with Teflon-lined caps

2.5 Test Systems

The test systems evaluated during this study were soils representative of the type of matrix this method was intended to analyze. The soils used for this ILV analysis were Clay Loam (SMV 14Dec16 Soil-B) and Loam soil (SMV 29Mar17 Soil-B). The soil characterization data are listed in the table below.

Soil Type	% Sand, Silt, Clay	Bulk Density (gm/cc)	CEC (meq/100 g)	% Organic Matter (Walkley Black)	% Moisutre at 1/3 Bar	pH in 1:1 Soil:Water Ratio
Clay loam	40, 28, 32	0.99	19.2	5.6	31.5	5.4
Loam	44, 36, 20	0.84	23.0	12.0	45.7	6.6

2.6 Preparation of Stock Solutions

Primary stock solutions were typically prepared as described in the table below. All volumes and masses may be scaled up or down as necessary.

Primary Stock ID	Amount Weighed (g), Net Weight	Amount Weighed (g), as Active Ingredient	Stock Solvent	Final Volume (mL)	Primary Stock Concentration (mg/L)	Primary Stock Use
8696B	0.0506	0.0502	Acetonitrile	50.0	1000	Fortification stock solution
8696D	0.0504	0.0500		50.0	1000	Fortification stock solution
8697C	0.05066	0.05005		50.0	1000	Fortification stock solution
8697D	0.0508	0.0502		50.0	1000	Fortification stock solution
8028P	0.0503	0.0500		50.0	1000	Fortification stock solution
8028Q	0.0503	0.0500		50.0	1000	Fortification stock solution
8029P	0.05013	0.05003		50.0	1000	Fortification stock solution
8029Q	0.0501	0.0500		50.0	1000	Fortification stock solution
8256C	0.05032	0.05020		50.0	1000	Fortification stock solution
8256D	0.0502	0.0501		50.0	1000	Fortification stock solution
8732C	0.05026	0.05000		50.0	1000	Fortification stock solution
8732D	0.0503	0.0500		50.0	1000	Fortification stock solution

Fortification and intermediate stock solutions were typically prepared as described in the table below.

Fortifying Stock ID	Fortifying Stock Concentration (mg/L)	Volume of Fortification (mL)	Final Volume (mL)	Stock Solvent	Stock ID	Stock Concentration (mg/L)	Stock Use
8696B	1000	0.250	50.0	Acetonitrile	Soil Fort. Sol. A	5.00	High-level recovery samples for clay loam and loam soils, and a fortification stock solution
8697C	1000	0.250				5.00	
8028P	1000	0.250				5.00	
8029P	1000	0.250				5.00	
8256C	1000	0.250				5.00	
8732C	1000	0.250				5.00	
8696D	1000	0.250	50.0	Acetonitrile	Soil Fort. Sol. D	5.00	Intermediate stock solution
8697D	1000	0.250				5.00	
8028Q	1000	0.250				5.00	
8029Q	1000	0.250				5.00	
8256D	1000	0.250				5.00	
8732D	1000	0.250				5.00	
Soil Fort. Sol. A	5.00[a]	5.00	50.0	Acetonitrile	Soil Fort. Sol. B	0.500[a]	LOQ-level recovery samples in clay loam and loam soils and an analytical, intermediate stock solution
Soil Fort. Sol. B	0.500[a]	5.00	50.0	40/60 Acetonitrile/purified reagent water (v/v)	Soil Ana. Sol. C	0.0500[a]	Calibration standards
Soil Fort. Sol. D	5.00[a]	5.00	50.0	Acetonitrile	Soil Fort. Sol. E	0.500[a]	An analytical, intermediate stock solution
Soil Fort. Sol. E	0.500[a]	5.00	50.0	40/60 Acetonitrile/purified reagent water (v/v)	Soil Ana. Sol. F	0.0500[a]	Calibration standards

[a] Mixed stock solution with expressed concentration for Benzobicyclon and metabolites 1315P-070, 1315P-076, 1315P-570, 1315P-683, and 1315P-960.

All stock solutions were stored in a freezer (-25 to -10 °C) in amber glass bottles fitted with Teflon-lined caps.

2.7 Liquid Reagent Preparation

All volumes and masses may be scaled up or down as necessary.

A 40/60 acetonitrile/purified reagent water (v/v) liquid reagent solution was typically prepared by combining 40 mL of acetonitrile and 60 mL of purified reagent water. The solution was mixed well using a stir bar and stir plate for five minutes.

A 0.2% formic acid in ultra-pure water mobile phase solution was typically prepared by adding 4.00 mL of formic acid to 2000 mL of ultra-pure water. The solution was mixed well using a stir bar and stir plate for five minutes, then degassed under vacuum with sonication for ten minutes.

A 0.2% formic acid in acetonitrile mobile phase solution was typically prepared by adding 4.00 mL of formic acid to 2000 mL of acetonitrile. The solution was mixed well using a stir bar and stir plate for five minutes, then degassed under vacuum with sonication for 10 minutes.

A 30/30/40 acetonitrile/methanol/purified reagent water (v/v/v) autosampler needle wash solution was typically prepared by combining 1500 mL of acetonitrile, 1500 mL of methanol, and 2000 mL of purified reagent water.

A 0.55 M citric acid in purified reagent water solution was typically prepared by bringing 52.53 g of citric acid to volume with purified reagent water in a 500-mL volumetric flask. The solution was mixed using a stir bar and stir plate for five minutes.

An 80/20 acetonitrile/0.55 M citric acid (v/v) liquid reagent solution was typically prepared by combining 1200 mL of acetonitrile and 300 mL of 0.55 M citric acid. The solution was mixed well using a stir bar and stir plate for five minutes.

2.8 Preparation of Calibration Standards

Standards were prepared in the final fraction of Ctrl C-C or Ctrl C-L using the 50.0 µg/L stock solution according to the tables below. Following fortification, each solution was vortex-mixed for 15 seconds, then standards were transferred to amber autosampler vials for analysis.

Clay Loam Soil

Fortifying Stock ID	Stock Concentration (µg/L)	Fortification Volume (mL)	Final Volume (mL)	Standard Concentration (µg/L)	Sample ID
Soil Ana. Sol. C	50.0	0.0200	10.0	0.100	Std CS-1
		0.0500	10.0	0.250	Std CS-2
		0.100	10.0	0.500	Std CS-3
		0.200	10.0	1.00	Std CS-4
		0.500	10.0	2.50	Std CS-5
		1.00	10.0	5.00	Std CS-6

Loam Soil

Fortifying Stock ID	Stock Concentration (µg/L)	Fortification Volume (mL)	Final Volume (mL)	Standard Concentration (µg/L)	Sample ID
Soil Ana. Sol. C	50.0	0.0200	10.0	0.100	Std LS-1
		0.0500	10.0	0.250	Std LS-2
		0.100	10.0	0.500	Std LS-3
		0.200	10.0	1.00	Std LS-4
		0.500	10.0	2.50	Std LS-5
		1.00	10.0	5.00	Std LS-6

2.9 Sample Fortification and Preparation

Thirteen aliquots of clay loam soil (12.20 g, wet weight) were weighed into individual 250-mL Nalgene centrifuge tubes. Five replicates were dosed with the 500 µg/L soil fortification solution and five aliquots were dosed with the 5000 µg/L soil fortification solution to obtain concentrations of 5.00 and 50.0 µg/kg (ppb), respectively. Three aliquots were left unfortified to serve as controls and an additional sample was extracted using only solvents as a reagent blank. The dosing procedure is detailed in the following table.

Clay Loam Soil

Sample ID	Stock ID	Fortifying Stock Concentration (µg/L)	Fortification Volume (mL)	Wet Soil Weight (g)	Nominal Concentration (µg/kg)
Reagent Blk A-C	NA[a]	NA	NA	NA	0.00
Control A-C, B-C, & C-C	NA	NA	NA	12.20	0.00
LOQ A-C, B-C, C-C, D-C, & E-C	Soil Fort. Sol. B	500	0.100	12.20	5.00
High A-C, B-C, C-C, D-C, & E-C	Soil Fort. Sol A	5000	0.100	12.20	50.0

[a] NA = Not Applicable

Thirteen aliquots of loam soil (12.99 g, wet weight) were weighed into individual 250-mL Nalgene centrifuge tubes. Five replicates were dosed with the 500 µg/L sub-stock solution and five aliquots were dosed with the 5000 µg/L sub-stock solution to obtain concentrations of 5.00 and 50.0 µg/kg (ppb), respectively. Three aliquots were left unfortified to serve as controls and an additional sample was extracted using only solvents as a reagent blank. The dosing procedure is detailed in the following table.

Loam Soil

Sample ID	Stock ID	Fortifying Stock Concentration (µg/L)	Fortification Volume (mL)	Wet Soil Weight (g)	Nominal Concentration (µg/kg)
Reagent Blk A-L	NA[a]	NA	NA	NA	0.00
Control A-L, B-L, & C-L	NA	NA	NA	12.99	0.00
LOQ A-L, B-L C-L, D-L, & E-L	Soil Fort. Sol. B	500	0.100	12.99	5.00
High A-L, B-L, C-L, D-L, & E-L	Soil Fort. Sol A	5000	0.100	12.99	50.0

[a] NA = Not Applicable

2.10 Extraction and Dilution of Fortified Recovery Samples

Samples were extracted once with 90 mL of 80/20 acetonitrile/0.55M citric acid (v/v). Samples were placed on an orbital shaker table for 30 minutes at 250 rpm, (refer to Method Differences).

The entirety of each sample was vacuum filtered through a 1-cm thick layer of celite on Whatman #5 filter paper. Each filtered extract was transferred to a 100-mLvolumetric flask. The filter-cake of each sample was rinsed with 10 mL of 80/20 acetonitrile/0.55 M citric acid (v/v). The rinsate was transferred to the 100-mL volumetric flask. The extracts were brought to volume with 80/20 acetonitrile/0.55 M citric acid (v/v) and vortex mixed for 15 seconds. An aliquot of each sample extract was filtered through a 0.45-µm PTFE syringe filter.

Following filtration, the samples were diluted into the calibration standard range with 40/60 acetonitrile/purified reagent water (v/v). Samples and calibration standards were transferred to autosampler vials for LC-MS/MS analysis.

Clay Loam soil samples for analysis were prepared as described in the following table.

Sample ID	Nominal Concentration (µg/kg)	Dry Soil Weight (g)	Final Volume[a] (mL)	Sub-Sample Volume (mL)	Final Volume[b] (mL)	Sub-Sample Volume (mL)	Final Volume[c] (mL)	Dilution Factor
Reagent Blk A-C	0.00	NA[d]	100	5.00	10.0	NA	NA	NA
Control A-C, B-C, & C-C	0.00	10.0	100	5.00	10.0	NA	NA	20.0
LOQ A-C, B-C, C-C, D-C, & E-C	5.00	10.0	100	5.00	10.0	NA	NA	20.0
High A-C, B-C, C-C, D-C, & E-C	50.0	10.0	100	5.00	10.0	NA	NA	20.0

[a] Extraction Solvent: 80/20 acetonitrile/0.55 M citric acid solution (v/v).
[b] Dilution solvent: purified reagent water and labelled sample id-M.
[c] Diluted with 40/60 acetonitrile/purified reagent water (v/v).
[d] NA = Not Applicable

Loam soil samples for analysis were prepared as described in the following table.

Sample ID	Nominal Concentration (µg/kg)	Dry Soil Weight (g)	Final Volume[a] (mL)	Sub-Sample Volume (mL)	Final Volume[b] (mL)	Sub-Sample Volume (mL)	Final Volume[c] (mL)	Dilution Factor
Reagent Blk A-L	0.00	NA[d]	100	5.00	10.0	NA	NA	NA
Control A-L, B-L, & C-L	0.00	10.0	100	5.00	10.0	NA	NA	20.0
LOQ A-L, B-L, C-L, D-L, & E-L	5.00	10.0	100	5.00	10.0	NA	NA	20.0
High A-L, B-L, C-L, D-L, & E-L	50.0	10.0	100	5.00	10.0	1.00[e]	10.0[e]	20/200[f]

[a] Extraction Solvent: 80/20 acetonitrile/0.55 M citric acid solution (v/v).
[b] Dilution solvent: purified reagent water.
[c] Diluted with final fraction of Ctrl C-L and labelled sample id-MM-3.
[d] NA = Not Applicable
[e] This final dilution was only performed for Benzobicyclon samples.
[f] Ratio expresses metabolites/Benzobicyclon dilution factors.

2.11 LC-MS/MS Instrumental Conditions

The LC-MS/MS analysis was conducted using the following instrumental conditions:

LC Parameters:

Column: Phenomenex Luna C18, 3 µm, 30 × 2 mm
Mobile Phase A: 0.2% formic acid in ultra-pure reagent water
Mobile Phase B: 0.2% formic acid in acetonitrile
Gradient:

Time (min.)	Flow rate (mL/min.)	Solvent A (%)	Solvent B (%)
0.01	0.500	90.0	10.0
7.00	0.500	5.00	95.0
8.00	0.500	5.00	95.0
8.10	0.500	90.0	10.0
9.50	0.500	90.0	10.0

Run time: 9.50 minutes
Injector Wash solvent: 30/30/40 acetonitrile/methanol/purified reagent water (v/v/v)
Column temperature: 40 °C
Sample temperature: 5 °C
Injection volume: 10.0 µL

Approximate Retention Times:

Analyte	Retention Time (min)
Benzobicyclon	4.54
1315P-070	3.46
1315P-076	2.16
1315P-570	2.25
1315P-683	2.08
1315P-960	2.24

MS Parameters:

Instrument:	MDS Sciex API 5000 mass spectrometer
Ionization Mode:	Positive (+) ESI
Resolution Q1/Q3:	Unit/Unit
Ion Spray Voltage:	5500 V
Scan type:	MRM
Source Temperature:	550 °C
Curtain Gas:	20.00
Ion Source – Gas 1 / Gas 2:	80.00 / 80.00
Collision Gas:	12.00
Collision Cell Entrance Potential:	10.00
Declustering Potential:	100.00

Analyte	Q1/Q3 *m/z*	Dwell Time (msec)	Collision Energy	Collision Cell Exit Potential
Benzobicyclon	447.44/257.07 (Primary)	40.00	52.00	17.00
	447.44/229.10 (Confirmatory)	20.00	58.00	17.00
1315P-070	355.35/165.09 (Primary)	20.00	36.20	20.00
	355.35/183.09 (Confirmatory)	20.00	33.50	20.00
1315P-076	398.30/208.14 (Primary)	40.00	41.50	20.00
	398.3/319.10 (Confirmatory)	20.00	42.80	20.00
1315P-570	354.34/164.10 (Primary)	40.00	42.40	20.00
	354.34/318.10 (Confirmatory)	20.00	37.00	20.00
1315P-683	319.33/240.12 (Primary)	40.00	53.00	20.00
	319.33/212.10 (Confirmatory)	20.00	57.00	20.00
1315P-960	412.33/176.15 (Primary)	40.00	54.50	20.00
	412.33/222.12 (Confirmatory)	20.00	38.60	20.00

2.11.1 Preparation of Calibration Standard Curve

Two sets of calibration standards were analyzed with each sample set; calibration standards were interspersed among analysis of the recovery samples, every two to six injections. Injection of recovery samples and calibration standards onto the chromatographic system was performed by programmed automated injection.

2.11.2 Method Differences

The analytical method used for Benzobiclyclon and its metabolites in this independent laboratory validation followed the procedures described in the original method validation. The analytical method used for Benzobicyclon and its metabolites in this independent laboratory validation required the following minor modifications from the original method validation.

- The validated method did not specify shaking speed. In this study, acceptable results were obtained when samples were placed on an orbital shaker table at 250 rpm for 30 minutes.
- Mass spectrometer parameters were optimized for sensitivity and linearity, as necessary.
- The validated method suggested bracketed standards with a curve check standard every two to six sample injections. In this study, acceptable results were obtained when calibration curves were generated from calibration standards run interspersed among recovery samples.
- The validated method, with the soils tested, utilized solvent-based standards for calibration curves. In this study, matrix effects were observed for the soil matrices tested. Acceptable results were obtained with the utilization of matrix-matched standards for calibration curves.

2.12 Evaluation of Precision, Accuracy, Specificity, and Linearity

The accuracy was reported in terms of percent recovery of the LOQ and 10 × LOQ recovery samples. Recoveries of 70.0 to 120% of nominal were considered acceptable, with no corrections made for procedural recoveries during the study. The precision was reported in terms of the standard deviation and relative standard deviation (RSD) for the retention time, the peak area quantitation, and the percent recovery values of the LOQ and 10 × LOQ recovery samples. The retention time should have an RSD of less than or equal to 2%. The RSD of the peak area based quantitation and of the recovery values should be less than or equal to 20%.

Specificity of the method was determined by examination of the control samples for peaks at the same retention time as Benzobicyclon and metabolites which might interfere with the quantitation of the analytes. Interferences with peak areas that are less than 50% of the LOQ are not considered significant. Linearity of the method was determined by the correlation coefficient (r), y-intercept, and slope of the regression line. A 1/x weighted linear regression was used for the LC-MS/MS analysis. The calibration curves were evaluated based on the correlation coefficient and the recoveries of the calibration standards. The signal response data should have an intercept close to zero and a correlation coefficient (r) not less than 0.995. The precision of the method at the LOQ was reported in terms of the coefficient of variation of the observed recovery values.

2.15 Critical Steps

For the particular soils that were investigated, the original analytical method utilized solvent-based standards for quantification. With the soils tested in this ILV, matrix effects were observed. Thus, it is recommended that prior to implementation of the analytical method that a matrix effects investigation be performed.

With analyses organized around bracketed standards (calibration standards before and after complete set of recovery samples), a number of calibration curves failed acceptance criteria. Calibration curves generated from matrix-matched standards interspersed among recovery samples met acceptance criteria and provided for quantification of analyte. Thus, it is recommended that the analysis be run with interspersed calibration standards.

3.0 CALCULATIONS

A calibration curve was constructed by plotting the analyte concentration (µg/L) of the calibration standards against the peak area of the analyte in the calibration standards. The equation of the line (equation 1) was algebraically manipulated to give equation 2. The concentration of test substance in each recovery sample was calculated using the slope and intercept from the linear regression analysis with 1/x weighting, the detector response, and the dilution factor of the recovery sample. Equations 2 and 3 were then used to calculate measured concentrations and analytical results.

$$(1)\quad y = mx + b$$

$$(2)\quad DC(x) = \frac{(y - b)}{m}$$

$$(3)\quad A = DC \times DF$$

where:

x	=	analyte concentration (µg/L)
y	=	detector response (peak area) from the chromatogram

b	=	y-intercept from the regression analysis
m	=	slope from the regression analysis
DC (x)	=	detected concentration (µg/L) in the sample
DF	=	dilution factor (final volume of the sample divided by the original sample volume (mL/g))
A	=	analytical result (µg/kg), concentration in the original sample

NOTE: A 1/x weighting was used for calibration curves and sample quantitation using Analyst software, version 1.6.

The Instrument LOD was calculated using the following equation:

$$(4)\ LOD = (3 \times SN_{ctl}) / Resp_{LS} \times Conc_{LS}$$

where:

SN_{ctl}	=	Mean signal to noise in height of the control samples (or blanks)
$Resp_{LS}$	=	Mean response in height of the two low calibration standards (0.100 µg/L)
$Conc_{LS}$	=	Concentration of the low calibration standard (µg/L)
LOD	=	Instrument Limit of Detection for the analysis (µg/L)

The Overall Method LOD was calculated using the following equation:

$$(5)\ LOD_{Overall} = LOD \times DF_{Ctl}$$

where:

LOD	=	Limit of Detection calculated from signal to noise ratio (µg/L)
DF_{Ctl}	=	Dilution factor for control sample
$LOD_{Overall}$	=	Overall Method Limit of Detection

PROTOCOL DEVIATIONS

No deviations from the protocol occurred during this study.

APPENDIX 1 – STUDY PROTOCOL

Independent Laboratory Validation (ILV) of the Analytical Method for the Determination of Benzobicyclon and its Metabolites 1315P-070, 1315P-076, 1315P-570, 1315P-683, and 1315P-960 in Soil by LC-MS/MS

1.0 INTRODUCTION

The purpose of this study is to confirm that an analytical method, developed by one group, can be independently validated by a second group in the absence of major interaction between the two. This study is required by EPA under Guideline OCSPP 850.6100: Environmental Chemistry Methods and Associated Independent Laboratory Validation [EPA 712-C-001], Guideline OCSPP 850.7100: Data Reporting for Environmental Chemistry Methods [EPA 712-C-96-348], and Guideline OCSPP 860.1340: Residue Analytical Method [EPA 712-C-96-174], as well as satisfies OECD guidance document ENV/JM/MONO(2007)17, EC guidance documents SANCO/3029/99 REV 4(2000) and SANCO/825/00 REV 8.1(2010). Independent labs are allowed to analyze three sample sets in order to validate the method as written. A complete set of samples should consist of, at a minimum, a reagent blank, two un-spiked matrix control samples, five matrix control samples fortified at the limit of quantification (LOQ), and five matrix control samples fortified at 10X LOQ for each distinct matrix. A complete set may include more than thirteen samples depending on the number of reagents, and un-fortified and fortified control matrix samples. It may be necessary, however, to divide a complete set into two subsets for efficient handling. Each subset should contain a reagent blank, two un-fortified matrix control samples, and five matrix control samples fortified at the LOQ or 10X LOQ.

If the performance data on the first set of samples at any of the required spiking levels is unsuccessful, the independent laboratory may contact the registrant to clarify the directions given in the method. Any contact with the registrant or developers during the method validation must be documented in writing in the final report submitted by the independent laboratory. If the independent laboratory cannot generate performance data that is similar to the registrant's or developers' after the second set of spiked samples, the independent laboratory may contact the registrant to further clarify the directions given in the method. If the independent laboratory cannot generate performance data that is similar to the registrant's or developers' after the third set, the method should be failed and a report will be sent to the registrant explaining why the method failed. The registrant should then decide whether to repeat the independent laboratory validation at another laboratory, further develop the method or withdraw it. This ILV trial will be conducted under FIFRA Good Laboratory Practice (GLP) standards as specified in 40 CFR part 160. A maximum of three sample sets are used by an independent laboratory to validate the method as written. A successful ILV trial will require adequate results on at least one complete set of samples on a given matrix.

The purpose of this protocol is to perform an ILV for the analytical method used to determine the test substance in two soil/sediment types (identified in the raw data and final report). The analytical method will be validated with regards to accuracy, precision, signal response, selectivity, and limits of quantitation.

2.0 OBJECTIVE

The objective of this study is to confirm that the analytical method for Benzobicyclon and its metabolites in soil, developed by one group, can be independently validated by a second group in the absence of major interaction between the two.

3.0 JUSTIFICATION OF THE TEST SYSTEM

The method validations described in this protocol are designed to conform to EPA Guideline OCSPP 850.6100: Environmental Chemistry Methods and Associated Independent Laboratory Validation [EPA 712-C-001], Guideline OCSPP 850.7100: Data Reporting for Environmental Chemistry Methods [EPA 712-C-96-348], and Guideline OCSPP 860.1340: Residue Analytical Method [EPA 712–C–96–174].

4.0 MATERIALS

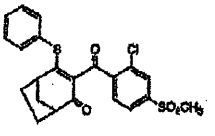
4.1 Test Substance

Upon arrival at Smithers Viscient, the test and reference substance(s) will be received by the Test Material Center. Records will be maintained in accordance with GLP requirements, and a Chain-of-Custody established. The condition of the external packaging of the test substance will be recorded and any damage noted. The packaging will be removed, the primary storage container inspected for leakage or damage, and the condition recorded. Any damage will be reported to the Sponsor and/or manufacturer.

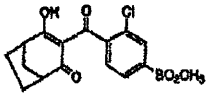
Each sample will be given a unique sample ID number and stored under the conditions specified by the Sponsor or manufacturer. The following information should be provided by the Study Sponsor, if applicable: test substance lot or batch number, test substance purity, water solubility (pH and temperature of solubility determination), vapor pressure, storage stability, methods of analysis of the test substance in water, MSDS, and safe handling procedures, and a verified expiration or reanalysis date.

4.1.1 Test Substance Information

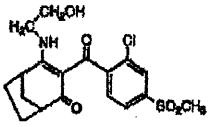
1. Name: Benzobicyclon
   Purity: 99.3%
   Batch or Lot #: 1L0108



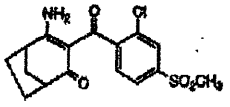
2. Name: 1315P-070
Purity: ~~99.7%~~ 98.8% updated per COA. KBB 14APR17
Batch or Lot #: 95Z25



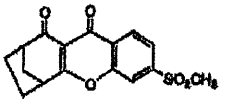
3. Name: 1315P-076
Purity: 99.5%
Batch or Lot #: TNA-10-074



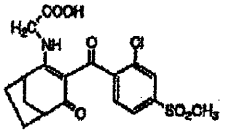
4. Name: 1315P-570
Purity: 99.8%
Batch or Lot #: TNA-9-186



5. Name: 1315P-683
Purity: 99.77%
Batch or Lot #: TM-8-198



6. Name: 1315P-960
Purity: 99.48%
Batch or Lot #: H/M-13-57-2



5.0 TEST SYSTEM IDENTIFICATION

Test solution preparation will be documented on data forms which include the amount of test substance, the volume or mass of the test solution, lot, batch or other sample designation of the test substance and date the solution was prepared. Individual sample containers will be labeled with a unique ID number.

6.0 ANALYTICAL METHOD

The analytical method used for this ILV is, "Analytical Method for the Determination of Benzobicyclon and its Metabolites 1315P-070, 1315P-076, 1315P-570, 1315P-683, and 1315P-960 in Soil by LC-MS/MS (M. Boatwright, GPL-MTH-088 Revision 1, effective January 15, 2015).

7.0 VALIDATION DESIGN

The standard curve will be comprised of at least five concentrations. The anticipated concentration range is 0.100-5.00 ppb. A smaller, larger, or shifted range may be necessary if achievable. The range will be documented in the study records and final report.

The limit of detection (LOD) will be established by evaluating the signal-to-noise (S/N) ratio from samples of known concentration and blank samples to establish the lowest level at which the analyte can be reliably detected. A S/N ratio of 3:1 is generally considered the minimum acceptable ratio for reliable detection.

7.1 Accuracy and Precision

The accuracy of the analytical method will be determined by applying the method to five samples of two soil/sediment types at the LOQ (5.0 ppb) and five samples at 10X LOQ (50.0 ppb) for each test substance. The accuracy will be reported in terms of percent recovery and the difference between the mean determined and the theoretical value. Recoveries of 70.0 to 120% of nominal are acceptable.

The precision will be calculated for the fortified samples in terms of the standard deviation (SD) and relative standard deviation (RSD or coefficient of variation (CV)) calculated for the retention

time, peak area based quantitation (i.e., µg/kg), and the observed recovery values. The retention time should have a RSD of less than or equal to 2%. The RSD of the peak area based quantitation (i.e., µg/kg) should be less than or equal to 20%. The RSD of the recovery values should be less than or equal to 20% as well.

7.2 Specificity

The specificity of the method will be determined by applying the method to two un-fortified matrix control samples for each matrix. Chromatograms will be obtained for the control samples and examined for peaks that might interfere with the quantitation of the analyte peak of interest. Peaks attributable to test substance should be sufficiently resolved from any peaks found in the samples of control matrix to enable quantification. Interferences with peak areas that are less than 50% at the limit of detection (LOD) are not considered significant.

7.3 Signal Response

The signal response of the method will be determined by preparing a calibration curve with a minimum of five standards to encompass approximately 70.0 to 120% of the test concentration.

The calibration data will be subjected to a regression analysis; a plot of the analyte concentration versus the detector response will be included in the report along with the correlation coefficient, y-intercept, and slope of the regression line. The signal response data should have an intercept close to zero and a correlation coefficient (r) not less than 0.995 (r^2 not less than 0.990). The responses of the standards shall be inserted into the regression equation, and a calculated concentration value calculated. This calculated value shall be within ±20% of the theoretical value. Deviations from these criteria will be addressed by reevaluating the calibration range, such that the calculated values meet these criteria.

8.0 CONTROL OF BIAS

Bias will be effectively controlled through techniques such as, but not limited to, preparation of replicate samples, replicate analysis, and maintenance of material balance.

9.0 RECORDS TO BE MAINTAINED

Records to be maintained will include, but will not be limited to, correspondence and other documents relating to the interpretation and evaluation of data as well as all raw data and documentation generated as a result of the study.

10.0 SAMPLE DISPOSAL

All study specimens, and/or samples collected during the study, and test materials and reference standards, etc., provided by the sponsor, client, or customer will either be returned to the originator, shipped to a third party archival facility on behalf of the study sponsor who will incur the costs of shipping and archival, or disposed of according to SMV SOPs.