The Office of Science and Technology (OST) is providing the fish tissue results from the 2022 National Lakes Assessment (NLA). The specific chemical analyses include:

- Mercury (total)
- Polychlorinated biphenyls (PCBs) (all 209 congeners)
- Per- and polyfluoroalkyl substances (PFAS) (40 compounds)

OST also is providing a separate file for the Lipid content of each sample.

This document represents the "data dictionary" for each type of chemical contaminant analysis. The field names and descriptions for the analytical results for mercury, PCBs, and PFAS are similar for each type of chemical analysis, but results from some analyses include additional information that may not apply to all analysis types. The lipid data file has a slightly different structure that is described in a separate table in this dictionary. OST is also providing information on the fish composite samples collected during the study and used to prepare the fillet tissue samples that were analyzed. The fish sample information for each contaminant data file and the lipid data file is identical, so only one version of the dictionary for the sample information is provided after the dictionary for the results for each type of contaminant analysis.

| Data Tabs for Mercury, PCBs, and PFAS | | |
|---------------------------------------|--|--|
| Field Name | Description | |
| EPA Region | The EPA Region in which the sample was collected. | |
| State | USPS 2-letter abbreviation for the state in which the sample was collected. | |
| Lake Name | Name of the lake from which the sample was collected. | |
| Site ID | The identifier assigned by EPA to the site. The first three characters are "NLA," the next two are the site selection year (22), followed by the State abbreviation and the 5-digit site location. | |
| EPA Sample ID | Unique 6-digit number assigned by EPA. | |
| Tissue Type | An indication of the tissue used for the analysis. For the 2022 NLA, all of the samples were prepared from composited fillet tissue. | |
| Method | An identifier for the analytical method used for each chemical. For the mercury and PCB results, the identifiers refer to the following standardized EPA methods: | |
| | EPA Method 1631E, Mercury in Water by Oxidation, Purge and Trap, and Cold Vapor Atomic Fluorescence Spectrometry, EPA-821-R-02-019, August 2002. | |
| | EPA Method 1668C, Chlorinated Biphenyl Congeners in Water, Soil, Sediment, Biosolids, and Tissue by HRGC/HRMS, EPA-820-R-10-005, April 2010. | |
| | 3 rd Draft EPA Method 1633, Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue Samples by LC-MS/MS, EPA 821-D-22-003, December 2022. | |
| | Common name or abbreviation for the chemical. | |
| Analyte | For PCBs, the abbreviation "PCB" is followed by the congener number (i.e., "PCB-7"). It is not practical to completely separate all 209 PCB congeners from one another during analysis, so congeners that elute from the gas chromatograph together are listed with a forward slash between each congener, in increasing congener number order, e.g., PCB-12/PCB-13. "Total PCBs" is the name given to the sum of the results for all 209 of the congeners (which includes the coeluting congener groups) reported in the fillet sample. This value was calculated by OST, using zero for any congener result that was "not detected" at the method detection limit. | |
| | For PFAS, the chemical names are the commonly used abbreviations for the anion form of the chemical (e.g., the "ate" form). The full names of the PFAS chemicals are given in a table later in this document. | |

| Data Tabs for Mercury, PCBs, and PFAS | | |
|---------------------------------------|--|--|
| Field Name | Description | |
| CAS Number | Chemical Abstracts Service Registry Number assigned by CAS to the chemical. CAS Numbers do not exist for the groups of coeluting PCB congeners. For PFAS, this is the CAS Number of the parent acid or amide form, because the anions do not have separate CAS Numbers. | |
| Amount | Concentration of the chemical, if detected. If this field is blank, then the chemical was not detected in the fillet sample. In order to accommodate the range of concentrations in these samples, all the results are presented with the same number of decimal places for a chemical (mercury) or chemical class (PCBs and PFAS). | |
| | For mercury, the amount field is presented to 1 decimal place. For the PCBs, the amount fields are presented to 5 decimal places. For PFAS, the amount field is presented to 3 decimal places. | |
| | However, these results mercury, PCBs, and PFAS have at most 3 significant figures, regardless of the number of decimal places (for example, a PFAS value of 19.000 does not imply 5 significant figures). | |
| MDL | The nominal method detection limit for the chemical, based on the procedure in 40 CFR part 136, not adjusted for actual sample size, in the units shown in the Units column. | |
| | For mercury, MDLs are reported to 2 decimal places. For PCBs, MDLs are reported to 6 decimal places. For PFAS, MDLs are reported to 3 decimal places. | |
| QL | The nominal quantitation limit (QL) or "Minimum Level" for the chemical, based on the lowest calibration standard analyzed, not adjusted for sample size, in the units shown in the Units column. | |
| Units 1 | The weight/weight units of nanograms per gram or ng/g | |
| Units 2 | The "parts per billion" notation ppb, which is equivalent to ng/g | |
| Lab Flag | The data qualifier flag(s) applied by the laboratory. For mercury, no lab qualifier flags were applied. U = Chemical not detected H = Result is an estimate (PFAS only) J = Result between the MDL and the QL B = Chemical also present in the method blank D = Result is from a diluted analysis K = Ion abundance ratio is outside of the acceptance limits, but the chemical meets all the other identification criteria NQ = Not quantified | |
| SCC Code | Qualifiers applied by the Sample Control Center staff at GDIT during data validation. The individual SCC codes applied to the PCB and PFAS results are identified and defined in the tables of SCC codes below. | |
| Comments | A text translation of the SCC code combinations applied to each result. | |
| Sort Order | Applies only to PCBs and PFAS. A field used to sort the chemical names in a consistent order within each chemical group. | |
| | For PCBs, the values in this field range from 1 to 169. For PFAS, the values in this field range from 1 to 40. | |

By the nature of the results, the method employed, and their limited use in this study (*i.e.*, none of the mercury, PCB, or PFAS results were normalized to the lipid content of the sample), the data tab for the lipids uses an abbreviated structure, as shown below.

| Data Tab for Lipids | | |
|--------------------------|---|--|
| Field Name | Description | |
| EPA Region | The EPA Region in which the sample was collected. | |
| State | USPS 2-letter abbreviation for the state in which the sample was collected. | |
| Lake Name | Name of the lake from which the sample was collected. | |
| Site ID | The identifier assigned by EPA to the site. The first three characters are "NLA," the next two are the site selection year (22), followed by the State abbreviation and the 5-digit site location. | |
| EPA Sample ID | Unique 6-digit number assigned by EPA. | |
| Tissue Type | An indication of the tissue used for the analysis. For the 2022 NLA, all of the samples were prepared from composited fillet tissue. | |
| Species - Common Name | Generally accepted common name based on Nelson et al. (2004). | |
| Method | The lipids were determined by solvent extraction followed by evaporating the solvent, weighing the residue, and subtracting the tare weight of the evaporation vessel. Thus, the method is listed as "Gravimetric" in this file. | |
| Lipid Result (%) | The percent lipid content of the sample determined by a solvent extraction and gravimetric determination procedure. "ND" in that field indicates that no lipids were detected by the gravimetric method employed. The results are reported to 2 decimal places. | |
| Reporting Limit (%) | The laboratory's reporting limit for the gravimetric procedure. | |
| Lab Flag | U = Lipids not detected | |
| Comments | Comments applied by the Sample Control Center staff at GDIT during data validation, limited to instances where the lipids were not detectable. | |

| Sample Information Tabs | | |
|---|---|--|
| Field Name | Description | |
| EPA Region | The EPA Region in which the sample was collected. | |
| State | USPS 2-letter abbreviation for the state in which the sample was collected. | |
| Site ID | The identifier assigned by EPA to the site. The first three characters are "NLA," the n | |
| Lake Name | Name of the lake from which the sample was collected. | |
| Latitude | Latitude, in decimal format, to 5 decimal places. | |
| Longitude | Longitude, in decimal format, to 5 decimal places. | |
| Sample Collection Date | Actual sampling date, in MM/DD/YYYY format. | |
| EPA Sample ID | Unique 6-digit number assigned by EPA. | |
| Specimen ID | The 6-digit EPA Sample ID, followed by a decimal point and a value between 1 and 10. The decimal portion identifies the number assigned to the individual fish specimen in the composite sample. | |
| Spec Sort | A specimen sorting field designed to account for the fact that samples with more than 9 specimens do not sort properly (i.e., XX.10 sorts before XX.2). | |
| Family | Latin name of the Family based on Nelson et al. (2004). | |
| Species - Scientific Name | Latin name (Genus and species) based on Nelson <i>et al.</i> (2004), <i>Common and Scientific</i> Names of Fishes from the United States, Canada, and Mexico, Sixth Edition. | |
| Species - Common Name | Generally accepted common name based on Nelson et al. (2004). | |
| Total Length (mm) | Length of each individual specimen in millimeters (mm). | |
| Included in Fillet Composite? | This field indicates if the specimen was included in the tissue sample for analysis or not. | |
| Predator or Bottom Dweller | All of the samples in the 2022 NLA were P = Predator species | |
| Composite Classification | Routine vs. Non-routine composite, based on the fish composite sample criteria specified in the human health fish sampling procedures. | |
| Deviation | For non-routine composites, the nature of the deviation from the criteria (e.g., number of fish, fish length, or both). | |
| Fillet Sample Preparation Instructions | Instructions from EPA/OW/OST to the sample preparation laboratory regarding which specimens to include in the fillet composite sample for analysis, based on specimen length, species, etc. | |

| Individual SCC Codes Applied to the Results | | | | |
|---|--|---|--|--|
| SCC Code | Comments | Implication | | |
| B, RMAX | Blank Contamination, Result is a Maximum Value | Blank contamination was observed and the target chemical was reported in the sample at a concentration between 5 and 10 times higher than the blank value. The result was considered to be of acceptable quality, but data users are cautioned that it may be a maximum value due to possible influence of contamination. | | |
| B, RNAF | Blank Contamination, Result is Not Affected | Blank contamination was present but was not considered to adversely impact the sample result. The presence of the chemical in the blank is not considered to adversely affect the data in cases where the sample results are more than 10 times the associated blank results or where the chemical is not detected in associated samples. | | |
| B, RNON | Blank Contamination, Result Reported as a Non-detect | When the sample result is less than five times the blank result, there are no means by which to ascertain whether or not the presence of the chemical may be attributed to contamination. Therefore, the result is reported in the database as a non-detect at the MDL, adjusted for sample size and dilution. | | |
| HIAR, J | High Ion Abundance Ratio, Estimated | Each chemical is identified and quantified based on the instrumental response for two specific ions and the ratio of those two ions was above the upper acceptance limit, suggesting a potential interference that may affect the sample result. Therefore, the result also is flagged as an estimated value. | | |
| HLBL, J | High Labeled Compound Recovery, Estimated | The labeled analog of the target chemical was recovered above acceptance criteria, suggesting the possible presence of matrix interferences. Isolated instances of high recovery are not uncommon, and patterns across multiple samples are more of a concern. | | |
| HLBL, RNAF | High Labeled Compound Recovery, Result is Not Affected | The labeled analog of the target chemical was recovered above acceptance criteria, suggesting the possible presence of matrix interferences. Isolated instances of high recovery are not uncommon, and patterns across multiple samples are more of a concern. If the chemical was not detected in a field sample, there is no concern and the RNAF is added to the HLBL flag. | | |
| HLCS, J | High Lab Control Sample Recovery, Estimated | The recovery in the LCS was high. Detected analytes also are considered estimated values. | | |
| HLCS, RNAF | High Lab Control Sample Recovery, Result is Not Affected | The recovery in the LCS was high, but the chemical was not detected in the associated fillet tissue sample, so there was no high bias concern and the RNAF flag was applied. | | |
| HRPD, J | High RPD, Estimated | The relative percent difference (RPD) between the results in the parent sample and the laboratory duplicate is above the acceptance limit. This may be due to inhomogeneity in the bulk sample or analytical variability. When high RPD was observed for a chemical, all the detected results for that chemical in any of the samples in the batch with the duplicate sample were qualified as estimated values. | | |
| HVER, RNAF | High CALVER, Result is Not Affected | The results for the calibration verification associated with the chemical were above the acceptance limit, suggesting a possible high bias. If the chemical was not detected in a field sample, there is no concern and the RNAF is added to the HVER flag | | |
| J | Estimated | When applied alone, this code indicates that the result is at or above the MDL, but below the QL. This flag also may be applied in conjunction with other flags to indicate the potential for greater uncertainty. | | |

| Individual SCC Codes Applied to the Results | | | | |
|---|--|--|--|--|
| SCC Code | Comments | Implication | | |
| LIAR, J | Low Ion Abundance Ratio, Estimated | Each analyte is identified and quantified based on the instrumental response for two specific ions and the ratio of those two ions was below the lower acceptance limit, suggesting a potential interference that may lower the sample result. Therefore, the result also is flagged as an estimated value. | | |
| LLBL | Low Labeled Compound Recovery | The labeled analog of the target chemical was recovered below acceptance criteria, suggesting the possible presence of matrix interferences or incomplete recovery of both the labeled compound and target chemical during the extract cleanup processes used in the analytical procedure. The use of isotope dilution quantitation automatically corrects the results for the target chemical, even when the labeled compound recovery is below expectations. This flag is applied when the chemical associated with the labeled analog is not detected in the sample. | | |
| LLBL, J | Low Labeled Compound Recovery, Estimated | The labeled analog of the target chemical was recovered below acceptance criteria, suggesting the possible presence of matrix interferences or incomplete recovery of both the labeled compound and target chemical during the extract cleanup processes used in the analytical procedure. The use of isotope dilution quantitation automatically corrects the results for the target chemical, even when the labeled compound recovery is below expectations. When the chemical associated with the labeled analog is detected in the sample, the result is also flagged as an estimated value. | | |
| LLCS | Low LCS result | The lab control sample (LCS) was a clean reference matrix. If recovery in the LCS was low, there may be a low bias for that chemical. When low LCS recovery was observed for a chemical, the results for that chemical were qualified in all of the samples in the batch with the LCS. | | |
| LLCS, J | Low LCS result, Estimated | The lab control sample (LCS) was a clean reference matrix. If recovery in the LCS was low, there may be a low bias for that chemical. When low LCS recovery was observed for a chemical, the results for that chemical were qualified in all of the samples in the batch with the LCS. | | |
| LVER | Low CALVER | The results for the calibration verification associated with the chemical were below the acceptance limit, suggesting a possible low bias. | | |
| NQ | Not Quantified | The chemical could not be quantified by isotope dilution and was reported as a non-detect at the MDL. | | |
| PIO, J | Peak Interference Observed, Estimated | An interference was observed in the peak for the analyte and therefore the result is flagged as an estimated value. | | |

Note: Commas are used to separate related parts of a single code (e.g., "B, RNAF is considered one code), while semicolons are used to separate different codes (e.g., "B, RNAF; J" is the combination of two codes).

| PFAS Abbreviation | PFAS Name |
|----------------------|---|
| Perfluoroalkyl carb | |
| PFBA | Perfluorobutanoic acid |
| PFPeA | Perfluoropentanoic acid |
| PFHxA | Perfluorohexanoic acid |
| PFHpA | Perfluoroheptanoic acid |
| PFOA | Perfluorooctanoic acid |
| PFNA | Perfluorononanoic acid |
| PFDA | Perfluorodecanoic acid |
| PFUnA | Perfluoroundecanoic acid |
| PFDoA | Perfluorododecanoic acid |
| PFTrDA | Perfluorotridecanoic acid |
| PFTeDA | Perfluorotetradecanoic acid |
| Perfluoroalkyl sulfo | onic acids |
| PFBS | Perfluorobutanesulfonic acid |
| PFPeS | Perfluoropentansulfonic acid |
| PFHxS | Perfluorohexanesulfonic acid |
| PFHpS | Perfluoroheptanesulfonic acid |
| PFOS | Perfluorooctanesulfonic acid |
| PFNS | Perfluorononanesulfonic acid |
| PFDS | Perfluorodecanesulfonic acid |
| PFDoS | Perfluorododecanesulfonic acid |
| Fluorotelomer sulfo | nic acids |
| 4:2FTS | 1 <i>H</i> ,1 <i>H</i> ,2 <i>H</i> ,2 <i>H</i> -Perfluorohexane sulfonic acid |
| 6:2FTS | 1H,1H,2H,2H-Perfluorooctane sulfonic acid |
| 8:2FTS | 1 <i>H</i> ,1 <i>H</i> ,2 <i>H</i> ,2 <i>H</i> -Perfluorodecane sulfonic acid |
| Perfluorooctane sul | fonamides |
| PFOSA | Perfluorooctanesulfonamide |
| NMeFOSA | N-methyl perfluorooctanesulfonamide |
| NEtFOSA | N-ethyl perfluorooctanesulfonamide |
| Perfluorooctane sul | fonamidoacetic acids |
| NMeFOSAA | N-methyl perfluorooctanesulfonamidoacetic acid |
| NEtFOSAA | N-ethyl perfluorooctanesulfonamidoacetic acid |
| | Perfluorooctane sulfonamide ethanols |
| NMeFOSE | N-methyl perfluorooctanesulfonamidoethanol |
| NEtFOSE | N-ethyl perfluorooctanesulfonamidoethanol |
| Per- and Polyfluoro | ether carboxylic acids |
| HFPO-DA | Hexafluoropropylene oxide dimer acid |
| ADONA | 4,8-Dioxa-3 <i>H</i> -perfluorononanoic acid |
| PFMPA | Perfluoro-3-methoxypropanoic acid |
| PFMBA | Perfluoro-4-methoxybutanoic acid |
| NFDHA | Nonafluoro-3,6-dioxaheptanoic acid |
| Ether sulfonic acids | |
| 9Cl-PF3ONS | 9-Chlorohexadecafluoro-3-oxanonane-1-sulfonic acid |
| 11Cl-PF3OUdS | 11-Chloroeicosafluoro-3-oxaundecane-1-sulfonic acid |
| PFEESA | Perfluoro(2-ethoxyethane)sulfonic acid |
| Fluorotelomer carb | |
| 3:3FTCA | 3-Perfluoropropyl propanoic acid |
| 5:3FTCA | 2 <i>H</i> ,2 <i>H</i> ,3 <i>H</i> ,3 <i>H</i> -Perfluorooctanoic acid |
| 7:3FTCA | 3-Perfluoroheptyl propanoic acid |
| | |