US Environmental Protection Agency Office of Pesticide Programs

Office of Pesticide Programs
Microbiology Laboratory
Environmental Science Center, Ft. Meade, MD

Disinfectant Towelette Test: Testing of *Mycobacterium bovis* (BCG)

SOP Number: MB-23-05

Date Revised: 04-20-22

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Title	Disinfectant Towelette Test: Testing of Mycobacterium bovis (BCG)
Revisions Made	• The Application was changed to read "The methodology described in this SOP is used to evaluate the performance of antimicrobial towelettes against Mycobacterium bovis (BCG)." • In section 10, Cautions, added statements on use of appropriate aseptic techniques for all test procedures involving the manipulation of test organism and required adherence to the procedure for validity of test results. • In section 11, added non-sterile gloves, 70% ethanol (v/v), and flame sterilized loops to the list of apparatus. • In section 12, added direction to conduct neutralization testing prior to testing to determine if secondary subculture tubes are necessary. • In section 12.6, added non-sterile gloves sprayed with 70% ethanol (v/v) as an option for handling a test towelette. • In Attachment 1, replaced "Middlebrook 7H9" with "M7H11" as a growth medium and added the incubation temperature (36±1°C) for the M7H11. • Minor editorial changes for clarification purposes.

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Title	Disinfectant Towelette Test: Testing of *Mycobacterium bovis* (BCG)
Scope	Describes the methodology to determine tuberculocidal activity of towelette-based disinfectants labeled to treat hard non-porous surfaces against *Mycobacterium bovis* (BCG). The test is based on AOAC Method 961.02 (Germicidal Spray Products as Disinfectants), see 15.1.
Application	The methodology described in this SOP is used to evaluate the performance of antimicrobial towelettes against *Mycobacterium bovis* (BCG).

	Approval	Date
SOP Developer:	Rebecca Pines Print Name: Rebecca Pines	04/20/22
SOP Reviewer	Michele Cottrill Print Name: Michele Cottrill	04/20/22
Quality Assurance Unit	Kiran Verma Print Name: Kiran Verma	04/20/22
Branch Chief	Rebecca Pines Print Name: Rebecca Pines	04/20/22

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1. **Definitions**	1. Additional abbreviations/definitions are provided in the text. 2. Carrier Set = One "carrier set" is defined as the primary MPB tube containing the carrier and duplicate tubes of the two additional subculture media (e.g., M7H9 broth, Kirchner's medium, or TB broth) inoculated from the carrier's corresponding neutralizer tube for a total of 5 tubes per carrier. There are 10 carrier sets per disinfectant tested.
2. **Health and Safety**	Follow procedures specified in SOP MB-01, Laboratory Biosafety. The Study Director and/or lead analyst should consult the Safety Data Sheet for specific hazards associated with products.
3. **Personnel Qualifications and Training**	Refer to SOP ADM-04, OPP Microbiology Laboratory Training.
4. **Instrument Calibration**	Refer to SOPs EQ-01 (pH meters), EQ-02 (thermometers), EQ-03 (weigh balances), EQ-04 (spectrophotometers), EQ-05 (timers), and QC-19 (pipettes).
5. **Sample Handling and Storage**	Refer to SOP MB-22, Preparation and Sampling Procedures for Antimicrobial Test Substances, and SOP COC-01, Chain of Custody Procedures.
6. **Quality Control**	For quality control purposes, the required information is documented on the appropriate form(s) (see section 14).
7. **Interferences**	1. Prior to inoculation, ensure that the carriers are dry (inside Petri dishes). Moisture can interfere with the concentration and drying of the inoculum on the glass slide carrier. 2. Do not use any inoculated carrier that is wet at the conclusion of the carrier drying period.
8. **Non-conforming Data**	1. An assessment of media quality (performance) is necessary to ensure the validity of the tuberculocidal efficacy results; tests will be invalidated if any media exhibit unsatisfactory performance. The media assessment may be conducted in advance of or concurrently with efficacy testing; refer to SOP MB-10, Media and Reagents: Preparation and Quality Evaluation. 2. Sterility and/or viability controls do not yield expected results. 3. The mean log density for control carriers (*TestLD*) falls outside the specified range. Note: The prescribed minimum and maximum carrier counts also account for the addition of 5% organic soil to the inoculum. a. The mean *TestLD* must be at least 4.0 (corresponding to a geometric mean density of 1.0×10^4) and not above 6.0

	(corresponding to a geometric mean density of 1.0×10^6). 4. Management of non-conforming data will be consistent with SOP ADM-07, Non-Conformance Reports.
9. **Data Management**	Data will be archived consistent with SOP ADM-03, Records and Archives.
10. **Cautions**	1. There are time sensitive steps in this procedure including the use-period of the inoculated carriers and the test chemical. Strict adherence to the procedure is necessary for validity of test results. 2. Verify the volume of dilution blanks, neutralizer tubes, and subculture tubes in advance and adjust accordingly. 3. Use appropriate aseptic techniques for all test procedures involving the manipulation of test organism and associated test compounds.
11. **Special Apparatus and Materials**	1. Culture media. a. *Modified Proskauer-Beck medium*. Dissolve 2.5 g KH_2PO_4, 5.0 g asparagine, 0.6 g $MgSO_4\times7H_2O$, 2.5 g magnesium citrate, 20.0 mL glycerol, 0.0046 g $FeCl_3$, and 0.001 g $ZnSO_4{\cdot}7H_2O$ in 1 L H_2O. Adjust to pH 7.2-7.4 with 1 N NaOH. Filter through Whatman No. 4 (or equivalent) filter paper, place 20 mL portions in separate 25×150 mm tubes, and steam sterilize 20 min at 121°C. Use this broth for propagating test cultures grown statically (25×150 mm tubes) and for recovery of test organism from treated carriers (38×100 mm tubes). b. *Middlebrook 7H9 broth (dehydrated M7H9 medium) with 0.1% (v/v) polysorbate 80.* Dissolve 4.7 g in 900 mL H_2O containing 1 mL polysorbate 80, 2 mL glycerol, and 1.0 g Bacto agar. Heat to boiling to dissolve completely. Steam sterilize 15 min at 121°C. Cool sterile medium to 45°C, add 100 mL Middlebrook ADC Enrichment under aseptic conditions and mix thoroughly. Store prepared medium at 2-5°C. Use this broth for propagating test cultures grown with agitation. c. *Middlebrook 7H11 agar (dehydrated M7H11 medium)*. Dissolve 21 g dehydrated M7H11 agar medium in 900 mL H_2O containing 5 mL glycerol. Swirl to obtain a smooth suspension; boil if necessary to completely dissolve the powder. Steam sterilize 15 min at 121°C. Cool sterile medium to 50-55°C, add 100 mL OADC enrichment under aseptic conditions, and mix thoroughly. Distribute in 20 mL portions in sterile 25×150 mm screw-capped tubes and slant or dispense a minimum of 30 mL into sterile Petri

plates. Alternatively, pre-made M7H11 agar plates may be purchased. Use slants to maintain stock culture and plates for inoculum isolation and enumeration.

d. *Middlebrook 7H9 broth (dehydrated M7H9 medium)*. Dissolve 4.7 g in 900 mL H_2O containing 2 mL glycerol and 1.0 g Bacto agar. Heat to boiling to dissolve completely. Distribute 18 mL portions in 25×150 mm tubes. Steam sterilize 10 min at 121°C, according to manufacturer's instructions. Cool sterile medium to approximately 40-45°C then add 2 mL Middlebrook ADC Enrichment to each tube under aseptic conditions and mix thoroughly. Store prepared medium at 2-5°C. Use for recovery of test organism from treated carriers.

e. *Kirchner's medium*. Dissolve 5 g asparagine, 2.5 g sodium citrate, 0.6 g magnesium sulfate (heptahydrate), 2.5 g monopotassium phosphate, and 1.5 g dipotassium phosphate, in 900 mL H_2O containing 20 mL glycerol and 1.0 g Bacto agar. Heat to boiling to dissolve completely. Steam sterilize 15 min at 121°C. Cool sterile medium to 45°C, add 100 mL Middlebrook ADC Enrichment under aseptic conditions, and mix thoroughly. Distribute in 20 mL portions in sterile 25×150 mm tubes. Use for recovery of test organism from treated carriers.

f. *TB broth base*. Dissolve 2.0 g yeast extract, 2.0 g proteose peptone No. 3, 2.0 g casitone, 1.0 g potassium phosphate monobasic, 2.5 g sodium phosphate dibasic, 1.5 g sodium citrate, and 0.6 g magnesium sulfate (heptahydrate) in 900 mL H_2O containing 50 mL glycerol and 1.0 g Bacto agar. Heat to boiling to dissolve completely. Steam sterilize 15 min at 121°C. Cool sterile medium to 45°C, add 100 mL Dubos Medium Serum under aseptic conditions, and mix thoroughly. Distribute in 20 mL portions in sterile 25×150 mm tubes. Use for recovery of test organism from treated carriers.

g. *Middlebrook 7H10 agar.* Dissolve 19 g in 900 mL H_2O containing 5 mL glyerol. Heat to boiling to dissolve completely. Steam sterilize 15 min at 121°C. Cool sterile medium to 45°C, add 100 mL Middlebrook ADC Enrichment under aseptic conditions and mix thoroughly. Use for initiating stock cultures.

2. Test organism.

a. *Mycobacterium bovis* (BCG) (ATCC #35743). For stock culture, streak inoculate M7H11 agar slants. Incubate 15-20 days at 36±1°C. Following incubation, maintain at 2-5°C for up to 6

	weeks. 3. Reagents a. *Sterile water*. Use reagent-grade water free of substances that interfere with analytical methods. Any method of preparation of reagent-grade water is acceptable provided that the requisite quality can be met. See Standard Methods for the Examination of Water and Wastewater and SOP QC-01, Quality Assurance of Purified Water for details on reagent-grade water. b. *0.1% polysorbate 80 in saline*. Add 0.1 mL polysorbate 80 to 100 mL sterile 0.85% aqueous saline (sodium chloride) solution, filter sterilize. Used in test culture preparation and dilution of culture grown with agitation. 4. Apparatus. a. *Specialized glassware*. For neutralizer/primary subcultures, use autoclavable 38×100 mm tubes (Bellco Glass Inc., Vineland, NJ). Cap tubes with closures before sterilizing. For glassware used to prepare test chemical, refer to SOP MB-22, Disinfectant Sample Preparation. b. *Tissue grinder*. Kimble glass tissue grinder (catalog number 885300-0015), for homogenization of the statically grown culture. c. *Inoculating loop*. For culture inoculation, 1 µL sterile disposable loops (Fisher Scientific). For culture harvest, 95% platinum, 3.5% rhodium alloy, 18 or 19 gauge, 4 mm loop with 75 mm shank (Baxter Scientific Products) or equivalent or disposable loops. d. *Carriers*. Glass Slide Carriers, 25 mm×75 mm (or comparable size) borosilicate glass cover slips with number 4 thickness or Fisherbrand® Premium Frosted Microscope Slides (Fisher Scientific, catalog number 12-544-2). Refer to SOP MB-03, Screening of Stainless Steel Cylinders, Porcelain Cylinders and Glass Slide Carriers Used in Disinfectant Efficacy Testing. e. *Non-sterile or Sterile surgical gloves*. For handling the towelette. f. *70% (v/v) ethanol*. For disinfecting non-sterile gloves and external surface of towelette container/canister/flatpack. g. *Forceps*. For manipulating glass slides. h. *Flame sterilized loop.* For spreading inoculum on the surface of the carriers. Make 4 mm inner diameter single loop at end of 50-75 mm (2-3 in.) Pt or Pt alloy wire No. 23 B&S gage or 4 mm loop

	fused on 75 mm (3 in.) shaft (available from Johnson Matthey, West Chester, PA 19380, USA). Fit the other end in suitable holder. Bend loop at 30° angle with stem. i. *Micropipettes*. For performing serial dilutions. j. *Positive displacement pipette.* With corresponding sterile tips able to deliver 0.01 mL (10 µL). k. *Timer*. Any certified timer that can display time in seconds. l. *Spectrophotometer*. Calibrated; for preparing standardized test culture. m. *Semi-microcuvette with cap.* For measuring percent transmittance. n. *TB Stain Kit.* For presumptive identification of test microbe. o. *Incubated Shaker.* To provide rotation at 150 rpm and 36±1°C for cultures grown with agitation.
12. **Procedure and Analysis**	One towelette is used to wipe ten carriers/slides. The area of the towelette used for wiping is folded and rotated to expose a new surface of the towelette for each carrier. The method may be altered to accommodate various towelette/carrier combinations (e.g., more than one towelette per set of ten slides). Prior to testing, perform the neutralization assay to determine if secondary subculture tubes are necessary (refer to SOP MB-11, Neutralization Confirmation Assay for Disinfectant Products Tested against *Mycobacterium bovis* (BCG)). Use the Disinfectant Towelette Test for *M. bovis* (BCG) Processing Sheet (see section 14) for tracking testing activities.
12.1 Test Culture Preparation: Agitated Culture	Refer to SOP MB-02 for the test microbe culture transfer notation. a. Transfer a 10 µL loopful of *M. bovis* (BCG) from an M7H11 stock slant to a 25×150 mm tube containing 10 mL of Middlebrook 7H9 broth with 0.1% (v/v) polysorbate 80 (M7H9/P80), parafilm the cap to the tube, and briefly vortex. Incubate the tube at 36±1°C on a rotary shaker at 150 rpm for 5-8 days. *This represents a primary (1°) culture and is never used as a test culture.* b. After incubation, vortex the 1° tube well and transfer 1 mL to a 250 mL flask containing 50 mL of M7H9/P80. Incubate at 36±1°C on a rotary shaker at 150 rpm for 6-10 days. *This represents the secondary (2°) culture and is the test culture.* c. On the test day (following the 6-10 day incubation period), harvest

	the culture: i. Transfer the 2° culture to sterile 25×150 mm test tubes. Allow the suspension to settle for 10-15 min. ii. Remove the upper portion of each culture (e.g., upper ¾), leaving behind any debris or clumps, and transfer to a sterile flask; pool cultures in the flask and swirl to mix. iii. Dilute the pooled culture with sterile saline with 0.1% polysorbate 80 (saline/P80) to achieve 20±1% transmittance at 650 nm. Use a semi-microcuvette with cap while measuring transmittance. Blank the spectrophotometer with M7H9/P80. d. If an organic soil load is specified in the test parameters for the product test, add the appropriate amount of organic soil to the pooled test culture prior to the inoculation of carriers. Swirl to mix. e. Inoculate glass slide carriers with the standardized culture within 10 min of standardization. Briefly mix culture prior to use.
12.2 Test Culture Preparation: Static Culture (alternative culture preparation procedure)	Refer to SOP MB-02 for the test microbe culture transfer notation. a. Initiate test culture by inoculating a sufficient number of 25×150 mm tubes containing 20 mL MPB (approximately 10) from stock culture slant(s) (M7H11 agar slants) by transferring 1-2 1 µL loopfuls from the stock culture onto the surface of the broth. Record all transfers on the Organism Culture Tracking Form (culture notation = –SL, indicating a transfer from slant to liquid). b. Note: Over-inoculation of MPB may lead to reduced viability due to excessive growth after 21±2 days; the resulting carrier counts may be negatively impacted. c. Incubate the tubes 21±2 days undisturbed at 36±1°C in a slanted position to increase surface area. d. On the test day: using a transfer loop, transfer culture to a sterile glass tissue grinder, add 1.0 mL saline/P80, grind continuously for approximately 1 min to break up large clumps or aggregates of the test organism. e. Dilute the homogenized culture with 9 mL MPB broth and transfer the suspension from the tissue grinder to a sterile test tube. Harvest and homogenize culture from multiple MPB broth tubes. f. Repeat 12.2d-e as necessary to obtain enough concentrated culture.

	g. Note: Growth from multiple tubes may be harvested and combined to prepare the concentrated culture prior to standardization. i. Allow the suspension to settle for 10-15 min. ii. Remove the upper portion of each culture (e.g., upper ¾), leaving behind any debris or clumps, and transfer to a sterile flask; pool cultures in the flask and swirl to mix. iii. Dilute the pooled culture with MPB broth to achieve 20±1% T at 650 nm. Use a semi-microcuvette with cap while measuring transmittance. Blank the spectrophotometer with MPB. h. If an organic soil load is specified in the test parameters for the product test, add the appropriate amount of organic soil to the pooled test culture prior to the inoculation of carriers. Swirl to mix. i. Aliquot a sufficient volume of culture into a sterile test tube. j. Inoculate glass slide carriers with the standardized culture within 10 min of standardization. Briefly mix culture prior to use.
12.3 Carrier Inoculation	Inoculate approximately 20 carriers; 10 carriers are required for testing, 3 for control carrier counts, and 3 for the viability controls. a. Vortex-mix the inoculum periodically during the inoculation of carriers. Use a calibrated positive displacement pipette to transfer 10 µL of the culture to the sterile test carrier in the Petri dish, at one end of the slide. Do not place inoculum in the middle of the slide. Immediately spread the inoculum uniformly over one third of the carrier surface using a sterile loop. Do not allow the inoculum to contact the edge of the glass slide carriers. Cover dish immediately. b. Dry carriers in incubator at 36±1°C for 30±2 min. Record the timed carrier inoculation activities on the Disinfectant Towelette Test for *M. bovis* (BCG) Processing Sheet (see section 14). Use inoculated carriers for testing within 2 h of drying. c. After completion of all slide inoculations, thoroughly wipe the micropipette with 70% ethanol prior to removal from the BSC.
12.4 Enumeration of viable bacteria from carriers (control	a. After inoculated carriers have dried, randomly select 3 inoculated carriers for assay. Assay 1 carrier immediately prior to conducting the efficacy test and 2 carriers immediately following the test. b. Place each of the inoculated, dried carriers in a 38×100 mm tube

carrier counts)	or a sterile 50 mL polypropylene conical tube containing 20 mL of MPB broth and vortex each tube for 15 s. Record the time of vortexing on the Disinfectant Towelette Test for *M. bovis* (BCG) Processing Sheet (see section 14). c. After vortexing, make serial ten-fold dilutions in 9 mL phosphate buffered dilution water. If the serial dilutions are not made and plated immediately, keep the vortexed tubes at 2-5°C until this step can be done; however, perform dilution and plating within 2 h of vortexing. d. Briefly mix each serial dilution tube prior to plating. Plate 0.1 mL aliquots of appropriate dilutions in duplicate on M7H11 using surface spread plating. Serial dilution tubes 10^{-1} through 10^{-3} should produce plates with CFU in the countable range. Spread inoculum evenly over the surface of the agar. Plates must be dry prior to incubation. e. Incubate plates (inverted) concurrently with the efficacy test subculture tubes at 36±1°C for 17-21 days. Place plates in sterile bags to reduce dehydration during the incubation period. f. Count colonies. Record plates that have colony counts over 300 as TNTC. Record counts on the Disinfectant Towelette Test for *M. bovis* (BCG) Carrier Counts Form (see section 14). See section 13 for data analysis.
12.5 Disinfectant Sample Preparation	a. Prepare disinfectant sample per SOP MB-22, Preparation and sampling procedures for antimicrobial substances. b. Wipe the outside of the towelette packet or dispenser with 70% ethanol and allow to air dry prior to opening.
12.6 Test Procedure	a. Record timed events on the Disinfectant Towelette Test for *M. bovis* (BCG) Time Recording Sheet for Carrier Transfers (see section 14). b. Use a new pair of sterile gloves (or non-sterile gloves sprayed with 70% [v/v] ethanol) when preparing to handle each test towelette. c. Aseptically remove several towelettes before aseptically removing a towelette to initiate testing. Fold towelette in half lengthwise one to two times depending on the size. Beginning at the bottom, fold up towards the top five times. The following steps in the "procedure" section are more conveniently done with two analysts – one to manage the Petri dishes and slides, and the other to perform the wiping procedure.

	d. Remove the lid from the Petri dish and aseptically remove the inoculated slide and hold it firmly against the rim of the Petri dish. e. Wipe the slide back and forth three times lengthwise with the towelette for a total of six passes across the inoculum or as specified by the study sponsor. Wiping should be done within ±5 seconds of specified time. Place slide in Petri dish, close the lid, and allow slide to sit undisturbed for the contact time. Maintain the wiped carriers in a horizontal position. f. Repeat with four additional slides, folding the used section of the towelette in such a way as to expose a new surface for wiping each slide. g. After the fifth slide, unfold the vertical fold in the towelette and reverse the towelette so that the used surface of the towelette faces inward. Continue wiping an additional five slides, folding the towelette between each slide to expose a new surface. h. After the last slide of a set (typically 10 slides) has been wiped and the exposure time is complete, sequentially transfer each slide into the neutralizer tube within the ±5 second time limit. Drain the carriers without touching the Petri dish or filter paper. Perform transfers with flame sterilized or autoclaved forceps. Place the inoculated/wiped end of the slide into the tube. i. After the slide is deposited, recap the neutralizer tube and shake it thoroughly; transfer the carrier to the tube containing 20 mL MPB broth within 5-10 minutes. Sterilize forceps after each carrier transfer. j. Once all carriers have been transferred to the MPB broth tubes, sequentially transfer 2 mL aliquots from each neutralizer tube into duplicate tubes of 2 additional subculture media, M7H9 broth, Kirchner's medium, or TB broth, as specified. This portion of the assay is not timed, but the aliquots should be sequentially transferred to the subculture media within approximately 30±5 min. Repeat this with each tube of neutralizer. Shake each subculture tube thoroughly. Slightly loosen caps of growth media prior to incubation. k. Incubate 60 days at 36±1°C. l. Report results as + (growth) or 0 (no growth).

	m. Record results at 60 days. If the 60th day of incubation falls on a weekend or holiday, record the results on the first workday following the 60th day of incubation. i. Tubes may be monitored beginning at day 21 for evidence of typical mycobacterial growth. If multiple tubes show significant growth prior to the 60th day, confirmatory tests (e.g., acid fast staining and streak isolation) may be initiated prior to day 60. If the results of the confirmatory test are indicative of *M. bovis* (BCG), the results may be recorded at that point to expedite the reporting process. ii. Provide justification when recording results on days other than 60 in the comments section of the Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet (see section 14). n. If no growth or occasional growth (insufficient for confirmatory tests) occurs within a set of tubes after 60 days, incubate the set an additional 30 days and record the results. After 30 days, if growth occurs check using standard confirmatory procedures (e.g., acid fast staining and growth on M7H11 agar) to ensure that no contamination is present. o. Record results at 90 days. If the 90th day of incubation falls on a weekend or holiday, record the results on the first workday following the 90th day of incubation. Recording of results beyond the 90th day should be notated in the comments section on the Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet (see section 14).
12.7 Sterility and viability controls	a. Sterility controls. Place one sterile, uninoculated carrier into a tube of MPB broth. In addition, add 2 mL sterile neutralizer to 1 tube of each subculture medium and incubate for quality control purposes. Shake each tube thoroughly and incubate all tubes with the efficacy test. Report results as + (growth) or 0 (no growth) as determined by presence or absence of turbidity or presence of culture growth. Growth should not occur in any tube. Record results on the Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet (see section 14). b. Viability controls. On the day of testing, place a dried inoculated carrier into a tube of MPB broth and a tube of each subculture medium. Incubate tubes as in the efficacy test. Report results as + (growth) or 0 (no growth) as determined by presence or absence of turbidity or presence of culture growth. Growth should occur in all

	tubes. Record results on the Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet (see section 14).
12.8 Test microbe identification	a. Presumptively confirm at least one positive subculture tube for each carrier set with growth. The maximum number of tubes subjected to confirmatory tests per disinfectant tested is 10. b. If more than one subculture tube for a carrier set is positive, confirm a minimum of one tube using acid fast staining and isolation on selective media (M7H11 agar plates). c. If the MPB in the set is positive, it is the representative subculture tube used for identification. If MPB is not positive, any of the other subculture media may be used for identification. d. If growth is observed in only one carrier set, then all subculture tubes showing growth for that carrier are subject to confirmatory tests. e. Growth for acid fast staining is taken from the selected positive tubes on the day that results are read. Acid fast rods are typical for *M. bovis* (BCG). The acid fast staining results should be read promptly prior to assigning a + or 0 to the results. If acid fast rods are observed from the selected tubes then a + is assigned to the results. If no cells are observed for the acid fast stain, apply a 0 to the results. f. In addition, streak isolate growth from positive tubes on M7H11 agar and incubate for 17-21 days at 36±1ºC. g. Following the 17-21 day incubation period, evaluate the colony morphology on M7H11 agar. *M. bovis* (BCG) typically appears as colorless to buff-colored, raised, rough growth on M7H11 agar (see Attachment 1). h. If a satisfactory smear cannot be obtained directly from the tube, take the smear for acid fast staining from the 17-21 day old M7H11 agar plate that was inoculated with the growth from the tube. i. In the event that no cells were observed with acid fast staining initially but typical growth was observed on the M7H11, correct the 0 to read + on the test sheet. An entry error will be noted in the comments section of the Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet (see section 14). j. Record results on the Disinfectant Towelette Test for *M. bovis* (BCG) Microbe Confirmation Sheet (see section 14).

13. **Data Analysis/ Calculations**	Calculations will be computed using a Microsoft Excel spreadsheet (see section 14). Both electronic and hard copies of the spreadsheet will be retained. Counts from 0 through 300 and their associated dilutions will be included in the calculations.
14. **Forms and Data Sheets**	1. Attachment 1: Typical Growth Characteristics of Strains of *M. bovis* (BCG) 2. Attachment 2: Culture Initiation and Stock Culture Generation for *Mycobacterium bovis* (BCG) 3. Test Sheets. Test sheets are stored separately from the SOP under the following file names: Physical Screening of Carriers Record — MB-03_F1.docx Organism Culture Tracking Form for *Mycobacterium bovis* (BCG) — MB-07_F5.docx Test Microbe Confirmation Sheet (Quality Control) — MB-07_F6.docx Disinfectant Towelette Test for *M. bovis* (BCG) Carrier Counts Form — MB-23-05_F1.docx Disinfectant Towelette Test for *M. bovis* (BCG) Time Recording Sheet for Carrier Transfers — MB-23-05_F2.docx Disinfectant Towelette Test for *M. bovis* (BCG) Information Sheet — MB-23-05_F3.docx Disinfectant Towelette Test for *M. bovis* (BCG) Results Sheet — MB-23-05_F4.docx Test Microbe Confirmation Sheet — MB-23-05_F5.docx Carrier Count Spreadsheet MS Excel spreadsheet: Carrier Count Template_CTBDTT_v4 — MB-23-05_F6.xlsx Disinfectant Towelette Test for *M. bovis* (BCG) Processing Sheet — MB-23-05_F7.docx
15. **References**	1. Official Methods of Analysis. Revised 2013. AOAC INTERNATIONAL, Gaithersburg, MD, (Method 961.02). 2. Official Methods of Analysis. 2012. 18th Ed., AOAC INTERNATIONAL, Gaithersburg, MD, (Method 965.12 In vitro Test for Determining Tuberculocidal Activity). 3. Standard Methods for the Examination of Water and Wastewater. 23rd Ed. American Public Health Association, 1015 15th Street, NW,

	Washington, DC 4. Holt, J., Krieg, N., Sneath, P., Staley, J., and Williams, S. eds.1994. Bergey's Manual of Determinative Bacteriology, 9th Edition. Williams & Wilkins, Baltimore, MD. 5. Sneath, P., Mair, N., Sharpe, M.E., and Holt, J. eds. 1986. Bergey's Manual of Systematic Bacteriology. Volume 2. Williams & Wilkins, Baltimore, MD.

Attachment 1

Typical Growth Characteristics of strains of *M. bovis* (BCG) (see ref. 15.4 and 15.5)

	M. bovis (BCG)*
Gram stain reaction	weakly (+)
Acid Fast stain reaction	(+)
Typical Growth Characteristics on Solid Media	
M7H11	rough, raised, thick colonies with a nodular or wrinkled surface and an irregular thin margin, off-white to faint buff, or even yellow
Typical Microscopic Characteristics	
Cell dimensions	0.3-0.6 µm in diameter by 1-4 µm in length*
Cell appearance	rods, straight or slightly curved, occurring singly and in occasional threads

*After 15-20 days of incubation at 36±1°C.

Attachment 2

Culture Initiation and Stock Culture Generation for *Mycobacterium bovis* (BCG)

A1.	Culture initiation. Refer to SOP MB-02 for establishment of the organism control number.	
	a.	Initiate new stock cultures from lyophilized cultures of *Mycobacterium bovis* (BCG) from ATCC after no more than 18 stock culture transfers.
	b.	Open ampule of freeze-dried organism as indicated by manufacturer. Using a tube containing 5-6 mL of M7H9 broth, aseptically withdraw 0.5 to 1.0 mL and rehydrate the lyophilized culture. Aseptically transfer the entire rehydrated pellet back into the original tube of broth. Mix well.
	c.	Use several drops of the suspension to inoculate two Middlebrook 7H10 agar plates and streak for isolation.
	d.	Incubate the tube of rehydrated culture and the plates at 36±1°C for 28±2 days.
A2.	Culture maintenance.	
	a.	Confirm the identity of a streak isolation plates and acid fast stain (see Attachment 1 for colony morphology and acid fast staining results).
	b.	Use an M7H10 streak isolation plate to streak M7H11 agar slants (stock slants). Based on anticipated use, streak approximately 10-20 stock slants.
	c.	Incubate the new stock transfers for 15-20 days at 36±1ºC. Store at 2-5ºC.
	d.	Every 6 weeks (42 days), generate an additional 10-20 M7H11 slants. Inoculate new M7H11 slants by streaking a loopful of *M. bovis* (BCG) growth from an established tube to each of the 10-20 tubes. Perform QC of stock cultures per section A3.
	e.	Incubate the stock culture slants at 36±1ºC for 15 to 20 days. Following incubation, maintain stock cultures at 2-5ºC for up to 6 weeks.
A3.	QC of stock cultures	
	a.	Up to every 6 weeks (42 days), streak a loopful of growth for isolation from the existing M7H11 stock slant used to inoculate new agar slants on a plate of M7H11 agar. Incubate the plate for 17-21 days at 36±1ºC.
	b.	Following the incubation period, record the colony morphology as observed on the M7H11 plate. See Attachment 1 for details on cell and colony morphology and stain reactions.
	c.	Perform an acid fast stain from growth taken from the M7H11 streak isolation plate according to the manufacturer's instructions. Observe the acid fast reaction by using

brightfield microscopy at 1000X magnification (oil immersion).

d. Record observations on the Test Microbe Confirmation Sheet (Quality Control) (see section 14).